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Metabolic adaption of epithelial cells in asthma: a window to the initiation of carcinogenesis?

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Abstract

Recent data has resulted in an interest in the metabolic shift in cellular metabolism to aerobic glycolysis, increased reactive oxygen species (ROS), and mitochondrial dysfunction associated with asthma. There has been a push to better understand the immune and metabolic changes in allergy to improve understanding of disease pathology and treatment. Aerobic glycolysis seen in epithelial cells in asthma promotes chronic inflammation and the production of inflammatory cytokines. Asthma epithelial cells share a number of features proposed in the stages of cancer initiation including aerobic glycolysis and increased apoptosis with proliferation, all within a chronic inflammatory microenvironment. Metabolic reprogramming in malignant cells has been widely investigated since the glycolytic characteristics were first described last century. It is still debated whether these metabolic changes are the cause or consequence of carcinogenesis and oncogenic cell-selective pressures. Although historic results have been conflicting, recent data has found an increased lung cancer risk in asthma patients, independent of risk factors. A review of emerging research on the metabolic changes seen in asthma helps us to propose a pathway between the initiation of aerobic glycolysis and the selective pressures of the epithelial microenvironment and resulting malignant transformation risk.

Keywords

Asthma, cancer, mitochondria, metabolism, epithelial cells, oxidative stress

Introduction

Asthma is a complex respiratory disease characterized by chronic airway inflammation caused by excessive T-helper 2 responses. Pathological features include airway hyper-responsiveness and remodeling, eosinophilic inflammation, and mucus hypersecretion. Studies have repeatedly found the presence of metabolic anomalies and pathway variations in asthmatic patients, which have opened a further potential avenue for new treatments.

Epithelial cells in the airway of asthmatics play a key role in the pathogenesis of asthma [1]. Mitochondrial dysfunction in epithelial cells has recently been linked to the pathogenesis of asthma and

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other respiratory diseases [2]. Defects in mitochondrial metabolism are involved in the airway remodeling seen in asthma and also lead to increased oxidative stress, reduced adenosine triphosphate (ATP) synthase activity, and apoptosis [3]. Cell growth and division increase ATP and biosynthetic demands, therefore proliferating cells present an altered metabolic profile.

Normal lungs have been noted to generate more ATP through aerobic glycolysis than other tissues, with around 40% of glucose converted to lactate regardless of oxygen status in studies of rat lungs [4]. Recent data suggests asthma is associated with a change in cellular metabolism towards increased aerobic glycolysis in several cell types [5]. Over the last decade, an increased shift to glycolysis has been confirmed in bronchial epithelial cells in asthmatic patients [6]. The cell aerobic glycolysis seen in epithelial cells in asthmatics results in an increase in inflammatory cytokines and promotes a chronic airway inflammatory response and immune activation [7]. In the 1920s, it was described how malignant cells largely perform aerobic glycolysis despite the presence of oxygen. Warburg [8] proposed this damage in mitochondrial respiration was followed by an increase in glycolysis to make up for the loss in energy availability.

We now know that the working model of carcinogenesis proposes that oncogenic signalling is the underlying driver of malignant cell transformation, which in turn alters cell metabolism pathways. The tumour suppressor protein, p53, has now been decisively linked to the altered balance between oxidative phosphorylation (OXPHOS) and glycolysis seen in carcinogenesis [9]. This data helps corroborate malignant gene mutations to the Warburg effect on the level of tumour suppressors. Chandel [10] reported that mitochondrial metabolism and electron transport chain (ETC) function are required for malignant cell growth, again moving away from the notion of the role of damaged mitochondria in carcinogenesis proposed by Warburg [8]. Researchers have suggested that this metabolic shift could be caused by cancerassociated hypoxia, however, it is a feature in many cells without an oxygen limitation [11]. Other models suggest that increased glycolysis can shunt biosynthetic precursors into anabolic reactions that branch from this pathway. However, again this is challenged as glycolytic intermediates are not necessarily elevated in proliferating cells [12]. It is still debated whether these metabolic changes are the cause or consequence of carcinogenesis and oncogenic cell selective pressures [13].

This phenotype is not unique to cancer and we now appreciate that aerobic glycolysis is exhibited by many proliferative non-transformed mammalian cells [14]. Along with this, activated T cells utilize aerobic glycolysis and it has been implicated in augmenting effector T cell response [15]. What drives aerobic glycolysis and why it is associated with proliferation in some cell populations have not yet been fully explained.

Large meta-analysis and cohort studies on adults with active asthma have found an increased lung cancer incidence, independent of smoking status [16]. We review the selective pressures of asthma epithelial chronic inflammatory microenvironment and the effect of this on apoptosis and metabolic pathway changes.

OXPHOS and reactive oxygen species

In a cell's mitochondrial inner membrane, ATP is generated by the transport of electrons to a series of transmembrane protein complexes: the ETC. The reduced co-enzymes nicotinamide adenine dinucleotide + hydrogen (NADH) or FADH₂ act as electron donors in this pathway. Complex I of the ETC accepts electrons from NADH and passes these onto complex II while oxidizing NADH to NAD⁺. As the electrons move through the ETC complexes I to IV, protons are moved from the mitochondrial matrix into the inter-membrane space by complexes I, III, and IV [16]. This results in an increased proton gradient across the mitochondrial matrix through complex V causing the synthesis of ATP through the transmembrane protein ATP synthase. In this pathway, oxygen behaves as the terminal electron acceptor, as it is a strong oxidizing agent [17]. The reduction of oxygen does involve the production of potentially damaging intermediates [18]. Although the transfer of four electrons and four protons reduces oxygen to water, the transfer of one or two electrons produces superoxide or peroxide anions: reactive oxygen species (ROS). Oxidative stress can be the

outcome of increased ROS production, an impaired antioxidant system, poor mitochondrial function, or a mixture of these [19].

