



The search for still unknown pathomechanisms of allergy

Gualtiero Leo^{1*} , Cristoforo Incorvaia², Stefania Arasi³ 

¹Pediatric Allergology and Pneumology Unit, Humanitas Medical Care – MCH S.r.l., 20089 Milan, Italy

²Allergy, Private Practice, 20100 Milan, Italy

³Allergy Diseases Research Area, Pediatric Allergology Unit, Bambino Gesù Children’s Hospital IRCCS, 00165 Rome, Italy

***Correspondence:** Gualtiero Leo, Pediatric Allergology and Pneumology Unit, Humanitas Medical Care – MCH S.r.l., via Manzoni, 113, 20089 Milan, Italy. gualtiero.leo@mc.humanitas.it

Academic Editor: Umit Murat Sahiner, Hacettepe University Faculty of Medicine, Turkey

Received: January 17, 2024 **Accepted:** April 11, 2024 **Published:** July 22, 2024

Cite this article: Leo G, Incorvaia C, Arasi S. The search for still unknown pathomechanisms of allergy. *Explor Asthma Allergy*. 2024;2:329–39. <https://doi.org/10.37349/ea.2024.00048>

Abstract

In recent decades, atopic diseases, such as atopic dermatitis (AD), allergic asthma (AA), allergic rhinitis (AR), and food allergy (FA) have been estimated rapidly increasing in prevalence. These diseases are characterized by the presence of specific immunoglobulin E (sIgE) and often relate to each other and develop in sequence (the so-called “atopic march”). AD may be the first early manifestation in infants followed by FA often within the first year of life. Moreover, AD is a risk factor for developing sensitization to airborne allergens later in life that can cause clinical manifestations of AA and AR. According to the dual-allergen exposure hypothesis, allergic sensitization to food allergens is promoted through cutaneous exposure, rather than the oral route. Moreover, there is evidence that exposure to food allergens, in particular peanuts, in the airway would also lead to food sensitization. The most frequent route of sensitization for inhalant allergens is still debated. Of note, a recent case report supports the development of sensitization to cat dander through a cat bite. Our review aims to provide an overview of current knowledge and unmet needs in the pathophysiology of respiratory and FAs.

Keywords

Allergy, mechanisms, skin barrier, airway

Introduction

The discovery of allergy dates back to 450 BC, when Greek doctors, using the methods of Hippocrates, identified a disease defined as “atopy” [ἄλλος (állos, “other”) + ἔργον (érgon, “work, activity”), i.e. “different or strange reaction”] for the definition caused by commonly harmless substances [1, 2]. Atopy was considered rare but, already in the Roman period of 30–40 BC, it appeared to be quite widespread. In particular, in this historical period, the first “diagnoses” of allergy were reported by Svetonius referring to eye swelling which prevented vision caused by contact with horses [2]. However, most substances with allergenic activity are not directly visible, and we had to wait until the years 1700–1800 to discover the etiologies of allergy, such as pollens and dust mites. Bostock discovered pollen allergy, which was defined

© The Author(s) 2024. This is an Open Access article licensed under a Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, sharing, adaptation, distribution and reproduction in any medium or format, for any purpose, even commercially, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.



as hay fever in 1819 [3]; in 1915, Voorhorst discovered the role of dust mites, and specially of *Dermatophagoides* species as triggering allergen [4]. Over time more and more allergenic sources have been identified, varying from biologic sources with very complex compositions (e.g., pollens, house dust mites, and foods), to single molecules such as chemicals (occupational allergy) or drugs (drug allergy) [5–7]. It is reasonable to imagine that new causes of allergy may be discovered in the future.

Furthermore, a dramatic rise in the prevalence of allergy has been reported in the last decades, as well as the development of increasingly effective treatments and management for allergy based on a better knowledge of underlying immunopathomechanisms [8].

In particular, intact skin and mucosal barriers play a key role in maintaining tissue homeostasis which protects tissues from harmful agents, whereas a defective epithelial barrier may facilitate allergic and autoimmune conditions [9].

This review focuses on the recent insights in the context of the pathomechanisms underlying allergy to airborne and food allergenic sources.

From the hygiene hypothesis to the skin-barrier hypothesis

The development of allergic diseases results from the interaction between genetic and environmental mechanisms. The increase in these diseases has occurred in relatively too short a time to be justified by genetic mutations for which considerable importance has been given to environmental factors linked to the modern lifestyle including changing dietary patterns, changing intestinal commensal bacteria, and vehicular pollution [10]. Studies suggest that environmental factors can produce epigenetic changes in gene expression and disease risk that may be potentially heritable across generations.

