

## **Open Access** Original Article



# Preparation and characterization of stearyl glycyrrhetinate/cyclodextrin complex using co-grinding

Momoko Ebisawa<sup>1</sup>, Nao Kodama<sup>1</sup><sup>®</sup>, Shun-ichi Mitomo<sup>1</sup><sup>®</sup>, Junki Tomita<sup>2</sup>, Mitsuaki Suzuki<sup>3</sup><sup>®</sup>, Yutaka Inoue<sup>1\*</sup><sup>®</sup>

<sup>1</sup>Laboratory of Nutri-Pharmacotherapeutics Management, Faculty of Pharmacy and Pharmaceutical Sciences, Josai University, Sakado 3500295, Japan

<sup>2</sup>Instrument Analysis Center, Josai University, Sakado 3500295, Japan

<sup>3</sup>Department of Chemistry, Faculty of Science, Josai University, Sakado 3500295, Japan

\*Correspondence: Yutaka Inoue, Laboratory of Nutri-Pharmacotherapeutics Management, Faculty of Pharmacy and Pharmaceutical Sciences, Josai University, 1-1 Keyakidai, Sakado, Saitama 3500295, Japan. yinoue@josai.ac.jp Academic Editor: Jayachandran Venkatesan, Yenepoya University, India Received: September 4, 2024 Accepted: February 10, 2025 Published: February 26, 2025

**Cite this article:** Ebisawa M, Kodama N, Mitomo Si, Tomita J, Suzuki M, Inoue Y. Preparation and characterization of stearyl glycyrrhetinate/cyclodextrin complex using co-grinding. Explor BioMat-X. 2025;2:101330. https://doi.org/10.37349/ebmx. 2025.101330

# Abstract

**Aim:** In this study, the physicochemical properties of stearyl glycyrrhetinate/ $\beta$ -cyclodextrin (SG/ $\beta$ CD) and SG/ $\gamma$ CD complexes were characterized, and the complexes were prepared using the co-milling method. The molecular interactions within the SG/CD complexes were also investigated using nuclear magnetic resonance (NMR) measurements to determine the mode of interaction.

**Methods:** Here, we evaluated the physicochemical properties of SG complexes with CDs prepared by ground mixtures (GM SG/ $\beta$ CD or  $\gamma$ CD = 1/1, 1/2).

**Results:** Powder X-ray diffraction (PXRD) showed that the characteristic SG and CD peaks disappeared after co-grinding with GM (SG/CD = molar ratio of 1/2), indicating a halo pattern. Differential scanning calorimetry (DSC) measurements showed that after co-grinding, the endothermic peak due to SG melting, as well as the dehydration peak and the endothermic peak due to SG melting, disappeared. Near-infrared (NIR) spectroscopy results showed that the peaks of SG-derived –CH moieties and CD-derived –OH and –CH moieties broadened in GM, suggesting their involvement in complex formation through SG/CDs intermolecular interactions. In GM (SG/CDs), NMR measurements showed broadened H-A and H-F peaks of the steroid skeleton derived from SG. In GM (SG/ $\beta$ CD = 1/2), the doublet peak, especially OH-3 at the wide edge of CD, shifted to a singlet peak. In GM (SG/ $\gamma$ CD = 1/2), the H-3 peak, which is the wide edge of  $\gamma$ CD, and the H-6 peak, which is the narrow edge, shifted to broad peaks, suggesting that  $\gamma$ CD is deeply encapsulated in the steroidal structure.

**Conclusions:** These findings suggest that complex formation occurred in SG/CDs and that inclusion behavior is different between GM (SG/ $\beta$ CD = 1/2) and GM (SG/ $\gamma$ CD = 1/2).

© The Author(s) 2025. This is an Open Access article licensed under a Creative Commons Attribution 4.0 International License (https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, sharing, adaptation, distribution and reproduction in any medium or format, for any purpose, even commercially, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.



# **Keywords**

Stearyl glycyrrhetinate, cyclodextrin, co-grinding, inclusion, nuclear magnetic resonance

## Introduction

The epidemic of new coronavirus infection has made wearing masks a habit, leading to an increasing number of people experiencing skin problems such as mask-induced steaming, acne, and pimples [1]. Skincare products are often used as a countermeasure against skin problems, and a few of these products contain ingredients that prevent skin irritation. One example is stearyl glycyrrhetinate (SG), a derivative of 18- $\beta$ -glycyrrhetinic acid (GA), a major metabolite of glycyrrhizic acid (GL) found in licorice root [2, 3]. SG is a dehydration-condensed ester of the hydroxyl group of stearyl alcohol on the carboxyl group of GL. SG is more lipophilic than dipotassium glycyrrhizinate [4], another derivative of GL, and is used in cosmetics such as creams and emulsions [5]. It has anti-inflammatory, skin-soothing, and antioxidant properties [6]. However, the physical properties of SG render them poorly soluble in water, necessitating a formulation that improves its solubility.