Oxidative stress, the imbalance between production and accumulation of ROS in cells, plays a key role in the development of asthma [20]. Data has confirmed extensive free radical oxidative effects in airway tissue of ovalbumin-sensitized mice in asthma models [21]. Oxidative stress can disrupt calcium homeostasis and ETC activity, cause mitochondrial DNA mutations, and increase mitochondrial membrane permeability [22]. A fall in mitochondrial membrane potential results in a reduced proton gradient and therefore limits OXPHOS and is a hallmark of mitochondrial dysfunction [23]. Oxidative stress can inhibit ETC complex activity, thus limiting NADH oxidation at complex I [24, 25]. Mitochondrial dysfunction has been implicated in bronchial asthma epithelial cells [26]. Such dysfunction has also been seen in T2 low asthma and therefore is independent of immune profile [27]. Evidence over the last few years has been increasing indicating that mitochondrial dysfunction is involved in the pathogenesis of asthma [2]. Chellappan et al. [2] also noted drugs to target mitochondrial function could also be used in asthma treatment.

As a process, chronic inflammation in asthma generates oxidative stress in pulmonary cells. Oxidative stress initiates the production of mediators of inflammation and ROS in pulmonary epithelial cells and other inflammatory cells [28]. ROS can result in oxidative damage to bronchial epithelial cells in asthma, which promotes inflammation through the release of cytokines [29]. Damage to the airway epithelium may play a role in the increase of particles reaching the airway submucosa, causing increased inflammation and airway remodeling [30].

An interesting observation was made by Raby et al. [31] to link the maternally-inherited asthma risk factor to mitochondrial inheritance, which is known to be exclusively maternally inherited. This highlights the importance of mitochondrial DNA and mitochondrial dysfunction in asthma pathogenesis.

Aerobic glycolysis

Glycolysis is a sequence of reactions that oxygen is used to reoxidise NADH to NAD⁺. This pathway starts with a molecule of glucose and ends up with two of pyruvate, which is usually only converted to lactate in the absence of oxygen. Glycolysis can occur in aerobic and anaerobic conditions. In aerobic conditions, pyruvate is directed towards the tricarboxylic acid cycle to ultimately undergo OXPHOS. In the first step of this in mitochondria, pyruvate is decarboxylated by pyruvate dehydrogenase (PDH) complex to form acetyl-CoA that is moved into the tricarboxylic acid cycle. PDH activity is limited by a low NAD⁺/NADH ratio, therefore PDH influences the extent a cell engages in this pathway [32]. In anaerobic conditions, pyruvate stays in the cytoplasm and is fermented to lactate by lactate dehydrogenase. In aerobic glycolysis, also known as the Warburg effect, pyruvate can be fermented to lactate even in the presence of oxygen or further oxidized. The pace of ATP generation in aerobic glycolysis is quicker than OXPHOS, which is thought to suit the energy demands of rapid proliferation [33]. Mechanism of regulation of glycolysis occurs through modification of rate-limiting enzymes, their allosteric activation or inhibition, and by hormonal control.

While a lot of research has been carried out in the area of cancer, we know that aerobic glycolysis is exhibited by many proliferative non-transformed mammalian cells. Current research suggests asthma to be associated with a shift in cell metabolism to aerobic glycolysis. Increased glycolysis was reported in bronchial epithelial cells from both obese and control asthma patients [6]. These changes have been linked to an overall increase in oxidative stress [34]. Inhibition of epithelial aerobic glycolysis was found to significantly decrease inflammatory cytokines, suggesting it is involved in the chronic inflammation seen in asthma [7]. A shift to aerobic glycolysis has emerged as a key feature of allergic asthma and has also been proposed as a feature of the development of mucus metaplasia, airway hyper-reactivity, and inflammation [35, 36]. Glycolysis generally occurs in oxygen-deficient conditions but in these studies cells or in vivo disease models were examined under normal oxygen conditions, or the oxygen consumption rates were monitored.

It has been demonstrated that the glycolytic phenotype is observed in human cancers and is often seen early in carcinogensis [37]. The glycolytic shift caused by oncogene activation and loss of tumour suppressors in carcinogenesis is thought to result in a selective advantage for tumours, by providing essential precursors for building the products to sustain growth and proliferation. The current working models of carcinogenesis propose DNA damage occurs, activating oncogenes with a subsequent metabolic shift [38].

The traditional thought that healthy proliferating cells require increased aerobic glycolysis does not always seem to be the case. A study of fibroblasts triggered into a quiet state compared to proliferating cells found no increase in glucose uptake in the proliferating population [39]. Recent findings in this field have found aerobic glycolysis is not just limited to supporting proliferation. Cells can alter their metabolism to adjust to energy requirements and signalling events [40].