Already in 1989, Strachan [11] found an inverse relationship between the development of hay fever and the number of the family units and the birth-order hypothesizing that the increase in the prevalence of hay fever was to be attributed to a reduced number of infections in childhood. This concept has subsequently undergone some evolutions by attributing the increase in allergies not to the reduction of infections in the early stages of life but rather to a reduction in the colonization of the organism by commensal microorganisms [12]. Allergic diseases such as atopic dermatitis (AD), allergic asthma (AA), and allergic rhinitis (AR) present an alteration of the epithelial structure of the respective affected organs which causes a dysfunction of the barrier effect [13]. This dysfunction can favor the passage of external agents such as microorganisms and allergens responsible for the release of alarmin cytokines, such as thymic stromal lymphopoietin (TSLP), IL-25, and IL-33, with the development of a Th2-type immunological reaction [13]. Allergic diseases begin to appear frequently early in life and longitudinal studies have shown that they often manifest themselves in progression. The classic sequence (the so-called “atopic march”) from AD and food allergy (FA) to AA and AR (Figure 1) [14, 15], has not been recently highlighted in a large population of children [16]. AD generally represented the first manifestation of the atopic march although it does not necessarily evolve in following atopic diseases. Furthermore, only a few children who presented AD early in life developed asthma and AR until adolescence, and among those who presented all three atopic diseases, only a few developed them according to sequences known as the atopic march [16]. Cutaneous food allergen exposure in children with AD may promote immunoglobulin E (IgE)-mediated sensitization. Furthermore, Th2 memory cells, IgE, TSLP, IL-33, and IL-25 can reach the digestive and respiratory tracts through the blood circulation and lead up to the development of FA, AA, and AR if they encounter the allergen again (Figure 2) [15]. AD is very common in industrialized countries where it occurs in approximately 15–30% of children and 2–10% of adults [17–20]. Epidemiological studies have found that in 45% of affected children, AD appears in the first 6 months of age, in 60% within the first year, and up to 90% before the age of 5 years [14, 17, 21]. Tsakok and collaborators [22] in a systematic review reported that food sensitization was up to 6 times more frequent in patients with AD than those without AD. The early onset and severity of AD increase the risk of developing food sensitization, AA, and AR. Dharmage et al. [23] demonstrated that children with AD in the first two years of life had a significantly increased risk of developing AA and AR during 6–7 years. Moreover, many studies found that children with increased

severity of AD had an increased risk of developing FA, AA, and AR [22, 24, 25]. Overall, current evidence supports a multimorbidity framework for atopic diseases, without a specific sequential atopic march progression [16].

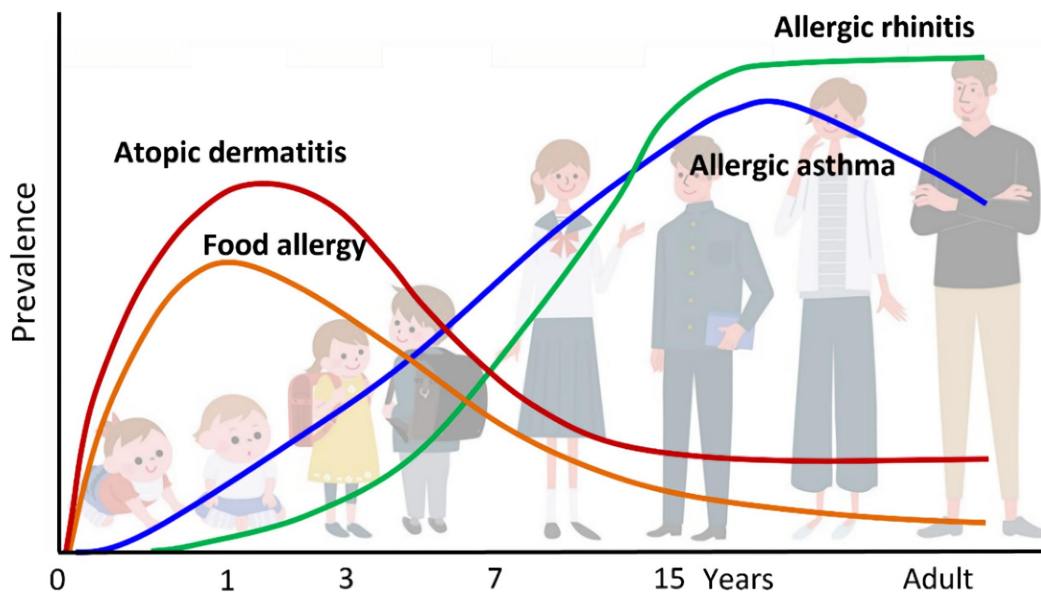


Figure 1. Classic atopic march. Atopic dermatitis (AD) generally occurs first, followed by immunoglobulin E (IgE)-mediated food allergy (FA), allergic asthma (AA), and allergic rhinitis (AR)

Note. Reprinted from “Current Insights into Atopic March” by Tsuge M, Ikeda M, Matsumoto N, Yorifuji T, Tsukahara H. *Children (Basel)*. 2021;8:1067 (<https://doi.org/10.3390/children8111067>). CC BY.

Routes of sensitization to food and airborne allergens

Sensitization to foods can occur not only through the gastrointestinal tract and oral cavity but also by different routes, such as the skin or the airways [26]. Sampson et al. [26] stated that even though oral tolerance is a usual response to swallowed antigens, a breakdown in this process apparently occurred in recent decades, resulting in a growing prevalence of sensitization to food allergens.

On the other hand, several studies demonstrated that breastmilk contains airborne allergens as well as food allergens derived from the maternal diet [27]. Inhalant allergens from *Dermatophagoides pteronyssinus* 1 (Der p 1) and *Blomia tropicalis* 5 (Blo t 5) have been detected in human milk [27]. Baiz and coworkers [28] demonstrated that children breast-fed by mothers with high Der p 1 levels had an increased risk of allergic sensitization and asthma and AR at five years of age. The mechanisms involved in natural history and therapeutically induced tolerance are thought to include suppression of Th2 cells, generation of T regulatory cells (Tregs), reduced production of IgE by B cells, augmented IgA and IgG4 production by B cells, defeat of effector T-cell migration to tissues, induction of IL-10-producing dendritic cells (DCs), and downfall of basophil, eosinophil, and mast cell activation [29]. Albeit with some variations, the same mechanisms which were reported to be accountable for the development of immune tolerance to allergens through different routes, such as Hymenoptera venom injected into the skin and pollens or dust mites inhaled into the respiratory tracts, are likely to be applicable in oral tolerance.

