



Plants and fungi metabolites as novel autophagy inducers and senescence inhibitors

Rivka Ofir^{1,2*} 

¹Dead Sea and Arava Science Center (DSASC), Central Arava Branch, Arava 8681500, Israel

²The Regenerative Medicine & Stem Cell (RMSC) Research Center, Ben Gurion University of the Negev (BGU), Beer Sheva 84105, Israel

***Correspondence:** Rivka Ofir, Dead Sea and Arava Science Center (DSASC), Central Arava Branch, Arava 8681500, Israel.

rivir@bgu.ac.il

Academic Editor: Juergen Reichardt, James Cook University, Australia

Received: April 9, 2024 **Accepted:** May 23, 2024 **Published:** July 1, 2024

Cite this article: Ofir R. Plants and fungi metabolites as novel autophagy inducers and senescence inhibitors. *Explor Drug Sci.* 2024;2:361–8. <https://doi.org/10.37349/eds.2024.00051>

Abstract

Premature aging can be partially explained by inefficient autophagy (the process of cellular self-digestion that recycles intracellular components) and premature senescence (cease of cellular division without cell death activation). Autophagy and senescence are among the basic biochemical pathways in plants and fungi suggesting that some of their metabolites have the potential to act as autophagy inducers (AI) and senescence inhibitors (SI) and to inhibit inflammation and human aging. Several compounds have already been identified: trehalose and resveratrol are natural compounds that act as AI; flavonoids found in fruit and vegetables (curcumin, quercetin, and fisetin) are among the first SI discovered so far. New AI/SI can be identified using various approaches like hypothesis-driven approach for screening receptor agonists using an in-silico library of thousands of natural compounds; cheminformatics studies of phytochemicals using docking and molecular dynamics simulation, structure similarities/mimicry in vitro, “blind” high throughput screening (HTS) of libraries of natural metabolites against relevant models, and more. This article aims to promote the use of plant and fungi novel resources to identify bioactive molecules relevant for healthy aging based on the knowledge that plants and fungi use autophagy and senescence mechanisms for their own survival and homeostasis. As autophagy and senescence are interconnected, how drugs targeting autophagy, senescence, or both could contribute to healthy aging in humans will be speculated.

Keywords

Autophagy, senescence, inflammation, aging, fungi, plant, metabolites, senescence-associated secretory phenotype (SASP)

Autophagy and senescence as targets for inhibiting inflammation and aging

Autophagy is responsible for cellular self-digestion that recycles intracellular components and for trafficking events that activate innate and adaptive immunity as well as autoinflammatory diseases [1].

© The Author(s) 2024. This is an Open Access article licensed under a Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, sharing, adaptation, distribution and reproduction in any medium or format, for any purpose, even commercially, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.



Senescence is the cease of cellular division without cell death activation, and immune-senescence is a series of age-related changes that affect the immune system [2], including a decline in coping with proinflammatory status [3]. Normally, functioning autophagy protects against neurodegeneration associated with intracytoplasmic aggregate-prone protein accumulation, in addition to its other roles, such as neuronal stem cell differentiation [4]. Neurodegenerative disorders share common pathogenic mechanisms, including the impairment of autophagic flux, which prevents the removal of neurotoxic misfolded proteins; effective disease-modifying strategies seek novel molecules exhibiting pro-autophagic potential [5].

Senescent cells are a major contributor to age-dependent cardiovascular tissue dysfunction and the integration of transcriptomes of senescent cell models representing multi-tissue patient samples has revealed that reduced collagen type VI alpha 3 chain (*COL6A3*) expression is one of the triggers of senescence [6]. Senescent cells represent a pharmacologic target for alleviating geriatric decline and chronic diseases [7]. Senolytic drugs like dasatinib, quercetin, fisetin, and navitoclax, were discovered using a hypothesis-driven approach; early pilot trials of senolytics suggest they decrease senescent cells, reduce inflammation, and alleviate frailty in humans [8]. Increased post mitotic senescence in aged human neurons is a pathological feature of Alzheimer's disease; senescent neurons gain an inflammatory senescence-associated secretory phenotype (SASP) and can be eliminated with senotherapeutics [5]. Microbiota sensing—free fatty acid receptor 2 signaling ameliorates amyloid- β induced neurotoxicity—can be activated by modulating proteolysis-senescence axis [9].