Why some cells engage in aerobic glycolysis remains poorly understood. Early proposals of mitochondrial dysfunction or hypoxia being the cause of these changes in carcinogenesis have been strongly challenged with a better understanding of the role of the functional ETC in malignant cells. Additionally, it is noted tumours have functional mitochondria, which are required for growth and progression. Healthy endothelial cells, in the process of vessel sprouting, need to differentiate and proliferate in a synchronized way. These healthy cells have been found to have high rates of aerobic glycolysis that results in lactate production rather than OXPHOS, regardless of the fact they are in direct contact with oxygenated blood, which again challenges the role of hypoxia as the cause of this metabolic change [41]. An additional previous proposal is that aerobic glycolysis provides essential biosynthetic precursors for anabolic reactions branching from glycolysis. However, conflicting data has noted most of the biomass from proliferating cells is derived from amino acids rather than glucose derivatives, which again challenges this notion [42]. Other authors have proposed that aerobic glycolysis may protect against the increased oxidative stress seen in a proliferating cell [43]. Aerobic glycolysis has recently been found to be a rate-limiting factor for the proliferation and survival of human bronchial epithelial cells [44].

NAD⁺ demand driving aerobic glycolysis

Why aerobic glycolysis is commonly associated with proliferation remains an ongoing question. However, a recent model of aerobic glycolysis has led to some promising data to help explain why some cells shift their metabolism. This showed that aerobic glycolysis reflects a state in which the demand for NAD⁺ for oxidation reactions is greater than the cell demand for ATP [45]. The authors found that when NAD⁺ regeneration by mitochondrial respiration became limited, fermentation was increased, despite available oxygen. They confirmed their hypothesis that mitochondrial respiration and ATP synthase activity were insufficient to support NAD⁺ regeneration so cells engaged in aerobic glycolysis. It was also noted that suppressing aerobic glycolysis decreased proliferation in both malignant and control cell populations. Inhibiting aerobic glycolysis decreased the NAD⁺/NADH ratio in cells and increased dependency on the mitochondrial ETC for NAD⁺ regeneration. The decreased NAD⁺/NADH ratio observed in aerobic glycolysis-inhibited cells suggested that NAD⁺ regeneration by mitochondrial respiration was not able to support increased proliferation. This research was consistent with previous evidence that the rate of NADH oxidation could limit biomass synthesis in aerobic conditions if proliferation is quick enough, in the context of impaired mitochondrial OXPHOS capacity [46]. This also supports previous proposals on the role of aerobic glycolysis in protecting against oxidative stress in proliferating cells [43].

Of note, lung mitochondrial NAD⁺/NADH ratios have been found to be significantly lower in models of airway inflammation [47]. The authors of this study also confirmed a shift to aerobic glycolysis and a much-reduced ETC complex I activity in asthmatics in a model of allergic airway inflammation. In asthma, recovering mitochondrial function, such as through physical exercise, could potentially reduce aerobic glycolysis as seen in malignancy [48].

Apoptosis and proliferation in asthmatic epithelial cells

In apoptosis, the intrinsic and extrinsic pathways mediate the cell death signalling cascade. The intrinsic pathway is mitochondrial-mediated and triggered by a number of cellular stresses, including oxidative stress. The extrinsic pathway refers to a death receptor-mediated activation pathway. Both pathways come together at the point of initiator and effector caspase activation. In the pathogenesis of asthma, epithelial cells respond to allergens by producing inflammatory cytokines. Epithelial cells can undergo apoptotic cell death in response to allergen exposure, which further drives the inflammatory cascade. Asthma and Thelper 2-mediated airway inflammation are associated with airway epithelial cell apoptosis [49]. Whilst epithelial apoptosis is commonly described, it has not universally been identified in models of asthma [50, 51]. Its relevance to asthma pathogenesis remains uncertain. Despite this, a number of interesting observations of apoptosis in asthma, particularly in severe disease, have been made. A number of factors, including infection and excess oxidative stress, have been described in mediating apoptosis in asthma [52]. Apoptotic epithelial cell clusters have been found in sputum and bronchoalveolar lavage samples in asthmatics [53]. An apoptosis pan-caspase inhibitor was found to decrease airway inflammation in an asthma model [54]. It has been described how T cells and eosinophils induce apoptosis in epithelial cells through the secretion of IFN-gamma and TNF-alpha [55]. A number of studies have demonstrated epithelial cell apoptosis after allergen exposure [56, 57].

There are multiple signals that initiate apoptosis including death receptors, cytokines, and oxidative stress. One such death receptor identified in epithelial cells is CD95 [58], which may also be increased in the asthmatic epithelium linking directly to the increase in apoptosis seen in asthma [49].

The intrinsic pathway of apoptosis is also strongly linked to asthma, as there is a significant overlap between asthma, mitochondrial dysfunction, oxidative stress, and apoptosis [59]. Epithelial cell apoptosis is closely linked with increased cell proliferation and may be stimulated by the same signals to maintain homeostasis [60]. Allergen insults have been found to stimulate epithelial cell proliferation in asthmatics [61]. It has been found that apoptosis and proliferation may both be elevated in chronic inflammatory disease states such as asthma [62]. More significant apoptosis, cell cycling, and proliferation are seen in asthma airway epithelium and may be linked to disease severity [62, 63].