Sensitization to food allergens

From tolerance to sensitization in the gastrointestinal tract

The gastrointestinal tract is continuously exposed to a wide range of non-self-antigens including both commensal bacteria (microbiota) and food proteins [30]. Under physiological conditions, ingested foods evoke an immunological response of tolerance which is characterized by an active suppression of cellular and humoral immune responses towards an antigen with which it had already come into contact via the oral route [31]. The immunological mechanisms leading to the development of food tolerance are not fully

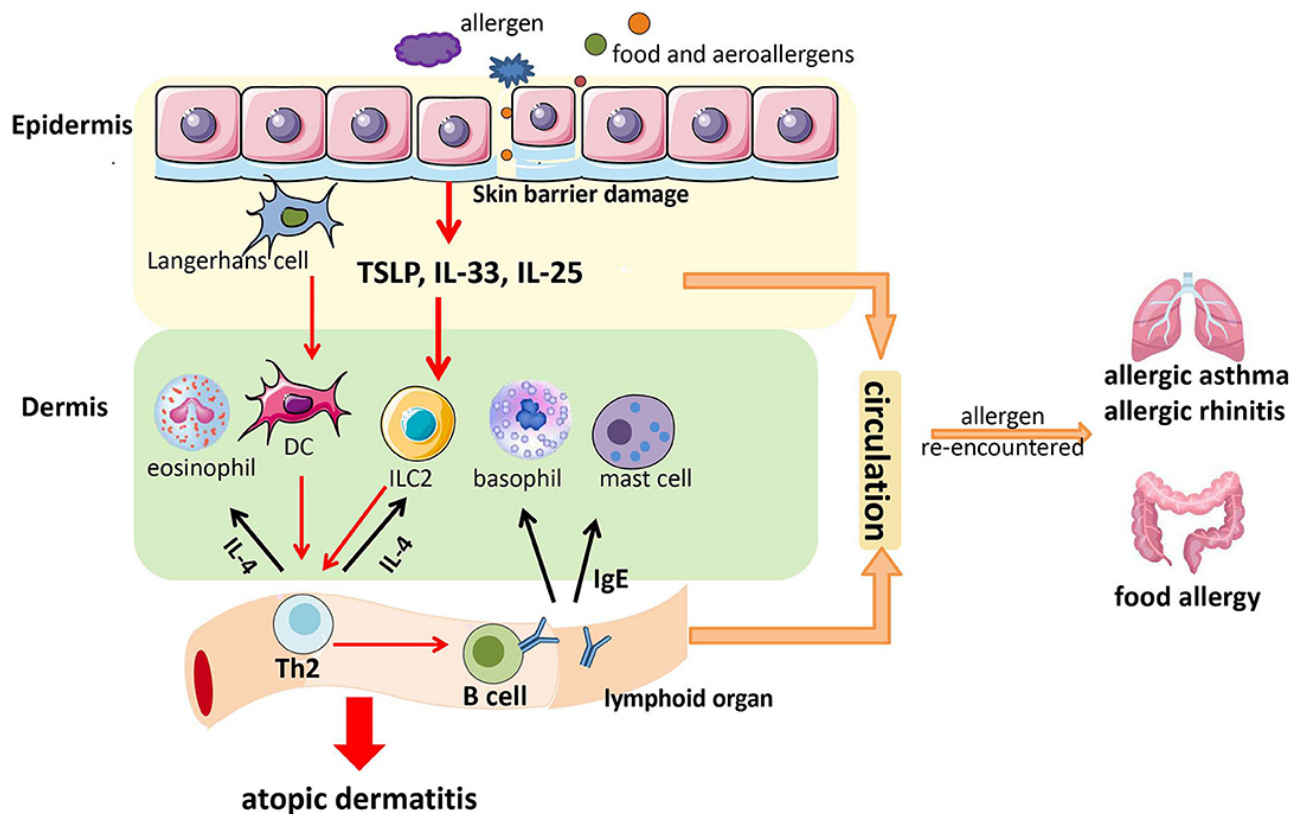


Figure 2. A possible mechanism through which food sensitization favors the development of food allergy (FA), allergic asthma (AA), and allergic rhinitis (AR). TSLP: thymic stromal lymphopoietin; DC: dendritic cell; ILC2: type 2 innate lymphoid cells; IgE: immunoglobulin E

Note. Reprinted from “Research Progress in Atopic March” by Yang L, Fu J, Zhou Y. *Front Immunol.* 2020;11:1907 (<https://doi.org/10.3389/fimmu.2020.01907>). CC BY.

understood and are mainly based on experimental mouse models. In principle, oral tolerance to food antigens is based on the effective production of Tregs within the intestinal mucosa [26].

After food ingestion, most of the food proteins are degraded into small peptides which, together with the remaining intact proteins, pass the intestinal barrier and are then presented to the immune cells [26]. The ways of overcoming the epithelial barrier include paracellular diffusion, endocytosis through microfold cells (M cells) and goblet cells, and the intervention of DCs which extend their dendrites into the intestinal lumen capturing food antigens [32]. Within the mucosa, food antigens are captured by antigen-presenting cells (APCs) which migrate to the intestinal draining mesenteric lymph nodes (mLNs; gut-draining mLNs) where cell differentiation occurs of naive T cells into Tregs [30]. Although Tregs specific to food allergens develop and localize in the intestine, they can also be found in the circulation where they maintain systemic tolerance [30].

The major mechanisms by which Tregs lead to tolerance to food antigens include at least the following: 1) encouraging B cells to produce non-inflammatory IgA-type responses, 2) suppressing Th2-type responses, and 3) inhibiting pro-inflammation, such as mast cells, basophils, and eosinophils leading to a state of non-responsiveness to ingested antigens or oral tolerance [30].