It has been shown that plants and fungal metabolites possess antiaging properties, and compounds isolated from plants and fungi modulate the cellular and physiological pathways that prolong lifespan and prevent age-related diseases in model organisms [10]. These compounds act through cellular processes such as autophagy and senescence and as such delay aging and prevent chronic diseases [11]. When autophagy is impaired, waste derived from tissue damage leads to organ deterioration. Thus, autophagy plays a critical role in antiaging processes and mTOR plays an important role in inhibiting autophagy. A chemo-informatics study of phytochemicals, using docking and molecular dynamics simulation, identified, among other compounds, the cyclo-trijuglone of *Juglans regia* L. as a potential ATP-competitive inhibitor of mTOR [12]. Senolytic compounds that selectively clear senescent cells, such as dasatinib, quercetin, fisetin and navitoclax, were discovered using a hypothesis-driven approach [8]; flavonoids quercetin and fisetin are found in fruits and vegetables [13] and have the potential to reduce the factors secreted by senescent cells (SASP) that lead to chronic inflammation and deterioration of healthy organs [14]. Recent in silico analysis of metabolites secreted by senescent cells may serve as tools to identify senescence inhibitors (SI) based on the mechanism of action [15].

Autophagy and senescence are interconnected

Autophagy and cellular senescence serve as stress responses to mammalian cells but the interconnection between these pathways is complex: autophagy sometimes suppress cellular senescence by removing damaged macromolecules or organelles, and in different scenarios, autophagy leads to cellular senescence and synthesis of SASPs [16]. Although autophagy and senescence interconnection may influence very different processes such as stem cells [17], aging, and cancer [18, 19], autophagy activators could be exploited to prevent the induction of senescence and drugs targeting the process of autophagy can indirectly contribute to blocking the process of senescence [18]. Autophagy and senescence converge in inducing triggers and signaling pathways such as the AMPK signaling pathway [20]. And autophagic degradation of the inhibitory p53 isoform $\Delta 133p53\alpha$ acts as a regulatory mechanism for p53-mediated senescence [21].

Autophagy regulates senescence and pathogen-induced cell death in plants [22]. This basic knowledge suggests that together, autophagy inducers (AI) and SI can contribute to healthy aging in humans through various cellular mechanisms.

Plants and fungi as resources for AI

Fungi (phytopathogenic or mycorrhizal) that interact with plants depend on autophagy as a mechanism that is responsible for recycling cell components, for the interaction of fungus-plant [23], and for affecting the pathogenicity potential of plant pathogens [24]. Several examples from the literature will be discussed below.

Plants and autophagy

Autophagy, a highly conserved self-degradation mechanism, involves the encapsulation of harmful intracellular content by double-membrane autophagic vacuoles for degradation in all parts of the plant, including roots, leaves, pollen, and more [25]. Plants must cope with diverse environmental stresses such as starvation, oxidative stress, drought stress, and invasion by phytopathogens; autophagy plays a critical role during plant differentiation, development, and aging processes [22]. The active ingredient of traditional Persian medicine, cyclo-triuglone of *Juglans regia* L., regulates autophagy through the mTOR pathway [12]. Plant natural compounds such as curcumin, resveratrol, paclitaxel, oridonin, quercetin, and plant lectin regulate core autophagic pathways involved in Ras-Raf signaling, Beclin-1 interactome, BCR-ABL, PI3KCI/Akt/mTOR, FOXO1 signaling, and p53 [26]. The process of autophagy in plant cells at various stages of development is controlled by intracellular signaling pathways TOR kinase activity, hormone signaling, ROS levels, and changes in environmental conditions [27].