Apoptosis-induced proliferation has been widely described in wound healing, cancer, and other disease processes, where activated caspases trigger compensatory proliferation [64]. Whilst the role of caspases in apoptosis is well understood, recent studies have found they can promote proliferation through autonomous regulation of the cell cycle as well as signaling neighbouring tissue [65]. Apoptosis-induced proliferation is believed to be used to maintain tissue morphology following unexpected cell population loss, where apoptotic cells trigger surviving cells to proliferate. It also involves a number of other mediators including increased oxidative stress and immune cell recruitment [66]. An increase in epidermal growth factor receptor expression has been found in the epithelium of asthmatic patients [67]. This plays a key role in the epithelial repair process by mediating cell proliferation and differentiation to maintain tissue morphology in the airway. While this is still not conclusive in current asthma models, increased apoptosis and increased epithelial proliferation have been observed in a number of studies, particularly related to the severity of asthma [62, 68]. Further work is needed to fully establish this pathway in asthma pathogenesis.

Metabolic pathway to asthma pathogenesis

From the evidence we have reviewed so far, we can make several observations (Table 1). Firstly, mitochondrial dysfunction and oxidative stress are now recognized to play a key role in asthma pathogenesis. There is a strong link between oxidative stress and the increase in apoptosis via the intrinsic pathway, particularly linked to the severity of asthma inflammation. Increased epithelial proliferation is observed in asthma, but it is not certain whether this is a result of increased apoptosis or a combination of other pathway pressures. However, apoptosis-induced proliferation has been appreciated in a number of chronic inflammatory diseases. Finally, the asthma epithelium has been shown to display increased oxidative stress, mitochondrial dysfunction, and a shift in metabolism to aerobic glycolysis.

Table 1. Summary of select	ed supporting literature
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Relevant findings	Author [Ref]	Date of publication	Study design
Mitochondrial dysfunction is increased in the asthmatic airway epithelium	Zhao et al. [27]	2023	Gene analysis of airway epithelial brushings from healthy and asthmatic patients
Increased oxidative stress and damage in asthmatic patients	Gazdík et al. [20]	2002	Levels of co-enzyme Q10 measured in asthmatic and control patients
Bronchial epithelial cells display increased glycolysis in asthmatics	Winnica et al. [<mark>6</mark>]	2019	Airway epithelial cells isolated from healthy and asthmatic patients
Aerobic glycolysis is up-regulated in airway epithelial cells in asthma	Yu et al. [7]	2022	Ovalbumin-sensitized mouse model and in vitro investigation of human bronchial epithelial cell lines
Asthma increases epithelial cell apoptosis	lwata et al. [54]	2003	Ovalbumin-sensitized mouse model
	Trautmann et al. [55]	2005	Evaluated epithelial layer of bronchial biopsies from asthma patients
	Truong-Tran et al. [56]	2002	Ovalbumin-sensitized mouse model
	Dorscheid et al. [57]	2003	Ovalbumin-sensitized mouse model
Proliferation is increased in asthma airway epithelial cells after allergen challenge	Hastie et al. [<mark>61</mark>]	2002	In vitro study of human epithelial cells from asthmatic and non-asthmatics
Epithelial cell proliferation contributes to airway remodeling in severe asthma	Cohen et al. [<mark>62</mark>]	2007	Epithelial cells examined from bronchial biopsy from asthmatic and healthy patients
Reduced ETC complex I and III activity in airway inflammation in asthmatics	Niu et al. [47]	2020	Lung tissue and monocytes from ovalbumin- sensitized mouse model evaluated
Increased demand for NAD ⁺ drives up- regulation of aerobic glycolysis	Luengo et al. [45]	2021	In vitro investigation of malignant and non- transformed cell populations (airway cell lines not involved in the study)
Aerobic glycolysis rate limiting factor for proliferation in airway epithelial cells	Park et al. [44]	2024	In vitro study of bronchial epithelial cells
Inflammation-induced cell proliferation potentiates DNA damage-induced mutations	Kiraly et al. [69]	2015	In vivo mouse model examining overall mutation rate
Asthma significantly associated with increased lung cancer risk	Qu et al. [16]	2017	Meta-analysis

ETC: electron transport chain

From these facts and the discussion on the metabolic switch to aerobic glycolysis in proliferating cells in health and cancer, we propose a pathway to the metabolic changes seen in the asthma epithelium and subsequent lung cancer risk (Figure 1).

Initial allergen and subsequent immune activation increases inflammatory pressures, cytokine production, and oxidative stress in epithelial cells and the airway microenvironment. This increases oxidative stress and ROS production in epithelial cell mitochondria. Subsequently, rates of cell apoptosis increase with resulting increased survivor epithelial cell proliferation, as found with current chronic inflammation models. NADH turnover decreases due to mitochondrial dysfunction in the ETC from oxidative stress [70]. ROS and oxidative stress can also affect complex I of the ETC to generate more free radicals and result in complex I dysfunction [70]. Complex I is thought to be the most susceptible to oxidative stress [71]. Chronic ROS exposure causes oxidative damage to mitochondrial proteins, whilst acute ROS exposure can deactivate iron-sulphur centres of ETC complex I and III resulting in mitochondrial dysfunction at these complexes [72]. Oxidative stress depletes cellular NAD⁺ further reducing availability for its role as a co-factor [73]. To further support this statement, monocytes from asthmatic patients, as well as lung tissue from mouse models, displayed greatly reduced ETC complex I and III activities [47].