Whether food tolerance does not develop or if an existing tolerance is broken down, sensitization to food antigens can develop, which is characterized by a Th2 response with the production of specific IgE (sIgE) [33]. The mechanisms by which some subjects develop allergic sensitization towards some foods while the majority of subjects develop and maintain tolerance are not fully understood, but there is evidence that some factors can alter at least in part the normal balance existing between tolerance and allergy [33].

These adjuvant factors include gut microbiota status, intestinal pathogen infections, dietary factors, dose and timing of exposure to food allergens, and route of contact and penetration of the allergens [33].

For years it has been believed that FA develops exclusively in the intestine in children through breast milk or following an early introduction into the diet [34]. Du Toit et al. [35] demonstrated that in a large population of children at high risk of developing peanut allergy, the early introduction of peanuts into the diet was accompanied by a significantly lower prevalence of peanut allergy up to the age of 5 years. Furthermore, it has been highlighted that the preventive effect of FA with the early introduction of the food is allergen specific. In the Learning Early About Peanut Allergy (LEAP) study, it was found that the early introduction of peanuts had not led to the prevention of allergy to nuts or sesame seeds [36].

Transcutaneous food sensitization

Numerous studies have highlighted the growing importance of allergic sensitization through a dysfunction of the skin barrier. The skin, unlike the other epithelial surfaces, has a structure composed of two barriers: the stratum corneum (SC), which is exclusive to the skin, and the tight junctions (TJs) also present in the other epithelial surfaces [37]. Both the SC and the TJs moderate on the one hand the entry and reaction towards external agents such as microorganisms, allergenic proteins, irritants, and toxins, and on the other hand they prevent the outward leakage of water through the skin [transepidermal water loss (TEWL)]. Filaggrin (FLG), a barrier protein, plays an important role in the normal structural composition of the SC [15]. FLG loss-of function mutations have been frequently associated with AD which is characterized by barrier effect dysfunction and subsequent inflammatory lesions caused by stimulation of Th2 cells [38]. On the other hand, FLG developmental genetic mutations have not been detected in many patients with AD and over 50% of subjects with the mutation do not develop AD [39]. FLG genetic mutations increase the risk of severe AD and FA in individuals with AD [40, 41]. A strong association was found between AD, sensitization to food allergens, and FA [15].

The role of the FLG genetic defect in the development of FA was demonstrated by Brough et al. [42] in a population of almost 1,200 children. The authors measured the content of peanut antigens in house dust during the first year of life of the children and evaluated the development of sensitization by prick test and immuno-Cap towards Ara h 1, 2, and 3 at the age of 8 and 11 years. The conclusions were that environmental exposure to peanuts early in life is associated with an increased risk of sensitization to peanuts in children carrying the *FLG* gene mutation [42].

The prevalence of FA in patients with AD varies according to the time of onset and the severity of AD. In a population of more than 600 children, Hill and Hosking [43] demonstrated that as AD severity increased, adverse reactions to food increased. In the ALSoPAC study, the application of emollient creams containing peanut oil to the inflamed skin of children was found to be an independent risk factor for the development of allergic sensitization to peanuts [24].

Airway sensitization to food allergens

The dual allergen exposure hypothesis proposed by Lack et al. [24] suggests that food sensitization can occur through the impaired skin barrier. Kulis et al. [44] proposed that exposure to food allergens, such as peanuts, in the airway is a possible route of sensitization. Peanut protein has been found not only in the home environment but also in school cafeterias and classrooms, where children spend a lot of their time [45]. Inhalational exposure to peanut extract with house dust resulted in the production of peanut-IgE and anaphylaxis in consequence of the peanut challenge [46]. The authors reported that exposure to peanut extract or dust alone did not cause peanut sensitization.

Sensitization to inhalant allergens

Custovic [47] in a review reports that despite contact with inhalant allergens (e.g., dust mites, epidermal derivatives of pets, and pollens), which occurs under physiological conditions at the level of the mucous membranes of the respiratory tract through inhalation of the air environment, it is unknown which is the most relevant sensitization pathway for inhalant allergens. The epithelium of the respiratory tract represents the first defense barrier against inhaled particles—such as allergens, microorganisms, and