Cyclodextrin (CD) is a cyclic oligosaccharide composed of  $\alpha$ -1,4-linked D-glucopyranose with a conical hydrophobic inner cavity and a hydrophilic outer surface. The natural CD varieties include those with at least six glucose units, with the most common CDs those that present six ( $\alpha$ CD), seven ( $\beta$ CD), and eight units ( $\gamma$ CD) [7]. Their compounds, cyclodextrins ( $\alpha$ -,  $\beta$ -, and  $\gamma$ -CDs), are "generally recognized as safe" by the U.S. Food and Drug Administration (FDA), according to Braga et al. [8]. For these advantages in drug delivery, various CDs, including  $\alpha$ -,  $\beta$ -, and  $\gamma$ -CDs, have already been registered as pharmaceutical excipients. The cavities of CDs are slightly hydrophobic and can incorporate lipophilic drugs and moieties. CDs have a three-dimensional shape like a hollow torus and exhibit different polarities on their inner and outer surfaces [9]. These primary and secondary hydroxyl moieties are located on the narrow and wide sides, respectively. In this sense, the primary hydroxyl moieties can rotate and reduce the diameter of CDs, whereas the secondary hydroxyl moieties form strong hydrogen bonds and give the CDs rigidity [10]. Due to their special structures, CDs can encapsulate guest molecules and form inclusion complexes. The inclusion complexation of hydrophobic compounds primarily occurs through hydrophobic interactions between the CD cavity wall and guest molecules. Other forces, such as dipole-dipole interactions and van der Waals interactions, may also contribute to guest binding [11]. Complex formation improves the water solubility of drugs, increases chemical stability [12], reduces or prevents irritation, and masks unpleasant tastes and odors [13]. Our laboratory has reported that the formation of a complex between the pentacyclic triterpenoid ursolic acid and  $\gamma$ CD enhanced solubility [14]. The inclusion complexes were prepared using a mechanochemical mixture.

Current studies suggest that SG/CD inclusion complexes can be prepared by co-grinding SG with CDs and that the physicochemical properties of SG may be modified, leading to the functional improvement of SG. This contributes to the anti-inflammatory effect of SG on mask rash and rough skin with improved formulation handling. In this study, it is proposed that co-grinding SG with CD to modify its physicochemical properties and prepare SG/CD complexes could expand the potential for improving SG functionality. Specifically, this approach may enhance the handling properties of SG formulations and further contribute to the anti-inflammatory effects of SG against mask-induced skin irritation and other skin problems. Therefore, in this study, SG/ $\beta$ CD and SG/ $\gamma$ CD were co-grinding to form complexes with poorly soluble drugs. Then, SG/CDs complexes were prepared and the intermolecular interactions in the solid state were investigated by nuclear magnetic resonance (NMR) measurements to evaluate the physical properties and the mode of complex formation.

# **Materials and methods**

## Materials

The SG was purchased from Tokyo Chemical Industry Co., Ltd. (Lot. TUWLL-ID, Tokyo, Japan).  $\beta$ CD and  $\gamma$ CD were donated by CycloChem Bio Co., Ltd. (Tokyo, Japan) and stored at 40°C and 82% relative humidity for 7 days (Figure 1). NMR solvents were purchased from Fujifilm Wako Pure Chemical Corporation (Tokyo, Japan). Other reagents were purchased from Fujifilm Wako Pure Chemical Corporation.



Figure 1. Chemical structure of (a) SG, (b)  $\beta$ CD, and (c)  $\gamma$ CD. SG: stearyl glycyrrhetinate; CD: cyclodextrin

#### Sample preparation

Physical mixtures (PM) were prepared by weighing SG and  $\beta$ CD or  $\gamma$ CD in a molar ratio of 1/1 or 1/2, respectively, and mixing them in a vortex mixer for 1 min. Ground mixtures (GM) were prepared by weighing out a total of 500 mg of PM (SG/ $\beta$ CD, SG/ $\gamma$ CD) and grinding it in a vibrating rod mill (TI-500ET, CMT Co., Ltd., Tokyo, Japan) for 30 min.

## Powder X-ray diffraction (PXRD) measurement

Diffraction intensity was measured with a NaI scintillation counter using a Miniflex II powder X-ray diffraction measuring device (Rigaku Corporation, Tokyo, Japan). The measurement conditions used were Cu radiation (30 kV, 15 mA), a scan condition of  $4^{\circ}$ /min for the X-ray diffraction measurement, and a measurement range of  $2\theta = 5-40^{\circ}$ . The measurements were performed by filling a powdered sample into a glass plate; therefore, the sample plane was flat.