Fungi and autophagy

The growth of filamentous fungus *Aspergillus niger* in carbon-starved cultures activates autophagy genes that, probably, protect these fungi from cell death in addition to promoting nutrient recycling [28]. It has been shown that the autophagy of several fungi involves endoproteases and contributes to their pathogenicity [23]. Pathways of autophagy processes play important roles in filamentous fungal pathogenicity [24] and regulate fungal virulence and sexual reproduction in *Cryptococcus neoformans* [29] and in both the development and infection mechanisms of *Phytophthora sojae* [30].

Plants and fungi as resources for SI

Dasatinib, quercetin, and fisetin were identified as the 1st senolytic drugs derived from plants and fungi; they activate apoptosis of senescent cells and as such extend lifespan using animal models [31]. They may be effective in delaying human aging and treating chronic diseases.

Plants and senescence

Leaf senescence is accompanied by changes in physiological metabolism; regulation of leaf senescence improves resistance to biotic and abiotic stresses and delay in leaf senescence of horticultural plants improves their yields [32]. Senescence has a role in plant pathogenesis and defense: pathogens often delay senescence to keep host cells alive, and resistance is achieved by senescence-like processes in the host that involve gene transcription and biosynthesis pathways [33]. Extending the shelf-life of fresh produce and cut flowers relies on delaying cell death by lowering storage temperatures and modifying the environment to slow down metabolism and reduce the senescence and cell death-promoting effects of ethylene [34]. Screening of plants with inhibitory activity on cellular senescence showed that fruit of *Physalis angulata* L. and the aerial part of *Synurus deltoides* (Aiton) Nakai inhibited cell-senescence on HUVEC cell model, and water extracted from the root of *Polygonatum odoratum* var. *pluriflorum* for *variegatum* Y. N. inhibited cell-senescence in human dermal fibroblast (HDF) models. *Isatis tinctoria* L. leaf extract inhibits replicative senescence in dermal fibroblasts by regulating mTOR-NF- κ B-SASP signaling [35]. Manipulation of plant senescence to improve biotic stress resistance showed that even the application of mycorrhiza can inhibit the senescence process of plants and improve their tolerance to stresses [36]. Interestingly, senescence in plants is not merely a deterioration process leading to death but rather a unique developmental state resembling dedifferentiation [37]. Several epigenetic mechanisms that control plant senescence lead to crop improvement [38]. Postharvest research challenges various materials (such as nitric oxide) for

controlling the quality of horticultural products by inhibiting senescence; interestingly, among others, hydrogen peroxide (H₂O₂) and calcium ions (Ca²⁺) are involved [39]. Peroxidase and phenylalanine ammonia lyase are the acting players relevant to inducing senescence in plant-fungus interactions; the process is accompanied by raising the concentration of flavonoids and phenolic compounds [40].

Fungi and senescence

Mushroom extracts inhibit ultraviolet B-induced cellular senescence in human keratinocytes through augmenting sirtuin-1 (*SIRT-1*) expression [41]. Senescence has an impact on the growth of fungal colonies due to dysfunctional oxidative phosphorylation [42]. Papilla formation and hypersensitive reactions, serve as defense mechanisms against infection attempts by *Mycosphaerella* spp. (*M. graminicola*), frequently occurred in plant leaves, leading to plant senescence [43].

The publications cited here and many more suggest that plants and fungi produce metabolites that regulate autophagy and senescence and among these metabolites, there are potential AI and SI.