pollutants, preventing their diffusion within the respiratory mucosa. Most of the particles inhaled into the respiratory tract are trapped in the mucus layer covering the epithelium and then eliminated by mucociliary clearance. In addition to exercising the physical barrier function, the cells of the respiratory epithelium have a key role in the immunological, inflammatory, and repair responses following contact with environmental factors present in the inhaled air [48]. In longitudinal studies that have studied the temporal evolution of allergic diseases, it has been found that they tend to manifest themselves with a progressive sequence defined as atopic march. In this natural evolution, the first manifestation is represented by AD followed by the development of FA and subsequently of AA and AR. The dysfunction of the skin barrier present in AD favors the penetration of allergens and causes Th2-mediated sensitization and inflammation in some children. Mutation of the *FLG* gene is reported as the major risk factor for the development of AD. Although there is a clear relationship between AD and sensitization to food allergens by transcutaneous absorption, the relationship to inhalant allergens is controversial [47, 49]. Wörnberg Gerdin et al. [50] demonstrated that in a population of nearly 2,000 infants with skin changes such as eczema, dry skin on the cheeks and/or extensors, and a high TEWL at 3 months of age, development of sensitization to food allergens was seen at the age of 6 months in 8.9% while sensitization to inhalants occurred only in 1.9%. Chiu and collaborators [51] studied a cohort of children from the Prediction of Allergies in Taiwanese Children (PATCH) study with the aim of detecting the development of sensitization to the most common food and inhalant allergens from 0 to 4 years. The authors found that sensitization to food allergens began before inhalant allergens and increased markedly from the age of 6 months reaching the peak at 1.5 years of age. In contrast, the prevalence of sensitization to inhalant allergens was only 5% at 2 years but increased markedly after 2 years of age [51]. Miwa and collaborators [52] demonstrated that FLG is also expressed in the human nasal epithelium of the inferior turbinate, both in the stratified squamous epithelium in the anterior portion, and in the pseudostratified ciliated epithelium in the posterior portion, as well as in the maxillary sinus mucosa. Moreover, bacterial and viral infections can modify *FLG* gene expression in human nasal mucosa. We can hypothesize that, particularly in children where upper respiratory tract infections are more frequent, these may favor the development of sensitization to inhalant allergens by increasing the permeability of the nasal mucosa epithelium in association with a Th2-type immune response. Studies have revealed the presence of airway epithelial dysfunction in both asthma and upper respiratory tract disease due to disturbances in TJ formation [53]. On the other hand, it is also known that some environmental allergens, such as Der p and various pollens are endowed with cysteine and serine proteases which are able to break the bonds of TJs [54, 55]. Similarly, molds of the genus *Aspergillus* and *Penicillium* have serine protease activity. Moreover, the role of proteolytic activity has not been demonstrated for human disease [54, 56]. In addition to the direct effect of inhalant allergens, allergic sensitization can lead to epithelial barrier dysfunction as a consequence of asthma-associated airway inflammation mediated by a Th2-type response [56]. This alteration can be caused by the intervention of both Th2 cells and type 2 innate lymphoid cells (ILC2) through the secretion of IL-13 which, in addition, causes other features of the respiratory epithelium in asthma, such as mucus production [57]. Furthermore, the destruction of barrier TJs can be induced by environmental pollutants such as particulate matter 2.5 and cigarette smoke [58].

New approaches for the diagnosis of allergy

The development of new insights into the pathomechanisms of allergies, and in particular the role of epithelial barriers, has stimulated new research fields. As we reported above, epithelial barriers represent the first line of defense against foreign substances, preventing the entry of potentially harmful substances through the epithelium. In the case of epithelial barrier dysfunction, allergens can penetrate through the epithelium followed by the release of cytokines (such as TSLP, IL-33, and IL-25) that cause type 2 immune responses. Recently, the first report was published on sensitization to cat allergen through cat bite in a child [59]. In this case report, sIgE assays showed positive results for cat extracts and rFel d 1. Identifying a new route of allergic sensitization to cat dander after decades of studies may appear unlikely. However, it must be considered that asking for information on possible cat bites is not included in the anamnesis. Previously,

Maeda and Akiyama [60] described a case of anaphylaxis after a cat bite in a 42-year-old woman already sensitized to cat allergen. These clinical reports consolidate the assumption that a cat bite could contain enough allergen to both trigger symptoms and develop possible sensitization. The intestinal epithelium has an important barrier function as on the one hand, it has to defend against microorganisms and toxins and on the other it has to exchange nutrients, water, and electrolytes [61]. Ogulur and collaborators [62] investigated if professional and household dishwashers and rinse aid agents may have an effect on gut barrier dysfunction. The authors showed no toxic effects after exposure to professional dishwasher detergent at all dilutions down to 1:250. In contrast, rinse aids caused cell death and impairment of epithelial barrier epithelium by destroying the TJs [62]. Baur and collaborators [63] demonstrated that exposure to triclosan on reconstructed human epidermis modifies keratinocyte function and increases skin barrier permeability.

A recent study showed that low levels of zinc in erythrocytes are associated with the development of allergic proctocolitis in children [64]. Zinc is an important trace element in the early stages of life both for the progress of the barrier function of the epithelia and of the immune system [65]. Previous studies had found a relationship between low zinc values and allergic diseases in children, such as AD and asthma, but the relationship with allergic proctocolitis had not been reported [66, 67]. Gunaydin et al. [64] found in a population of children with a mean age of (7.1 ± 2.9) months affected by allergic proctocolitis, intracellular zinc values were significantly lower than those in a control group of the same number and mean age.

Microbiome

Many studies have suggested that the increase in allergic diseases in recent decades is due to an alteration of the intestinal microbiota resulting from better hygienic and sanitary conditions, more frequent cesarean sections and use of antibiotics, and a diet low in fiber and high in fat and sugar. Moreover, Roudit et al. [68] demonstrated that strategies to increase short-chain fatty acid (SCFA) levels, particularly butyrate and propionate, in the first year of life are associated with a lower risk of developing FA, asthma, and AR between 3 and 6 years of life. These data suggest that prebiotics and postbiotics in the first year of life may help prevent the development of allergic diseases [69].

Conclusions

Understanding how sensitizations to food and inhalant allergens occur is an important prerequisite in order to be able to implement prevention strategies [70].

Skin barrier dysfunction in AD is a risk factor for developing FA and a possible onset of the atopic march. More data are also needed to better understand the role of microbiome and nutrition in order to provide specific and targeted preventive and therapeutic measures also in this context whether the rationale and evidence will be supported. In the near future, new immunological and cohort studies are needed to better understand the pathomechanism of allergic diseases and implement proper preventive and therapeutic strategies at a population and individual level.

Abbreviations

AA: allergic asthma

AD: atopic dermatitis

AR: allergic rhinitis

FA: food allergy

FLG: filaggrin

IgE: immunoglobulin E

SC: stratum corneum

sIgE: specific immunoglobulin E

TJs: tight junctions

Treg: T regulatory cell

TSLP: thymic stromal lymphopoietin

Declarations

Acknowledgments

The authors thank Stanley Norman for the English revision.