## Differential scanning calorimetry (DSC) measurements

Measurements were performed using a high-sensitivity differential scanning calorimeter (Thermo Plus Evo; Rigaku Corporation, Tokyo, Japan). A sample weighing 1.5–2 mg was precisely measured and placed in a sealed aluminum pan for analysis. The measurement conditions were nitrogen gas flow rate of 60 mL/min, temperature increase rate of 10.0 °C/min, and measurement range of 30–350°C.

#### Near-infrared (NIR) absorption spectrum measurement

Measurements were performed using a Fourier-transform near-infrared spectrometer (JASCO V-770; JASCO Corporation, Tokyo, Japan). Approximately 3 mg of each sample was placed in a powder cell for measurement, and measurements were performed at 5 nm intervals along the optical path. Measurement conditions were a measurement range of 4,000–10,000 cm<sup>-1</sup>, measurement time of 8 s, and measurement temperature of 25°C. The obtained spectra were subjected to second-order differentiation and analysis.

#### Scanning electron microscopy (SEM)

SEM measurements were performed using a JSM-IT800 (SHL) instrument (JEOL, Tokyo, Japan) at an accelerating voltage of 1 kV. The vacuum gold steaming time of the samples was 60 s.

#### <sup>1</sup>H-NMR spectrum measurement

NMR spectroscopy was performed using an AVENCE NEO 600 cryosystem (Bruker, Yokohama, Japan) operating at 600.13 MHz in DMSO-d<sub>6</sub>. The other conditions were as follows: 90° pulse, 7.775  $\mu$ s; waiting time, 1 s; integration frequency, 64; and temperature, 25°C. Note: SG is a pentacyclic triterpenoid with low solubility in aqueous media. Therefore, SG was first dissolved in DMSO-d<sub>6</sub>, and then diluted in aqueous media to be used in the NMR assay [15].

## **Results**

#### **PXRD** measurement

The crystalline state of the SG/CDs (molar ratio of 1/1, 1/2) prepared by co-grinding was confirmed via PXRD measurements. Characteristic diffraction peaks were observed for intact SG at  $2\theta = 6.7^{\circ}$ ,  $10.3^{\circ}$ ,  $13.9^{\circ}$ , and  $17.5^{\circ}$  (Figure 2a);  $\beta$ CD  $2\theta = 8.8^{\circ}$  and  $12.5^{\circ}$  (Figure 2b); and  $\gamma$ CD  $2\theta = 12.2^{\circ}$ ,  $13.8^{\circ}$ , and  $16.3^{\circ}$  (Figure 2g). Characteristic diffraction peaks derived from SG and CDs were observed for PM [SG/CDs (molar ratio of 1/1, 1/2)] (Figure 2c, d, h, i). In GM (SG/ $\beta$ CD = 1/1) and GM (SG/ $\gamma$ CD = 1/1), the diffraction peaks derived from SG did not disappear (Figure 2e, j). In GM (SG/ $\beta$ CD = 1/2) and GM (SG/ $\gamma$ CD = 1/2), the characteristic diffraction peak derived from SG disappeared. Furthermore, the diffraction peaks derived from  $\beta$ CD and  $\gamma$ CD disappeared, and a halo pattern was exhibited (Figure 2f, k).

#### **DSC analysis**

The PXRD analysis results confirmed a halo pattern for GM (SG/CD = 1/2) in the solid state. Therefore, DSC analysis was carried out to determine the thermal behavior of GM (SG/ $\beta$ CD, SG/ $\gamma$ CD). An endothermic peak due to the melting point of SG was observed at approximately 80°C for intact SG (Figure 3a). In  $\beta$ CD, an endothermic peak due to dehydration was observed at approximately 109°C, and a peak due to decomposition of  $\beta$ CD was observed at approximately 310°C (Figure 3b). In  $\gamma$ CD, an endothermic peak due to dehydration was observed at approximately 310°C (Figure 3g). In PM (SG/ $\beta$ CD = 1/1, 1/2, SG/ $\gamma$ CD = 1/1, 1/2) and GM (SG/ $\beta$ CD = 1/1, SG/ $\gamma$ CD = 1/1), an endothermic peak due to the melting point of SG was observed at approximately 72° C (Figure 3c, d, e, h, i, j). Above 300°C, a peak of decomposition due to  $\beta$ CD or  $\gamma$ CD was observed. In contrast, the disappearance of the endothermic peak due to SG was observed in GM (SG/ $\beta$ CD = 1/2, SG/ $\gamma$ CD = 1/2) (Figure 3f, k).