NAD⁺ is an essential electron acceptor and is a key requirement to increase biomass during proliferation [74]. Proliferating cells are unable to meet their NAD⁺ needs through OXPHOS, so show a metabolic switch to aerobic glycolysis, as seen in current working models [45]. Recent data proposed a role in NAD⁺ boosting therapy to suppress mast cell degranulation in a model of allergy, which is relevant to this discussion [75].



Figure 1. The pathway from chronic airway inflammation in asthma to the up-regulation in aerobic glycolysis in airway epithelial cells, and resulting lung cancer risk. ETC: electron transport chain; NADH: nicotinamide adenine dinucleotide + hydrogen. ---->: discussed under "Apoptosis and proliferation in asthmatic epithelial cells" section

Chronic inflammation-induced cell proliferation is thought to increase the frequency of DNA mutations and drive cancer initiation [69]. A cell metabolic shift to aerobic glycolysis can inhibit pathways that halt proliferation, including p53, again possibly increasing selection for cells with a suppressed p53 phenotype and a high rate of proliferation in the airway epithelium [76].

Asthma is associated with an increased incidence of lung cancer, independent of smoking risks [16]. This risk is directly linked to the severity of presenting asthma. A significant percentage of lung cancers are thought to originate from epithelial cells so the metabolic microenvironment in this setting is likely to play a key role in carcinogenesis [77]. As we have reviewed, there is considerable overlap between the metabolism of epithelial cells in asthma and the malignant microenvironment with mitochondrial dysfunction. This has interesting implications for the metabolic shift prior to malignant transformation in this setting.

Conclusions

The evolving understanding of the metabolic pathways seen in the bronchial epithelium and in other diseases provides an opportunity for the development of new strategies in asthma treatment. We have

proposed a pathway to the metabolic changes seen in asthma from the inflammation in the epithelial microenvironment and the resulting increase in oxidative stress and mitochondrial dysfunction. Reduced ETC NADH turnover in the inflammatory airway environment means cells are unable to meet increased NAD⁺ demand in proliferation, which results in a subsequent shift to aerobic glycolysis; this is supported by current models. Increased apoptosis and proliferation are observed in bronchial asthmatic epithelial cells, resulting in airway remodeling. In vivo studies have found that inflammation-induced cell proliferation potentiates the frequency of DNA mutations. Higher rates of cancer are seen in adults with active asthma, linked to disease severity. It would be easy to propose how inflammation, subsequent shift to aerobic glycolysis and proliferation, then DNA mutation, would lead to the initiation of carcinogenesis in this setting, especially as a large proportion of lung cancers are thought to originate from epithelial cells in the airways.

Interestingly, this puts cell metabolic shift to aerobic glycolysis and proliferation before DNA mutation in the pathway of carcinogenesis in the asthma microenvironment. The Warburg effect is currently believed to be an early event in carcinogenesis that is a consequence of an initial oncogenic mutation. Therefore, this is a significant change when compared to current working models of carcinogenesis by proposing the Warburg effect precedes DNA mutation in at least this setting. This also highlights the importance of controlling airway inflammation in asthma management in line with recent shifts in the Global Initiative for Asthma treatment guidelines to earlier use of inhaled steroids [78].

This hypothesis is obviously dependent on the current working models of aerobic glycolysis activation translating to pathways in healthy airway cell populations, related to proliferation and increased NAD⁺ demand with mitochondrial dysfunction. Caution also has to be taken applying in vitro data, in the context of abundant nutrients, to the underlying changes in asthmatic epithelial cells in a very different micro-environment. Further investigation is required in this area to further define these metabolic pathways and establish this proposal.

Abbreviations

ATP: adenosine triphosphate ETC: electron transport chain NADH: nicotinamide adenine dinucleotide + hydrogen OXPHOS: oxidative phosphorylation PDH: pyruvate dehydrogenase ROS: reactive oxygen species

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Author contributions

TD: Conceptualization, Writing—original draft, Writing—review & editing.

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The author declares that he has no conflicts of interest.