Author contributions

GL, CI, and SA: Writing—original draft, Writing—review & editing. All authors have read and agreed to the final version of the manuscript.

Conflicts of interest

The authors declare that they have no conflicts of interest related to this manuscript.

Ethical approval

Not applicable.

Consent to participate

Not applicable.

Consent to publication

Not applicable.

Availability of data and materials

Not applicable.

Funding

Not applicable.

Copyright

© The Author(s) 2024.

References

1. Bergmann KC, Ring J, editors. History of allergy. Karger Medical and Scientific Publishers; 2014.
2. Vierucci A. Allergie: breve storia e pietre miliari. RIAP. 2019;2–14. Italian.
3. Ramachandran M, Aronson JK. John Bostock's first description of hayfever. J R Soc Med. 2011;104: 237–40. [DOI] [PubMed] [PMC]
4. Spieksma FTM, Dieges PH. The history of the finding of the house dust mite. J Allergy Clin Immunol. 2004;113:573–6. [DOI] [PubMed]
5. allergen.org [Internet]. [cited 2024 Jan 4]. Available from: <https://www.allergen.org/>
6. allergenonline.org [Internet]. University of Nebraska–Lincoln; c2019 [cited 2024 Jan 4]. Available from: <http://www.allergenonline.org/>
7. allergome.org [Internet]. Allergome; c2003–2015 [cited 2024 Jan 4]. Available from: <https://www.allergome.org/>
8. Ring J. History of Allergy: Clinical Descriptions, Pathophysiology, and Treatment. In: Traidl-Hoffmann C, Zuberbier T, Werfel T, editors. Allergic Diseases – From Basic Mechanisms to Comprehensive Management and Prevention. Cham: Springer International Publishing; 2022. pp. 3–19. [DOI] [PubMed]

9. Akdis CA. Does the epithelial barrier hypothesis explain the increase in allergy, autoimmunity and other chronic conditions? *Nat Rev Immunol*. 2021;21:739–51. [DOI] [PubMed]
10. Prescott S, Allen KJ. Food allergy: riding the second wave of the allergy epidemic. *Pediatr Allergy Immunol*. 2011;22:155–60. [DOI] [PubMed]
11. Strachan DP. Hay fever, hygiene, and household size. *BMJ*. 1989;299:1259–60. [DOI] [PubMed] [PMC]
12. Lynch SV, Boushey HA. The microbiome and development of allergic disease. *Curr Opin Allergy Clin Immunol*. 2016;16:165–71. [DOI] [PubMed] [PMC]
13. Goleva E, Berdyshev E, Leung DY. Epithelial barrier repair and prevention of allergy. *J Clin Invest*. 2019;129:1463–74. [DOI] [PubMed] [PMC]
14. Tsuge M, Ikeda M, Matsumoto N, Yorifuji T, Tsukahara H. Current Insights into Atopic March. *Children (Basel)*. 2021;8:1067. [DOI] [PubMed] [PMC]
15. Yang L, Fu J, Zhou Y. Research Progress in Atopic March. *Front Immunol*. 2020;11:1907. [DOI] [PubMed] [PMC]
16. Haider S, Fontanella S, Ullah A, Turner S, Simpson A, Roberts G, et al.; STELAR/UNICORN investigators. Evolution of Eczema, Wheeze, and Rhinitis from Infancy to Early Adulthood: Four Birth Cohort Studies. *Am J Respir Crit Care Med*. 2022;206:950–60. [DOI] [PubMed] [PMC]
17. Bieber T. Atopic dermatitis. *N Engl J Med*. 2008;358:1483–94. [DOI] [PubMed]
18. Odhiambo JA, Williams HC, Clayton TO, Robertson CF, Asher MI; ISAAC Phase Three Study Group. Global variations in prevalence of eczema symptoms in children from ISAAC Phase Three. *J Allergy Clin Immunol*. 2009;124:1251–8.e23. [DOI] [PubMed]
19. Silverberg JI, Barbarot S, Gadkari A, Simpson EL, Weidinger S, Mina-Osorio P, et al. Atopic dermatitis in the pediatric population: A cross-sectional, international epidemiologic study. *Ann Allergy Asthma Immunol*. 2021;126:417–28.e2. [DOI] [PubMed]
20. Langan SM, Irvine AD, Weidinger S. Atopic dermatitis. *Lancet*. 2020;396:345–60. [DOI] [PubMed]
21. Weidinger S, Novak N. Atopic dermatitis. *Lancet*. 2016;387:1109–22. [DOI] [PubMed]
22. Tsakok T, Marrs T, Mohsin M, Baron S, du Toit G, Till S, et al. Does atopic dermatitis cause food allergy? A systematic review. *J Allergy Clin Immunol*. 2016;137:1071–8. [DOI] [PubMed]
23. Dharmage SC, Lowe AJ, Matheson MC, Burgess JA, Allen KJ, Abramson MJ. Atopic dermatitis and the atopic march revisited. *Allergy*. 2014;69:17–27. [DOI] [PubMed]
24. Lack G, Fox D, Northstone K, Golding J; Avon Longitudinal Study of Parents and Children Study Team. Factors associated with the development of peanut allergy in childhood. *N Engl J Med*. 2003;348:977–85. [DOI] [PubMed]
25. Zheng T, Yu J, Oh MH, Zhu Z. The atopic march: progression from atopic dermatitis to allergic rhinitis and asthma. *Allergy Asthma Immunol Res*. 2011;3:67–73. [DOI] [PubMed] [PMC]
26. Sampson HA, O'Mahony L, Burks AW, Plaut M, Lack G, Akdis CA. Mechanisms of food allergy. *J Allergy Clin Immunol*. 2018;141:11–9. [DOI] [PubMed]
27. Macchiaverni P, Rekima A, van den Elsen L, Renz H, Verhasselt V. Allergen shedding in human milk: Could it be key for immune system education and allergy prevention? *J Allergy Clin Immunol*. 2021;148:679–88. [DOI] [PubMed]
28. Baiz N, Macchiaverni P, Tulic MK, Rekima A, Annesi-Maesano I, Verhasselt V; EDEN Mother-Child Cohort Study Group. Early oral exposure to house dust mite allergen through breast milk: A potential risk factor for allergic sensitization and respiratory allergies in children. *J Allergy Clin Immunol*. 2017;139:369–72.e10. [DOI] [PubMed]
29. Akdis CA, Akdis M. Advances in allergen immunotherapy: aiming for complete tolerance to allergens. *Sci Transl Med*. 2015;7:280ps6. [DOI] [PubMed]
30. Wambre E, Jeong D. Oral Tolerance Development and Maintenance. *Immunol Allergy Clin North Am*. 2018;38:27–37. [DOI] [PubMed] [PMC]