#### **NIR** analysis

The results of PXRD and DSC indicated that GM (SG/ $\beta$ CD = 1/2) and GM (SG/ $\gamma$ CD = 1/2) in the solid state formed inclusion complexes at a molar ratio of 1/2, respectively. To confirm the intermolecular interactions between SG/ $\beta$ CD and SG/ $\gamma$ CD in the solid state, NIR absorption spectroscopy was performed (Figures 4, 5). This analysis is particularly useful for detecting CH, NH, and OH groups [16]. The CH-derived peaks of SG were identified at 4,367 cm<sup>-1</sup>. The OH group peaks of  $\beta$ CD were observed at 4,808 cm<sup>-1</sup> and 6,920 cm<sup>-1</sup>, while the water content peak of  $\beta$ CD was observed at 5,208 cm<sup>-1</sup> (Figure 4). In PM (SG/ $\beta$ CD = 1/2), the peak derived from -CH of SG was observed at 4,367 cm<sup>-1</sup>, and no shift was observed in  $\beta$ CD. In GM (SG/ $\beta$ CD = 1/2), the peak of 4,367 cm<sup>-1</sup> derived from the CH group of SG was observed to shift to 4,386 cm<sup>-1</sup> at a higher



Figure 2. PXRD patterns of SG intact, SG/ $\beta$ CD, and SG/ $\gamma$ CD systems. (a) SG intact, (b)  $\beta$ CD intact, (c) PM (SG/ $\beta$ CD = 1/1), (d) PM (SG/ $\beta$ CD = 1/2), (e) GM (SG/ $\beta$ CD = 1/1), (f) GM (SG/ $\beta$ CD = 1/2), (g)  $\gamma$ CD intact, (h) PM (SG/ $\gamma$ CD = 1/1), (i) PM (SG/ $\gamma$ CD = 1/2), (j) GM (SG/ $\gamma$ CD = 1/1), (k) GM (SG/ $\gamma$ CD = 1/2). SG: stearyl glycyrrhetinate; CD: cyclodextrin; GM: ground mixtures; PM: physical mixtures. Open circle: SG original; blue solid triangle:  $\beta$ CD original; green solid square:  $\gamma$ CD original



Figure 3. DSC curves of SG intact, SG/ $\beta$ CD, and SG/ $\gamma$ CD systems. (a) SG intact, (b)  $\beta$ CD intact, (c) PM (SG/ $\beta$ CD = 1/1), (d) PM (SG/ $\beta$ CD = 1/2), (e) GM (SG/ $\beta$ CD = 1/1), (f) GM (SG/ $\beta$ CD = 1/2), (g)  $\gamma$ CD intact, (h) PM (SG/ $\gamma$ CD = 1/1), (i) PM (SG/ $\gamma$ CD = 1/2), (j) GM (SG/ $\gamma$ CD = 1/1), (k) GM (SG/ $\gamma$ CD = 1/2). Heating rate: 10 °C/min. SG: stearyl glycyrrhetinate; CD: cyclodextrin; GM: ground mixtures; PM: physical mixtures



Figure 4. Second differentiation NIR absorption spectra of SG/ $\beta$ CD systems. (a) 7,500–6,500 cm<sup>-1</sup>, (b) 5,500–4,500 cm<sup>-1</sup>, (c) 4,600–4,200 cm<sup>-1</sup>. SG: stearyl glycyrrhetinate; CD: cyclodextrin; GM: ground mixtures; PM: physical mixtures



Figure 5. Second differentiation NIR absorption spectra of SG/ $\gamma$ CD systems. (a) 7,500–6,500 cm<sup>-1</sup>, (b) 5,500–4,500 cm<sup>-1</sup>, (c) 4,600–4,200 cm<sup>-1</sup>. SG: stearyl glycyrrhetinate; CD: cyclodextrin; GM: ground mixtures; PM: physical mixtures