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References

- 1. Calvén J, Ax E, Rådinger M. The Airway Epithelium—A Central Player in Asthma Pathogenesis. Int J Mol Sci. 2020;21:8907.
- 2. Chellappan DK, Paudel KR, Tan NW, Cheong KS, Khoo SSQ, Seow SM, et al. Targeting the mitochondria in chronic respiratory diseases. Mitochondrion. 2022;67:15–37.
- 3. Qian L, Mehrabi Nasab E, Athari SM, Athari SS. Mitochondrial signaling pathways in allergic asthma. J Investig Med. 2022;70:863–82.
- 4. Liu G, Summer R. Cellular Metabolism in Lung Health and Disease. Annu Rev Physiol. 2019;81:403–28.
- 5. Qin Z, Chen Y, Wang Y, Xu Y, Liu T, Mu Q. Immunometabolism in the pathogenesis of asthma. Immunology. 2024;171:1–17.
- 6. Winnica D, Corey C, Mullett S, Reynolds M, Hill G, Wendell S, et al. Bioenergetic Differences in the Airway Epithelium of Lean *Versus* Obese Asthmatics Are Driven by Nitric Oxide and Reflected in Circulating Platelets. Antioxid Redox Signal. 2019;31:673–86.
- 7. Yu C, Huang W, Zhou Z, Liang S, Zhou Z, Liu J, et al. Short isoform thymic stromal lymphopoieten reduces inflammation and aerobic glycolysis of asthmatic airway epithelium by antagonizing long isoform thymic stromal lymphopoientin. Respir Res. 2022;23:75.
- 8. Warburg O. The Metabolism of Carcinoma Cells. J Cancer Res. 1925;9:148–63.
- 9. Vousden KH, Ryan KM. p53 and metabolism. Nat Rev Cancer. 2009;9:691–700.
- 10. Chandel NS. Mitochondria and cancer. Cancer Metab. 2014;2:8.
- 11. Vander Heiden M, Cantley L, Thompson C. Understanding the Warburg effect: the metabolic requirements of cell proliferation. Science. 2009;324:1029–33.
- 12. Lunt SY, Muralidhar V, Hosios AM, Israelsen WJ, Gui DY, Newhouse L, et al. Pyruvate kinase isoform expression alters nucleotide synthesis to impact cell proliferation. Mol Cell. 2015;57:95–107.
- 13. Devic S. Warburg Effect a Consequence or the Cause of Carcinogenesis? J Cancer. 2016;7:817–22.
- 14. Epstein T, Xu L, Gillies RJ, Gatenby RA. Separation of metabolic supply and demand: aerobic glycolysis as a normal physiological response to fluctuating energetic demands in the membrane. Cancer Metab. 2014;2:7.
- 15. Peng M, Yin N, Chhangawala S, Xu K, Leslie CS, Li MO. Aerobic glycolysis promotes T helper 1 cell differentiation through an epigenetic mechanism. Science. 2016;354:481–4.
- 16. Qu YL, Liu J, Zhang LX, Wu CM, Chu AJ, Wen BL, et al. Correction: Asthma and the risk of lung cancer: a meta-analysis. Oncotarget. 2017;8:48525. Erratum for: Oncotarget. 2017;8:11614–20.
- 17. Kumari A. Sweet Biochemistry: Remembering Structures, Cycles, and Pathways by Mnemonics. Elsevier; 2018.
- 18. Davies KJ. The Oxygen Paradox, oxidative stress, and ageing. Arch Biochem Biophys. 2016;595:28–32.
- 19. Cui H, Kong Y, Zhang H. Oxidative stress, mitochondrial dysfunction, and aging. J Signal Transduct. 2012;2012:646354.
- 20. Gazdík F, Gvozdjakova A, Horvathova M, Weissova S, Kucharska J, Pijak MR, et al. Levels of coenzyme Q10 in asthmatics. Bratisl Lek Listy. 2002;103:353–6.

- 21. Reddy PH. Mitochondrial Dysfunction and Oxidative Stress in Asthma: Implications for Mitochondria-Targeted Antioxidant Therapeutics. Pharmaceuticals (Basel). 2011;4:429–56.
- 22. Kowalczyk P, Sulejczak D, Kleczkowska P, Bukowska-Ośko I, Kucia M, Popiel M, et al. Mitochondrial Oxidative Stress—A Causative Factor and Therapeutic Target in Many Diseases. Int J Mol Sci. 2021;22: 13384.
- 23. Liu S, Liu S, He B, Li L, Wang J, Cai T, et al. OXPHOS deficiency activates global adaption pathways to maintain mitochondrial membrane potential. EMBO Rep. 2021;22:e51606.
- 24. Cortés-Rojo C, Vargas-Vargas MA, Olmos-Orizaba BE, Rodríguez-Orozco AR, Calderón-Cortés E. Interplay between NADH oxidation by complex I, glutathione redox state and sirtuin-3, and its role in the development of insulin resistance. Biochim Biophys Acta Mol Basis Dis. 2020;1866:165801.
- 25. Balogh E, Veale D, McGarry T, Orr C, Szekanecz Z, Ng CT, et al. Oxidative stress impairs energy metabolism in primary cells and synovial tissue of patients with rheumatoid arthritis. Arthritis Res Ther. 2018;20:95.
- 26. Aghapour M, Remels A, Pouwels S, Bruder D, Hiemstra P, Cloonan, et al. Mitochondria: at the crossroads of regulating lung epithelial cell function in chronic obstructive pulmonary disease. Am J Physiol Lung Cell Mol Physiol. 2020;318:L149–64.
- 27. Zhao L, Gao J, Chen G, Huang C, Kong W, Feng Y, et al. Mitochondrial dysfunction in airway epithelial cells is associated with type 2-low asthma. Front Genet. 2023;14:1186317.
- 28. Valavanidis A, Vlachogianni T, Fiotakis K, Loridas S. Pulmonary oxidative stress, inflammation and cancer: respirable particulate matter, fibrous dust and ozone as major causes of lung carcinogenesis through reactive oxygen species mechanisms. Int J Environ res Public Health. 2013;10:3886–907.
- 29. Boonpiyathad T, Sözener ZC, Satitsuksanoa P, Akdis CA. Immunologic mechanisms in asthma. Semin Immunol. 2019;46:101333.
- 30. Gon Y, Hashimoto S. Role of airway epithelial barrier dysfunction in pathogenesis of asthma. Allergol Int. 2018;67:12–7.
- Raby BA, Klanderman B, Murphy A, Mazza S, Camargo Ca Jr, Silverman EK, et al. A common mitochondrial haplogroup is associated with elevated total serum IgE levels. J Allergy Clin Immunol. 2007;120:351–8.
- 32. Grassian AR, Metallo CM, Coloff JL, Stephanopoulos G, Brugge JS. Erk regulation of pyruvate dehydrogenase flux through PDK4 modulates cell proliferation. Genes Dev. 2011;25:1716–33.
- 33. Vazquez A, Liu J, Zhou Y, Oltvai ZN. Catabolic efficiency of aerobic glycolysis: the Warburg effect revisited. BMC Syst Biol. 2010;4:58.
- 34. Holguin F. Oxidative stress in airway diseases. Ann Am Thorac Soc. 2013;10:S150–7.
- 35. Qian X, Aboushousha R, van de Wetering C, Chia SB, Amiel E, Schneider RW, et al. IL-1/inhibitory κB kinase ε–induced glycolysis augment epithelial effector function and promote allergic airways disease. J Allergy Clin Immunol. 2018;142:435–50.E10.
- 36. van de Wetering C, Aboushousha R, Manuel AM, Chia SB, Erickson C, MacPherson MB, et al. Pyruvate Kinase M2 Promotes Expression of Proinflammatory Mediators in House Dust Mite–Induced Allergic Airways Disease. J Immunol. 2020;204:763–74.
- 37. Cruz MD, Ledbetter S, Chowdhury S, Tiwari AK, Momi N, Wali RK, et al. Metabolic reprogramming of the premalignant colonic mucosa is an early event in carcinogenesis. Oncotarget. 2017;8:20543–57.
- 38. Couch DB. Carcinogenesis: basic principles. Drug chem Toxicol. 1996;19:133–48.
- 39. Lemons JM, Feng XJ, Bennett BD, Legesse-Miller A, Johnson EL, Raitman I, et al. Quiescent Fibroblasts Exhibit High Metabolic Activity. PLoS Biol. 2010;8:e1000514.
- 40. Jones W, Bianchi K. Aerobic glycolysis: beyond proliferation. Front Immunol. 2015;6:227.
- 41. De Bock K, Georgiadou M, Schoors S, Kuchnio A, Wong BW, Cantelmo AR, et al. Role of PFKFB3-Driven Glycolysis in Vessel Sprouting. Cell. 2013;154:651–63.