31. Chehade M, Mayer L. Oral tolerance and its relation to food hypersensitivities. *J Allergy Clin Immunol.* 2005;115:3–12. [DOI] [PubMed]
32. Ali A, Tan H, Kaiko GE. Role of the Intestinal Epithelium and Its Interaction With the Microbiota in Food Allergy. *Front Immunol.* 2020;11:604054. [DOI] [PubMed] [PMC]
33. Pelz BJ, Bryce PJ. Pathophysiology of Food Allergy. *Pediatr Clin North Am.* 2015;62:1363–75. [DOI] [PubMed]
34. Committee on Nutrition. Hypoallergenic infant formulas. *Pediatrics.* 2000;106:346–9. [DOI] [PubMed]
35. Du Toit G, Roberts G, Sayre PH, Bahnson HT, Radulovic S, Santos AF, et al.; LEAP Study Team. Randomized trial of peanut consumption in infants at risk for peanut allergy. *N Engl J Med.* 2015;372:803–13. [DOI] [PubMed] [PMC]
36. du Toit G, Sayre PH, Roberts G, Lawson K, Sever ML, Bahnson HT, et al.; Immune Tolerance Network Learning Early About Peanut Allergy study team. Allergen specificity of early peanut consumption and effect on development of allergic disease in the Learning Early About Peanut Allergy study cohort. *J Allergy Clin Immunol.* 2018;141:1343–53. [DOI] [PubMed] [PMC]
37. Beck LA, Cork MJ, Amagai M, De Benedetto A, Kabashima K, Hamilton JD, et al. Type 2 Inflammation Contributes to Skin Barrier Dysfunction in Atopic Dermatitis. *JID Innov.* 2022;2:100131. [DOI] [PubMed] [PMC]
38. Brough HA, Nadeau KC, Sindher SB, Alkotob SS, Chan S, Bahnson HT, et al. Epicutaneous sensitization in the development of food allergy: What is the evidence and how can this be prevented? *Allergy.* 2020;75:2185–205. [DOI] [PubMed] [PMC]
39. Rodríguez E, Baurecht H, Herberich E, Wagenpfeil S, Brown SJ, Cordell HJ, et al. Meta-analysis of filaggrin polymorphisms in eczema and asthma: robust risk factors in atopic disease. *J Allergy Clin Immunol.* 2009;123:1361–70.e7. [DOI] [PubMed]
40. Palmer CNA, Irvine AD, Terron-Kwiatkowski A, Zhao Y, Liao H, Lee SP, et al. Common loss-of-function variants of the epidermal barrier protein filaggrin are a major predisposing factor for atopic dermatitis. *Nat Genet.* 2006;38:441–6. [DOI] [PubMed]
41. Hudson TJ. Skin barrier function and allergic risk. *Nat Genet.* 2006;38:399–400. [DOI] [PubMed]
42. Brough HA, Simpson A, Makinson K, Hankinson J, Brown S, Douiri A, et al. Peanut allergy: effect of environmental peanut exposure in children with filaggrin loss-of-function mutations. *J Allergy Clin Immunol.* 2014;134:867–75.e1. [DOI] [PubMed] [PMC]
43. Hill DJ, Hosking CS. Food allergy and atopic dermatitis in infancy: an epidemiologic study. *Pediatr Allergy Immunol.* 2004;15:421–7. [DOI] [PubMed]
44. Kulis MD, Smeekens JM, Immormino RM, Moran TP. The airway as a route of sensitization to peanut: An update to the dual allergen exposure hypothesis. *J Allergy Clin Immunol.* 2021;148:689–93. [DOI] [PubMed] [PMC]
45. Sheehan WJ, Brough HA, Makinson K, Petty CR, Lack G, Phipatanakul W. Distribution of peanut protein in school and home environments of inner-city children. *J Allergy Clin Immunol.* 2017;140:1724–6. [DOI] [PubMed] [PMC]
46. Smeekens JM, Immormino RM, Balogh PA, Randell SH, Kulis MD, Moran TP. Indoor dust acts as an adjuvant to promote sensitization to peanut through the airway. *Clin Exp Allergy.* 2019;49:1500–11. [DOI] [PubMed] [PMC]
47. Custovic A. To what extent is allergen exposure a risk factor for the development of allergic disease? *Clin Exp Allergy.* 2015;45:54–62. [DOI] [PubMed]
48. Celebi Sözen Z, Cevhertas L, Nadeau K, Akdis M, Akdis CA. Environmental factors in epithelial barrier dysfunction. *J Allergy Clin Immunol.* 2020;145:1517–28. [DOI] [PubMed]
49. Leung DYM, Berdyshev E, Goleva E. Cutaneous barrier dysfunction in allergic diseases. *J Allergy Clin Immunol.* 2020;145:1485–97. [DOI] [PubMed] [PMC]
50. Wärnberg Gerdin S, Lie A, Asarnoj A, Borres MP, Lødrup Carlsen KC, Färdig M, et al. Impaired skin barrier and allergic sensitization in early infancy. *Allergy.* 2022;77:1464–76. [DOI] [PubMed]