“The wisdom of the desert”—desert plants as novel AI and SI

Desert plants have adapted to stressful environments by synthesizing secondary metabolites and accumulating ions as osmoticum (a substance that acts to supplement osmotic pressure in a cell). Desert environments are one of the harshest places on earth due to low precipitation, limited soil nutrients, and high irradiation. The predictive metabolomics of multiple Atacama plant species unveils a core set of generic metabolites for extreme climate resilience [44]. The mechanisms to survive in harsh conditions suggest that these plants have generated unique metabolites, termed here by us “the wisdom of the desert” [45]. Studies showed variations in flavonoid metabolites along an altitudinal gradient in the desert medicinal plant *Agriophyllum squarrosum* [46]. Phytochemical analysis of secondary metabolites (alkaloids, terpenoids, tannins, saponins, flavonoids, and phenolics) in 26 plants from the desert of Egypt showed that flavonoids, phenolics, and tannins were present in all the examined species while saponin and terpenoid compounds were detected only in fifteen species. Such a resource of natural metabolites of plants, used traditionally for treatment, may be considered a new, biologically active source of medicinal compounds [47]. Among them, one can be expected to find AI and SI.

The harsh conditions of the desert also influence the biosynthesis of metabolites in fungi and microbes. Bioactive secondary metabolites from endophytic strains of *Neocamarosporium betae*, *Chaetomium globosum* (Chaetomiaceae), and *Rhinocladiella similis* collected from desert plants could be a new resource for bioactive natural products [48–50].

Filamentous cyanobacteria use unique extracellular polysaccharide-based biosynthetic pathways to survive in the desert. In addition to the extracellular polysaccharide, chaperones (to maintain protein integrity), oxidative stress protection system, synthesis of compatible solutes and ion channels, and upregulation of DNA repair mechanism are examples of the strategies cyanobacteria use for coping with desiccation/rehydration cycles in the desert [51]. These metabolites that facilitate the adaptation to extremely arid environment may contain potential AI and SI. Analysis of Sonoran desert fungi (*Aspergillus* strains) occurring in the rhizosphere of *Ambrosia ambrosoides* and in the rhizosphere of *Anicasanthus thurberi*, identified unusual new secondary metabolites terrequinone A, terrefuranone and 4R,5S-dihydroxy-3-methoxy-5-methylcyclohex-2-enone, 6-methoxy-5(6)-dihydropenicillic acid, respectively, with medicinal properties like selective toxicity against cancer cells (and not against healthy cells) [52]. These metabolites that enable growth in harsh arid environments may contain potential AI and SI.

Conclusions

Novelty: Many novel AI and SI are waiting to be discovered in plants, fungi, and microbes. As aging is characterized by systemic chronic inflammation, which is accompanied by impaired autophagy and by cellular senescence (including SASP), elimination of inflammation could be a potential healthy aging strategy. [Table 1](#) summarizes the suggested mechanism of action of AI and SI discussed in this Perspective.

Table 1. Summary of autophagy inducers (AI) and senescence inhibitors (SI) along with their potential target mechanisms

Compounds		Target/mechanism	References
AI	Resveratrol	Sirtuin-1	[53]
	Quercetin	Ras-Raf signaling, Beclin-1 interactome, BCR-ABL, PI3KCI/Akt/mTOR, FOXO1 signaling, and p53	[26]
	Plant lectin	Ras-Raf signaling, Beclin-1 interactome, BCR-ABL, PI3KCI/Akt/mTOR, FOXO1 signaling, and p53	[26]
	Trehalose	Antioxidant and more	[54]
	Cyclo-triuglone of <i>Juglans regia</i> L.	Inhibitor of mTOR	[12]
	Resveratrol	Ras-Raf signaling, Beclin-1 interactome, BCR-ABL, PI3KCI/Akt/mTOR, FOXO1 signaling, and p53	[26]
	Paclitaxel	Ras-Raf signaling, Beclin-1 interactome, BCR-ABL, PI3KCI/Akt/mTOR, FOXO1 signaling, and p53	[26]
	Oridonin	Ras-Raf signaling, Beclin-1 interactome, BCR-ABL, PI3KCI/Akt/mTOR, FOXO1 signaling, and p53	[26]
SI	Curcumin	Anti-inflammatory, immune-regulatory, anti-oxidative, and lipid-modifying properties	[55]
	Quercetin	Selectively clear senescent cells; reduce senescence-associated secretory phenotype (SASP); Ras-Raf signaling, Beclin-1 interactome, BCR-ABL, PI3KCI/Akt/mTOR, FOXO1 signaling, and p53	[26]
	Fisetin	Selectively clear senescent cells; reduce SASP	[8, 14]
	Dasatinib	Selectively clear senescent cells	[8]
	Navitoclax	Selectively clear senescent cells	[8]

Challenges (before moving to clinical trials in humans): i. Suitable in vitro and in vivo models are needed for screening the novel agents for toxicity/safety dosage, mode of application, efficacy, and selectivity. ii. Understanding of the underlying mechanisms linking autophagy, senescence, inflammation, and aging will enable optimization of therapeutic strategies.