- 42. Hosios AM, Hecht VC, Danai LV, Johnson MO, Rathmell JC, Steinhauser ML, et al. Amino Acids Rather than Glucose Account for the Majority of Cell Mass in Proliferating Mammalian Cells. Dev Cell. 2016; 36:540–9.
- 43. Brand K. Aerobic Glycolysis by Proliferating Cells: Protection against Oxidative Stress at the Expense of Energy Yield. J Bioenerg Biomembr. 1997;29:355–64.
- 44. Park SH, Kim G, Yang GE, Yun HJ, Shin TH, Kim ST, et al. Disruption of phosphofructokinase activity and aerobic glycolysis in human bronchial epithelial cells by atmospheric ultrafine particulate matter. J Hazard Mater. 2024;464:132966.
- 45. Luengo A, Li Z, Gui DY, Sullivan LB, Zagorulya M, Do BT, et al. Increased demand for NAD⁺ relative to ATP drives aerobic glycolysis. Mol Cell. 2021;81:691–707.E6.
- 46. Fernandez-de-Cossio-Diaz J, Vazquez A. Limits of aerobic metabolism in cancer cells. Sci Rep. 2017;7: 13488.
- 47. Niu Y, Nie Q, Dong L, Zhang J, Liu SF, Song W, et al. Hydrogen Attenuates Allergic Inflammation by Reversing Energy Metabolic Pathway Switch. Sci Rep. 2020;10:1962.
- 48. Clemente-Suárez VJ, Martín-Rodríguez A, Redondo-Flórez L, Ruisoto P, Navarro-Jiménez E, Ramos-Campo DJ, et al. Metabolic Health, Mitochondrial Fitness, Physical Activity, and Cancer. Cancers (Basel). 2023;15:814.
- 49. White S. Apoptosis and the airway epithelium. J Allergy (Cairo). 2011;2011:948406.
- 50. Lambrecht BN, Hammad H. Death at the epithelium in asthma. Cell Res. 2013;23:588–9.
- 51. Ordoñez C, Ferrando R, Hyde DM, Wong HH, Fahy JV. Epithelial desquamation in asthma, artifact or pathology? Am J Respir Crit Care Med. 2000;162:2324–9.
- 52. Tesfaigzi Y. Roles of apoptosis in airway epithelia. Am J Respir Cell Mol Biol. 2006;34:537–47.
- 53. Juncadella IJ, Kadl A, Sharma AK, Shim YM, Hochreiter-Hufford A, Borish L, et al. Apoptotic cell clearance by bronchial epithelial cells critically influences airway inflammation. Nature. 2023;493: 547–51.
- 54. Iwata A, Nishio K, Winn RK, Chi EY, Henderson WR Jr, Harlan JM. A broad spectrum caspase inhibitor attenuates allergic airway inflammation in murine asthma model. J Immunol. 2003;170:3386–91.
- 55. Trautmann A, Kruger K, Akdis M, Muller-Wening D, Akkaya A, Brocker EB, et al. Apoptosis and loss of adhesion of bronchial epithelial cells in asthma. Int Arch Allergy Immunol. 2005;138:142–50.
- 56. Truong-Tran AQ, Ruffin RE, Foster PS, Koskinen AM, Coyle P, Philcox JC, et al. Altered zinc homeostasis and caspase-3 activity in murine allergic airway inflammation. Am J Respir Cell Mol Biol. 2002;27:286–96.
- 57. Dorscheid DR, Low E, Conforti A, Shifrin S, Sperling AI, White SR. Corticosteroid-induced apoptosis in mouse airway epithelium: effect in normal airway and after allergen-induced airway inflammation. J Allergy Clin Immunol. 2003;111:360–6.
- 58. Hamann KJ, Dorscheid DR, Ko FD, Conforti AE, Sperling AI, Rabe KF, et al. Expression of Fas (CD95) and FasL (CD95L) in Human Airway Epithelium. Am J Respir Cell Mol Biol. 1998;19:537–42.
- 59. Liu L, Zhou L, Wang LL, Zheng P, Zhang FQ, Mao ZY, et al. Programmed Cell Death in Asthma: Apoptosis, Autophagy, Pyroptosis, Ferroptosis, and Necroptosis. J Inflamm Res. 2023;16:2727–54.
- 60. Jyonouchi H. Airway epithelium and apoptosis. Apoptosis. 1999;4:407–17.
- 61. Hastie AT, Kraft WK, Nyce KB, Zangrilli JG, Musani AI, Fish JE, et al. Asthmatic epithelial cell proliferation and stimulation of collagen production: human asthmatic epithelial cells stimulate collagen type III production by human lung myofibroblasts after segmental allergen challenge. Am J Respir Crit Care Med. 2002;165:266–72.
- 62. Cohen L, E X, Tarsi J, Ramkumar T, Horiuchi TK, Cochran R, et al.; NHLBI Severe Asthma Research Program (SARP). Epithelial cell proliferation contributes to airway remodeling in severe asthma. Am J Respir Crit care Med. 2007;176:138–45.