51. Chiu CY, Huang YL, Tsai MH, Tu YL, Hua MC, Yao TC, et al. Sensitization to food and inhalant allergens in relation to atopic diseases in early childhood: a birth cohort study. *PLoS One*. 2014;9:e102809. [DOI] [PubMed] [PMC]
52. Miwa M, Hasan S, Miwa M, Okubo K. Filaggrin exists in human nose. *Allergol Int*. 2016;65:338–40. [DOI] [PubMed]
53. Hellings PW, Steelant B. Epithelial barriers in allergy and asthma. *J Allergy Clin Immunol*. 2020;145:1499–509. [DOI] [PubMed] [PMC]
54. Reithofer M, Jahn-Schmid B. Allergens with Protease Activity from House Dust Mites. *Int J Mol Sci*. 2017;18:1368. [DOI] [PubMed] [PMC]
55. Celebi Sozener Z, Ozdel Ozturk B, Cerci P, Turk M, Gorgulu Akin B, Akdis M, et al. Epithelial barrier hypothesis: Effect of the external exposome on the microbiome and epithelial barriers in allergic disease. *Allergy*. 2022;77:1418–49. [DOI] [PubMed] [PMC]
56. Heijink IH, Kuchibhotla VNS, Roffel MP, Maes T, Knight DA, Sayers I, et al. Epithelial cell dysfunction, a major driver of asthma development. *Allergy*. 2020;75:1902–17. [DOI] [PubMed] [PMC]
57. Sugita K, Steer CA, Martinez-Gonzalez I, Altunbulakli C, Morita H, Castro-Giner F, et al. Type 2 innate lymphoid cells disrupt bronchial epithelial barrier integrity by targeting tight junctions through IL-13 in asthmatic patients. *J Allergy Clin Immunol*. 2018;141:300–10.e11. [DOI] [PubMed]
58. Fukuoka A, Yoshimoto T. Barrier dysfunction in the nasal allergy. *Allergol Int*. 2018;67:18–23. [DOI] [PubMed]
59. Leo G, Incorvaia C, Arasi S. Could a bite trigger the onset of cat allergy? *Pediatr Allergy Immunol*. 2022;33:e13841. [DOI] [PubMed] [PMC]
60. Maeda Y, Akiyama K. Anaphylaxis after a cat bite. *Allergol Int*. 2012;61:511–2. [DOI] [PubMed]
61. Groschwitz KR, Hogan SP. Intestinal barrier function: molecular regulation and disease pathogenesis. *J Allergy Clin Immunol*. 2009;124:3–20. [DOI] [PubMed] [PMC]
62. Ogulur I, Pat Y, Aydin T, Yazici D, Rückert B, Peng Y, et al. Gut epithelial barrier damage caused by dishwasher detergents and rinse aids. *J Allergy Clin Immunol*. 2023;151:469–84. [DOI] [PubMed]
63. Baur R, Kashon M, Lukomska E, Weatherly LM, Shane HL, Anderson SE. Exposure to the anti-microbial chemical triclosan disrupts keratinocyte function and skin integrity in a model of reconstructed human epidermis. *J Immunotoxicol*. 2023;20:2148781. [DOI] [PubMed] [PMC]
64. Gunaydin NC, Celikkol A, Nalbantoglu A. Assessment of intracellular zinc levels in infants with food protein-induced allergic proctocolitis. *Allergol Immunopathol (Madr)*. 2023;51:9–15. [DOI] [PubMed]
65. Maywald M, Rink L. Zinc homeostasis and immunosenescence. *J Trace Elem Med Biol*. 2015;29:24–30. [DOI] [PubMed]
66. Karabacak E, Aydin E, Kutlu A, Ozcan O, Muftuoglu T, Gunes A, et al. Erythrocyte zinc level in patients with atopic dermatitis and its relation to SCORAD index. *Postepy Dermatol Alergol*. 2016;33:349–52. [DOI] [PubMed] [PMC]
67. Podlecka D, Jerzyńska J, Sanad K, Polańska K, Bobrowska-Korzeniowska M, Stelmach I, et al. Micronutrients and the Risk of Allergic Diseases in School Children. *Int J Environ Res Public Health*. 2022;19:12187. [DOI] [PubMed] [PMC]
68. Roduit C, Frei R, Ferstl R, Loeliger S, Westermann P, Rhyner C, et al.; PASTURE/EFRAIM study group. High levels of butyrate and propionate in early life are associated with protection against atopy. *Allergy*. 2019;74:799–809. [DOI] [PubMed]
69. Akagawa S, Kaneko K. Gut microbiota and allergic diseases in children. *Allergol Int*. 2022;71:301–9. [DOI] [PubMed]
70. Perkin MR, Togias A, Koplin J, Sicherer S. Food Allergy Prevention: More Than Peanut. *J Allergy Clin Immunol Pract*. 2020;8:1–13. [DOI] [PubMed]