Figure 6. Scanning electron microscope images of SG intact, SG/ $\beta$ CD, and SG/ $\gamma$ CD systems. (a) SG intact, (b)  $\beta$ CD intact, (c) PM (SG/ $\beta$ CD = 1/1), (d) PM (SG/ $\beta$ CD = 1/2), (e) GM (SG/ $\beta$ CD = 1/1), (f) GM (SG/ $\beta$ CD = 1/2), (g)  $\gamma$ CD intact, (h) PM (SG/ $\gamma$ CD = 1/1), (i) PM (SG/ $\gamma$ CD = 1/2), (j) GM (SG/ $\gamma$ CD = 1/1), (k) GM (SG/ $\gamma$ CD = 1/2). SG: stearyl glycyrrhetinate; CD: cyclodextrin; GM: ground mixtures; PM: physical mixtures

wavenumber. In addition, the peak derived from the –OH group of  $\beta$ CD shifted to a lower wavenumber of 4,751 cm<sup>-1</sup>, and the peak derived from the water content of  $\beta$ CD shifted to a lower wavenumber of 5,181 cm<sup>-1</sup>. The OH group peaks of  $\gamma$ CD were observed at 4,739 cm<sup>-1</sup> and 6,897 cm<sup>-1</sup>, and the water-containing peak of  $\gamma$ CD was observed at 5,195 cm<sup>-1</sup> (Figure 5). In PM (SG/ $\gamma$ CD = 1/2), the peak derived from –CH of SG was observed at 4,357 cm<sup>-1</sup>, and no shift was observed in  $\gamma$ CD. In GM (SG/ $\gamma$ CD = 1/2), the peak derived from the CH group at 4,357 cm<sup>-1</sup> was observed to shift to a lower wavenumber at 4,367 cm<sup>-1</sup>. In addition, the OH group peak of  $\gamma$ CD related to 4,739 cm<sup>-1</sup> was confirmed to shift to a higher wavenumber at 4,796 cm<sup>-1</sup>. Although no peak shift was observed in the water-containing peak of  $\gamma$ CD observed at 5,195 cm<sup>-1</sup>, the peak was broadened. The peak of the OH group of  $\gamma$ CD at 6,897 cm<sup>-1</sup> was observed to shift to a higher wavenumber at 6,920 cm<sup>-1</sup>.

#### Assessment of morphology via SEM

SEM measurements were performed to examine the morphology of SG/CDs (Figure 6). The surface of SG was observed to be flat, with small debris adhering to larger, thinner debris (Figure 6a). In  $\beta$ CD and  $\gamma$ CD, smooth and angular surfaces were observed with a size of about 50–80 µm. (Figure 6b, g). In PM, the crystals of SG,  $\beta$ CD, and  $\gamma$ CD were 50 µm in diameter, rough, and angular (Figure 6c, d, h, i). Generally, inclusions formed by CDs have been described as cubic [17]. In GM (SG/ $\beta$ CD = 1/1) and GM (SG/ $\gamma$ CD = 1/1), angular cubic particles that looked like aggregates of small fragments 30 µm in diameter were observed, as well as cubic crystals in GM (SG/ $\beta$ CD = 1/2) and GM (SG/ $\gamma$ CD = 1/2) (Figure 6e, f, j, k).

#### <sup>1</sup>H-NMR spectrum measurement

In the solid state, PXRD, DSC, and NIR measurements suggested that SG formed an inclusion complex with the CDs. <sup>1</sup>H NMR spectroscopy was performed to predict the relative interactions between SG and CDs. Proton NMR measurements were performed to examine the intermolecular interactions in the solution. The results of GM (SG/ $\beta$ CD = 1/2) (DMSO-d<sub>6</sub>) are shown in Figure 7. In intact SG, H-A was observed at approximately 2.98–3.03 ppm, and the H-F moiety derived from hydroxyl moieties was observed at approximately 4.25 ppm. In intact CD, H-3,5,6 derived from the CD cavity was observed at approximately 3.50 to 3.62 ppm. Furthermore, OH-2,3,6 (OH-2: 5.71 ppm, OH-3: 5.66 ppm, OH-6: 4.44 ppm) derived from outside the  $\beta$ CD cavity were observed [18]. In PM (SG/ $\beta$ CD = 1/1) and PM (SG/ $\beta$ CD = 1/2), SG intact and  $\beta$ CD-derived proton peaks were observed, and no change in peaks was observed. Similarly, no change in the peaks was observed in GM (SG/ $\beta$ CD = 1/1) and GM (SG/ $\beta$ CD = 1/2).



**Figure 7.** <sup>1</sup>**H-NMR spectra of SG**/β**CD systems (DMSO-d<sub>6</sub>).** SG: stearyl glycyrrhetinate; CD: cyclodextrin; GM: ground mixtures; PM: physical mixtures



**Figure 8.** <sup>1</sup>**H-NMR spectra of SG/γCD systems (DMSO-d<sub>6</sub>).** SG: stearyl glycyrrhetinate; CD: cyclodextrin; GM: ground mixtures; PM: physical mixtures