Abbreviations

AI: autophagy inducers

SASP: senescence-associated secretory phenotype

SI: senescence inhibitors

Declarations

Author contributions

RO: Writing—original draft, Writing—review & editing.

Conflicts of interest

The author declares that there are no conflicts of interest.

Ethical approval

Not applicable.

Consent to participate

Not applicable.

Consent to publication

Not applicable.

Availability of data and materials

Not applicable.

Funding

Rivka Ofir received support from the Israeli Ministry of Innovation, Science and Technology [alona23568]. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Copyright

© The Author(s) 2024.

References

1. Zhou XJ, Zhang H. Autophagy in immunity: implications in etiology of autoimmune/autoinflammatory diseases. *Autophagy*. 2012;8:1286–99. [DOI] [PubMed] [PMC]
2. Nikolich-Zugich J. The twilight of immunity: emerging concepts in aging of the immune system. *Nat Immunol*. 2018;19:10–9. Erratum in: *Nat Immunol*. 2018;19:1146. [DOI] [PubMed]
3. Liu Z, Liang Q, Ren Y, Guo C, Ge X, Wang L, et al. Immunosenescence: molecular mechanisms and diseases. *Signal Transduct Target Ther*. 2023;8:200. [DOI] [PubMed] [PMC]
4. Menzies FM, Fleming A, Caricasole A, Bento CF, Andrews SP, Ashkenazi A, et al. Autophagy and Neurodegeneration: Pathogenic Mechanisms and Therapeutic Opportunities. *Neuron*. 2017;93:1015–34. [DOI] [PubMed]
5. Thellung S, Corsaro A, Nizzari M, Barbieri F, Florio T. Autophagy Activator Drugs: A New Opportunity in Neuroprotection from Misfolded Protein Toxicity. *Int J Mol Sci*. 2019;20:901. [DOI] [PubMed] [PMC]
6. Savić R, Yang J, Koplev S, An MC, Patel PL, O'Brien RN, et al. Integration of transcriptomes of senescent cell models with multi-tissue patient samples reveals reduced COL6A3 as an inducer of senescence. *Cell Rep*. 2023;42:113371. [DOI] [PubMed] [PMC]
7. Gasek NS, Kuchel GA, Kirkland JL, Xu M. Strategies for Targeting Senescent Cells in Human Disease. *Nat Aging*. 2021;1:870–9. [DOI] [PubMed] [PMC]
8. Kirkland JL, Tchkonja T. Senolytic drugs: from discovery to translation. *J Intern Med*. 2020;288:518–36. [DOI] [PubMed] [PMC]
9. Razazan A, Karunakar P, Mishra SP, Sharma S, Miller B, Jain S, et al. Activation of Microbiota Sensing – Free Fatty Acid Receptor 2 Signaling Ameliorates Amyloid- β Induced Neurotoxicity by Modulating Proteolysis-Senescence Axis. *Front Aging Neurosci*. 2021;13:735933. [DOI] [PubMed] [PMC]
10. Ullah H, Khan H, Prieto Lage MA, Daglia M. Editorial: Herbal medical products and natural products targeting aging and age-related disorders--ethnopharmacological perspectives. *Front Pharmacol*. 2024;15:1368389. [DOI] [PubMed] [PMC]
11. Martel J, Ojcius DM, Ko YF, Chang CJ, Young JD. Antiaging effects of bioactive molecules isolated from plants and fungi. *Med Res Rev*. 2019;39:1515–52. [DOI] [PubMed]
12. Mosaddeghi P, Eslami M, Farahmandnejad M, Akhavein M, Ranjbarfarrokhi R, Khorraminejad-Shirazi M, et al. A systems pharmacology approach to identify the autophagy-inducing effects of Traditional Persian medicinal plants. *Sci Rep*. 2021;11:336. Erratum in: *Sci Rep*. 2021;11:12481. [DOI] [PubMed] [PMC]
13. Kashyap D, Garg VK, Tuli HS, Yerer MB, Sak K, Sharma AK, et al. Fisetin and Quercetin: Promising Flavonoids with Chemopreventive Potential. *Biomolecules*. 2019;9:174. [DOI] [PubMed] [PMC]
14. Li X, Li C, Zhang W, Wang Y, Qian P, Huang H. Inflammation and aging: signaling pathways and intervention therapies. *Signal Transduct Target Ther*. 2023;8:239. [DOI] [PubMed] [PMC]
15. Oguma Y, Alessio N, Aprile D, Dezawa M, Peluso G, Di Bernardo G, et al. Meta-analysis of senescent cell secretomes to identify common and specific features of the different senescent phenotypes: a tool for developing new senotherapeutics. *Cell Commun Signal*. 2023;21:262. [DOI] [PubMed] [PMC]
16. Kwon Y, Kim JW, Jeoung JA, Kim MS, Kang C. Autophagy Is Pro-Senescence When Seen in Close-Up, but Anti-Senescence in Long-Shot. *Mol Cells*. 2017;40:607–12. [DOI] [PubMed] [PMC]