- 63. Trautmann A, Schmid-Grendelmeier P, Krüger K, Crameri R, Akdis M, Akkaya A, et al. T cells and eosinophils cooperate in the induction of bronchial epithelial cell apoptosis in asthma. J Allergy Clin Immunol. 2002;109:329–37.
- 64. Fogarty CE, Bergmann A. Killers creating new life: caspases drive apoptosis-induced proliferation in tissue repair and disease. Cell Death Differ. 2017;24:1390–400.
- 65. Pérez-Garijo A. When dying is not the end: Apoptotic caspases as drivers of proliferation. Semin Cell Dev Biol. 2018;82:86–95.
- 66. Fogarty CE, Diwanji N, Lindblad JL, Tare M, Amcheslavsky A, Makhijani K, et al. Extracellular reactive oxygen species drive apoptosis-induced proliferation via *Drosophila* macrophages. Curr Biol. 2016;26: 575–84.
- 67. Inoue H, Akimoto K, Homma T, Tanaka A, Sagara H. Airway Epithelial Dysfunction in Asthma: Relevant to Epidermal Growth Factor Receptors and Airway Epithelial Cells. J Clin Med. 2020;9:3698.
- 68. Heijink IH, Kuchibhotla VNS, Roffel MP, Maes T, Knight DA, Sayers I, et al. Epithelial cell dysfunction, a major driver of asthma development. Allergy. 2020;75:1902–17.
- 69. Kiraly O, Gong G, Olipitz W, Muthupalani S, Engelward BP. Inflammation-Induced Cell Proliferation Potentiates DNA Damage-Induced Mutations *In Vivo*. PLoS Genet. 2015;11:e1004901.
- 70. Srivastava S. Emerging therapeutic roles for NAD⁺ metabolism in mitochondrial and age-related disorders. Clin Transl Med. 2016;5:25.
- 71. Sharma L, Lu J, Bai Y. Mitochondrial respiratory complex I: structure, function and implication in human diseases. Curr Med Chem. 2009;16:1266–77.
- 72. Ghezzi D, Zeviani M. Assembly factors of human mitochondrial respiratory chain complexes: physiology and pathophysiology. Adv Exp Med Biol. 2012;748:65–106.
- 73. Massudi H, Grant R, Guillemin G, Braidy N. NAD⁺ metabolism and oxidative stress: the golden nucleotide on the crown of thorns. Redox Rep. 2012;17:28–46.
- 74. Han X, Simon MC. NAD⁺ regeneration drives cancer cell proliferation. Nat Metab. 2022;4:647–8.
- 75. Kim HW, Ryoo GH, Jang HY, Rah SY, Lee DH, Kim DK, et al. NAD⁺-boosting molecules suppress mast cell degranulation and anaphylactic responses in mice. Theranostics. 2022;12:3316–28.
- 76. Mason EF, Zhao Y, Goraksha-Hicks P, Coloff JL, Gannon H, Jones SN, et al. Aerobic glycolysis suppresses p53 activity to provide selective protection from apoptosis upon loss of growth signals or inhibition of BCR-Abl. Cancer Res. 2011;70:8066–76.
- 77. Sutherland KD, Berns A. Cell of origin of lung cancer. Mol Oncol. 2010;4:397–403.
- 78. 2023 GINA Report, Global Strategy for Asthma Management and Prevention [Internet]. Fontana: Global Initiative for Asthma; c2024 [cited 2023 Jul 10]. Available from: https://ginasthma.org/2023-g ina-main-report/