The results of GM (SG/ $\gamma$ CD = 1/2) (DMSO-d<sub>6</sub>) are shown in Figure 8. In  $\gamma$ CD, H-3, H-5, and H-6 protons within the CD cavity were observed in the range of 3.50 to 3.62 ppm, while the OH-2, OH-3, and OH-6 protons outside the CD cavity appeared at approximately 5.75, 5.73, and 4.51 ppm, respectively. For PM (SG/ $\gamma$ CD = 1/1) and PM (SG/ $\gamma$ CD = 1/2), intact SG and  $\gamma$ CD-derived proton peaks were detected, with no significant shifts observed. However, in GM (SG/ $\gamma$ CD = 1/1), the H-3 and H-6 peaks of  $\gamma$ CD within its cavity (3.50–3.69 ppm) shifted slightly downfield (H-3: from 3.61 to 3.62 ppm, H-6: from 3.58 to 3.59 ppm). In GM (SG/ $\gamma$ CD = 1/2), the H-3 and H-6 peaks within the  $\gamma$ CD cavity (3.56–3.64 ppm) also showed a downfield shift (H-3: from 3.61 to 3.62 ppm, H-6: from 3.58 to 3.59 ppm). Additionally, peak shifts were observed for

the H-A (2.98–3.03 ppm) and H-F (4.29 ppm) protons in SG, as well as for the OH-2 (5.75 ppm), OH-3 (5.73 ppm), and OH-6 (4.51 ppm) protons of  $\gamma$ CD. Interestingly, the OH-2 and OH-3 peaks of  $\gamma$ CD changed from sharp signals to broad peaks, while the OH-6 peak transitioned from a sharp triplet to a broad peak. Notably, the H-A and H-F peaks of SG also exhibited significant broadening (H-A: multiple peaks to broad doublet peaks, H-F: doublet peak to broad peak) in GM (SG/ $\gamma$ CD = 1/2).

# Discussion

The results of PXRD measurements showed that the characteristic diffraction peaks of SG,  $\beta$ CD, and  $\gamma$ CD disappeared in GM (SG/ $\beta$ CD = 1/2) and GM (SG/ $\gamma$ CD = 1/2), indicating a halo pattern [19, 20]. Furthermore, co-grinding can cause disorders due to the breakdown of the crystalline structure, and these amorphous solids exhibit halo patterns. When the PXRD pattern of a poorly soluble drug exhibits a halo pattern, it indicates that the material is in an amorphous state. In the amorphous state, molecules are arranged in a disordered manner compared to the ordered structure of crystalline solids. This state has higher energy, which enhances interactions with solvents and is expected to improve solubility compared to crystalline materials. In this study, mechanochemical treatment is specifically proposed to modify the surface energy of drug particles, thereby increasing their affinity with solvents. Therefore, mechanical energy in the form of distortion or percussion may induce mechanochemical interactions between SG and  $\beta$ CD in the solid state [21].

The DSC results of GM (SG/ $\beta$ CD = 1/2) and GM (SG/ $\gamma$ CD = 1/2) showed the disappearance of the endothermic peak of SG and the decrease of the dehydration-derived peak of CD, suggesting the formation of an inclusion complex [16, 22]. This was presumably due to the loss of water from the CD, which was not involved in the inclusion complex. Thermal analysis showed changes in the thermal behavior, and the above findings suggest intermolecular interactions between GM (SG/CDs) and changes in the SG properties. The PXRD and DSC results suggested that SG/CD complexes may be formed at an SG/CD molar ratio of 1/2. Therefore, to confirm the intermolecular interactions of SG/ $\beta$ CD and SG/ $\gamma$ CD in the solid state, NIR absorption spectroscopy, which is useful for analyzing the –CH and OH groups, was performed [23]. The results of the NIR measurements suggest that both GM (SG/ $\beta$ CD = 1/2) and GM (SG/ $\gamma$ CD = 1/2) have significant peak shifts of the CH groups of SG and the OH moieties of the cavities of  $\beta$ CD and  $\gamma$ CD, suggesting that molecular interaction occurs by inclusion in the CD cavities. In SEM observation, generally, when CDs form inclusion complexes, cubic crystals are reported to appear. In the GM of SG/CDs (1/1), the particle size was reduced to around 30 µm due to milling, and the particles exhibited a cubic shape with fine surface attachments, suggesting the formation of an inclusion complex.