17. Capasso S, Alessio N, Squillaro T, Di Bernardo G, Melone MA, Cipollaro M, et al. Changes in autophagy, proteasome activity and metabolism to determine a specific signature for acute and chronic senescent mesenchymal stromal cells. *Oncotarget*. 2015;6:39457–68. [DOI] [PubMed] [PMC]
18. Russo M, Bono E, Ghigo A. The Interplay Between Autophagy and Senescence in Anthracycline Cardiotoxicity. *Curr Heart Fail Rep*. 2021;18:180–90. [DOI] [PubMed] [PMC]
19. Cassidy LD, Narita M. Autophagy at the intersection of aging, senescence, and cancer. *Mol Oncol*. 2022;16:3259–75. [DOI] [PubMed] [PMC]
20. Hela F, Aguayo-Mazzucato C. Interaction between Autophagy and Senescence in Pancreatic Beta Cells. *Biology (Basel)*. 2023;12:1205. [DOI] [PubMed] [PMC]
21. Horikawa I, Fujita K, Jenkins LM, Hiyoshi Y, Mondal AM, Vojtesek B, et al. Autophagic degradation of the inhibitory p53 isoform $\Delta 133p53\alpha$ as a regulatory mechanism for p53-mediated senescence. *Nat Commun*. 2014;5:4706. [DOI] [PubMed] [PMC]
22. Yoshimoto K, Takano Y, Sakai Y. Autophagy in plants and phytopathogens. *FEBS Lett*. 2010;584:1350–8. [DOI] [PubMed]
23. Juárez-Montiel M, Clark-Flores D, Tesillo-Moreno P, de la Vega-Camarillo E, Andrade-Pavón D, Hernández-García JA, et al. Vacuolar proteases and autophagy in phytopathogenic fungi: A review. *Front Fungal Biol*. 2022;3:948477. [DOI] [PubMed] [PMC]
24. Zhu XM, Li L, Wu M, Liang S, Shi HB, Liu XH, et al. Current opinions on autophagy in pathogenicity of fungi. *Virulence*. 2019;10:481–9. [DOI] [PubMed] [PMC]
25. Wang S, Hu W, Liu F. Autophagy in the Lifetime of Plants: From Seed to Seed. *Int J Mol Sci*. 2022;23:11410. [DOI] [PubMed] [PMC]
26. Zhang X, Chen LX, Ouyang L, Cheng Y, Liu B. Plant natural compounds: targeting pathways of autophagy as anti-cancer therapeutic agents. *Cell Prolif*. 2012;45:466–76. [DOI] [PubMed] [PMC]
27. Wang P, Wang T, Han J, Li M, Zhao Y, Su T, et al. Plant Autophagy: An Intricate Process Controlled by Various Signaling Pathways. *Front Plant Sci*. 2021;12:754982. [DOI] [PubMed] [PMC]
28. Nitsche BM, Burggraaf-van Welzen AM, Lamers G, Meyer V, Ram AF. Autophagy promotes survival in aging submerged cultures of the filamentous fungus *Aspergillus niger*. *Appl Microbiol Biotechnol*. 2013;97:8205–18. [DOI] [PubMed]
29. Jiang ST, Chang AN, Han LT, Guo JS, Li YH, Liu TB. Autophagy Regulates Fungal Virulence and Sexual Reproduction in *Cryptococcus neoformans*. *Front Cell Dev Biol*. 2020;8:374. [DOI] [PubMed] [PMC]
30. Chen L, Zhang X, Wang W, Geng X, Shi Y, Na R, et al. Network and role analysis of autophagy in *Phytophthora sojae*. *Sci Rep*. 2017;7:1879. [DOI] [PubMed] [PMC]
31. Martel J, Ojcius DM, Wu CY, Peng HH, Voisin L, Perfettini JL, et al. Emerging use of senolytics and senomorphics against aging and chronic diseases. *Med Res Rev*. 2020;40:2114–31. [DOI] [PubMed]
32. Zhao W, Zhao H, Wang H, He Y. Research progress on the relationship between leaf senescence and quality, yield and stress resistance in horticultural plants. *Front Plant Sci*. 2022;13:1044500. [DOI] [PubMed] [PMC]
33. Häffner E, Konietzki S, Diederichsen E. Keeping Control: The Role of Senescence and Development in Plant Pathogenesis and Defense. *Plants (Basel)*. 2015;4:449–88. [DOI] [PubMed] [PMC]
34. Kacprzyk J, Burke R, Armengot L, Coppola M, Tattrie SB, Vahldick H, et al. Roadmap for the next decade of plant programmed cell death research. *New Phytol*. 2024;242:1865–75. [DOI] [PubMed]
35. Woo J, Shin S, Ji H, Ryu D, Cho E, Kim Y, et al. *Isatis tinctoria* L. Leaf Extract Inhibits Replicative Senescence in Dermal Fibroblasts by Regulating mTOR-NF- κ B-SASP Signaling. *Nutrients*. 2022;14:1979. [DOI] [PubMed] [PMC]
36. Barna B. Manipulation of Senescence of Plants to Improve Biotic Stress Resistance. *Life (Basel)*. 2022;12:1496. [DOI] [PubMed] [PMC]
37. Rapp YG, Ransbotyn V, Grafi G. Senescence Meets Dedifferentiation. *Plants (Basel)*. 2015;4:356–68. [DOI] [PubMed] [PMC]