The <sup>1</sup>H-NMR spectra measurement results showed no peak shift in GM (SG/ $\beta$ CD = 1/1 and 1/2). However, in GM (SG/ $\gamma$ CD = 1/2), it was observed that the OH-2,3,6 peaks of  $\gamma$ CD were broadened. In particular, in GM (SG/ $\gamma$ CD = 1/2), the broadening of the H-A and H-F peaks of SG suggests an interaction involving the steroid skeleton containing H-A and H-F. Moreover, the wider ring structure of  $\gamma$ CD compared to  $\beta$ CD may facilitate more effective encapsulation of the steroid skeleton of SG. We have previously reported that the inclusion complex of ursolic acid (which also has a steroid skeleton) with  $\gamma$ CD adopts an inclusion mode where UA is encapsulated from the narrow edge of the CD [16]. Based on these findings, we speculate that in solution, GM (SG/ $\gamma$ CD = 1/2) forms a complex with a CD:2 interaction involving the stearyl group of SG, along with potential interactions between the steroid backbone and  $\gamma$ CD. Furthermore, the complex formation behavior appears to differ between GM (SG/ $\beta$ CD = 1/1 and 1/2) and GM (SG/ $\gamma$ CD = 1/1 and 1/2). As a next step, we will try single crystal structure analysis as a promising method for structural analysis of SG/CD complexes, which will greatly facilitate the discussion and interpretation of NMR results that strongly support the present results.

Limitations of the present study include the fact that it is generally recommended to examine the solubility phase diagram and the inclusion of molar ratio by Job's plot to elucidate the complex formation between CD and the drug. However, as a limitation of our study, we could not quantify SG because its solubility in water was too low [SG <  $0.5 \mu g/mL$  (D.W.)]. In addition, SG was not soluble in D<sub>2</sub>O in NMR

measurement, and the proton peak by NMR measurement could not be detected. Therefore, DMSO-d<sub>6</sub> was used. Therefore, in this study, the complex was not prepared by the solution method, but by the GM method using mechanochemical processes, and physical properties in the solid state were evaluated as a basic step, followed by the elucidation of intermolecular interactions by NMR techniques. It is necessary to establish a quantitative method other than HPLC to confirm the solubility of SG. Alternatively, as a next research topic, preparing complexes using CD derivatives (e.g., HP- $\beta$ CD and SBE- $\beta$ CD) may help improve solubility. In the future, the SG/CD complexes obtained in this study could be further investigated for their anti-inflammatory effects and solubility. This would contribute to skin application research, addressing issues such as rough skin and other skin problems.

# Abbreviations

CD: cyclodextrin DSC: differential scanning calorimetry GA: 18-β-glycyrrhetinic acid GL: glycyrrhizic acid GM: ground mixtures NIR: near-infrared NMR: nuclear magnetic resonance PM: physical mixtures PXRD: powder X-ray diffraction SEM: scanning electron microscopy SG: stearyl glycyrrhetinate

# **Declarations**

## Acknowledgments

The authors are grateful to Cyclo Chem Bio Co., Ltd. for providing the cyclodextrin samples. The authors wish to thank the Instrument Analysis Center of Josai University for helpful advice regarding NMR spectroscopy. We thank the Laboratory of Nutri-Pharmacotherapeutics Management, Josai University, for research support while taking measures against COVID-19 (SARS-CoV2) infection.

## Author contributions

ME: Investigation, Data curation, Writing—original draft. NK: Data curation. SM: Conceptualization, Data curation. JT, MS: Investigation, Validation, Writing—review & editing, Project administration, Supervision. YI: Investigation, Conceptualization, Validation, Writing—review & editing, Project administration, Supervision. All the authors have read and approved the submitted manuscript.

## **Conflicts of interest**

The authors declare that this study received  $\beta$ -cyclodextrin from Cyclo Chem Co., Ltd., which was not involved in the study design, collection, analysis, interpretation of data, the writing of this article, or the decision to submit it for publication. The authors declare that the research was conducted in the absence of commercial or financial relationships that could be construed as potential conflicts of interest.

## **Ethical approval**

Not applicable.

#### **Consent to participate**

Not applicable.

### **Consent to publication**

Not applicable.

## Availability of data and materials

The original contributions presented in the study are included in the article, further inquiries can be directed to the corresponding author.

## Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

## Copyright

© The Author(s) 2025.

# **Publisher's note**

Open Exploration maintains a neutral stance on jurisdictional claims in published institutional affiliations and maps. All opinions expressed in this article are the personal views of the author(s) and do not represent the stance of the editorial team or the publisher.