38. Zhang Y, Huang D, Miao Y. Epigenetic control of plant senescence and cell death and its application in crop improvement. *Front Plant Sci.* 2023;14:1258487. [DOI] [PubMed] [PMC]
39. Zhu Y, Du M, Jiang X, Huang M, Zhao J. Nitric Oxide Acts as an Inhibitor of Postharvest Senescence in Horticultural Products. *Int J Mol Sci.* 2022;23:11512. [DOI] [PubMed] [PMC]
40. Flors V, Miralles C, Cerezo M, González-Bosch C, García-Agustín P. Effect of a novel chemical mixture on senescence processes and plant–fungus interaction in Solanaceae plants. *J Agric Food Chem.* 2001;49:2569–75. [DOI] [PubMed]
41. Chong Z, Matsuo H, Kuroda M, Yamashita S, Parajuli GP, Manandhar HK, et al. Mushroom extract inhibits ultraviolet B-induced cellular senescence in human keratinocytes. *Cytotechnology.* 2018;70:1001–8. [DOI] [PubMed] [PMC]
42. Maheshwari R, Navaraj A. Senescence in fungi: the view from *Neurospora*. *FEMS Microbiol Lett.* 2008;280:135–43. [DOI] [PubMed]
43. Bertelsen JR, De Neergaard E, Smedegaard-Petersen V. Fungicidal effects of azoxystrobin and epoxiconazole on phyllosphere fungi, senescence and yield of winter wheat. *Plant Pathol.* 2001;50:190–205. [DOI]
44. Dussarrat T, Prigent S, Latorre C, Bernillon S, Flandin A, Díaz FP, et al. Predictive metabolomics of multiple Atacama plant species unveils a core set of generic metabolites for extreme climate resilience. *New Phytol.* 2022;234:1614–28. [DOI] [PubMed] [PMC]
45. Ofir R, Lev R, Ron M, Stavi I. Analysis of herbal medicine among Bedouin of the Saint Catherine Protectorate (southern Sinai Peninsula) and its comparison to modern drug design. *Sustainable Environ.* 2023;9:2278831. [DOI]
46. Zhou S, Yan X, Yang J, Qian C, Yin X, Fan X, et al. Variations in Flavonoid Metabolites Along Altitudinal Gradient in a Desert Medicinal Plant *Agriophyllum squarrosum*. *Front Plant Sci.* 2021;12:683265. [DOI] [PubMed] [PMC]
47. Lotfy RA, Fahmy DM, Ahmed FA. QUALITATIVE AND QUANTITATIVE DETERMINATION OF SECONDARY METABOLITES OF 26 MEDICINAL PLANTS FROM SOUTHEASTERN OF EGYPT. *Egypt J Desert Res.* 2015;65:309–26. [DOI]
48. Liu P, Tan Y, Yang J, Wang YD, Li Q, Sun BD, et al. Bioactive secondary metabolites from endophytic strains of *Neocamarosporium betae* collected from desert plants. *Front Plant Sci.* 2023;14:1142212. [DOI] [PubMed] [PMC]
49. Zhang XY, Tan XM, Yu M, Yang J, Sun BD, Qin JC, et al. Bioactive metabolites from the desert plant-associated endophytic fungus *Chaetomium globosum* (Chaetomiaceae). *Phytochemistry.* 2021;185:112701. [DOI] [PubMed]
50. Li LY, Wang YD, Liu ZL, Sun BD, Yu M, Niu SB, et al. Resorcylic acid analogs from the desert plant endophytic fungus *Rhinocladiella similis*. *Mycosystema.* 2020;39:589–98. [DOI]
51. Dabravolski SA, Isayenkov SV. Metabolites Facilitating Adaptation of Desert Cyanobacteria to Extremely Arid Environments. *Plants (Basel).* 2022;11:3225. [DOI] [PubMed] [PMC]
52. He J, Wijeratne EM, Bashyal BP, Zhan J, Seliga CJ, Liu MX, et al. Cytotoxic and other metabolites of *Aspergillus* inhabiting the rhizosphere of Sonoran desert plants. *J Nat Prod.* 2004;67:1985–91. [DOI] [PubMed]
53. Meng T, Xiao D, Muhammed A, Deng J, Chen L, He J. Anti-Inflammatory Action and Mechanisms of Resveratrol. *Molecules.* 2021;26:229. [DOI] [PubMed] [PMC]
54. S N Chaitanya N, Devi A, Sahu S, Alugoju P. Molecular mechanisms of action of Trehalose in cancer: A comprehensive review. *Life Sci.* 2021;269:118968. [DOI] [PubMed]
55. Sadeghi M, Dehnavi S, Asadirad A, Xu S, Majeed M, Jamialahmadi T, et al. Curcumin and chemokines: mechanism of action and therapeutic potential in inflammatory diseases. *Inflammopharmacology.* 2023;31:1069–93. [DOI] [PubMed] [PMC]