# References

- Althobaiti HM, Althobaiti H, Khan M, Alsatti H, Samarkandy SJ. The Association Between Facial Dermatosis and Face-Mask Wearing During COVID-19 in Saudi Arabia. Cureus. 2022;14:e22265. [DOI] [PubMed] [PMC]
- 2. Gomaa AA, Abdel-Wadood YA. The potential of glycyrrhizin and licorice extract in combating COVID-19 and associated conditions. Phytomed Plus. 2021;1:100043. [DOI] [PubMed] [PMC]
- 3. Santonocito D, Puglia C, Torrisi C, Giuffrida A, Greco V, Castelli F, et al. Calorimetric Evaluation of Glycyrrhetic Acid (GA)- and Stearyl Glycyrrhetinate (SG)-Loaded Solid Lipid Nanoparticle Interactions with a Model Biomembrane. Molecules. 2021;26:4903. [DOI] [PubMed] [PMC]
- Oyama K, Kawada-Matsuo M, Oogai Y, Hayashi T, Nakamura N, Komatsuzawa H. Antibacterial Effects of Glycyrrhetinic Acid and Its Derivatives on Staphylococcus aureus. PLoS One. 2016;11:e0165831.
  [DOI] [PubMed] [PMC]
- Sakata O, Fujii M, Koizumi N, Nakade M, Kameyama K, Watanabe Y. Effects of oils and emulsifiers on the skin penetration of stearyl glycyrrhetinate in oil-in-water emulsions. Biol Pharm Bull. 2014;37: 486–9. [DOI] [PubMed]
- 6. Piquero-Casals J, Hexsel D, Mir-Bonafé JF, Rozas-Muñoz E. Topical Non-Pharmacological Treatment for Facial Seborrheic Dermatitis. Dermatol Ther (Heidelb). 2019;9:469–77. [DOI] [PubMed] [PMC]
- Cid-Samamed A, Rakmai J, Mejuto JC, Simal-Gandara J, Astray G. Cyclodextrins inclusion complex: Preparation methods, analytical techniques and food industry applications. Food Chem. 2022;384: 132467. [DOI] [PubMed]
- 8. Braga SS, Barbosa JS, Santos NE, El-Saleh F, Paz FAA. Cyclodextrins in Antiviral Therapeutics and Vaccines. Pharmaceutics. 2021;13:409. [DOI] [PubMed] [PMC]
- 9. Ho S, Thoo YY, Young DJ, Siow LF. Cyclodextrin encapsulated catechin: Effect of pH, relative humidity and various food models on antioxidant stability. LWT Food Sci Tech. 2017;85:232–39. [DOI]
- 10. Przybyla MA, Yilmaza G, Becer CR. Natural cyclodextrins and their derivatives for polymer synthesis. Polym Chem. 2020;11:7582–602. [DOI]

- 11. Cheirsilp B, Rakmai J. Inclusion complex formation of cyclodextrin with its guest and their applications. Biol Eng Med. 2016;2:1–6. [DOI]
- 12. Jansook P, Ogawa N, Loftsson T. Cyclodextrins: structure, physicochemical properties and pharmaceutical applications. Int J Pharm. 2018;535:272–84. [DOI] [PubMed]
- 13. Fenyvesi É, Vikmon M, Szente L. Cyclodextrins in food technology and human nutrition: Benefits and limitations. Crit Rev Food Sci Nutr. 2016;9;56:1981–2004. [DOI]
- 14. Inoue Y, Motoda A, Tanikawa T, Takao K, Arce F Jr, See GL, et al. Inclusion complexes of ursolic acid with cyclodextrin-based metal-organic framework-1 enhance its solubility. J Drug Deliv Sci Technol. 2023;89:104986. [DOI]
- 15. Alfei S, Schito AM, Zuccari G. Considerable Improvement of Ursolic Acid Water Solubility by Its Encapsulation in Dendrimer Nanoparticles: Design, Synthesis and Physicochemical Characterization. Nanomaterials (Basel). 2021;11:2196. [DOI] [PubMed] [PMC]
- 16. Inoue Y, Yoshida M, Ezawa T, Tanikawa T, Arce F Jr, See GL, et al. Inclusion Complexes of Daidzein with Cyclodextrin-Based Metal-Organic Framework-1 Enhance Its Solubility and Antioxidant Capacity. AAPS PharmSciTech. 2021;23:2. [DOI] [PubMed]
- 17. Prabu S, Sivakumar K, Swaminathan M, Rajamohan R. Preparation and characterization of host-guest system between inosine and β-cyclodextrin through inclusion mode. Spectrochim Acta A Mol Biomol Spectrosc. 2015;147:151–7. [DOI] [PubMed]
- Bucur S, Niculaua M, Ciobanu CI, Lungu NC, Mangalagiu I. A Simple Synthesis Route for Selectively Methylated β-Cyclodextrin Using a Copper Complex Sandwich Protecting Strategy. Molecules. 2021; 26:5669. [DOI] [PubMed] [PMC]
- 19. Ogawa N, Higashi K, Nagase H, Endo T, Moribe K, Loftsson T, et al. Effects of cogrinding with βcyclodextrin on the solid state fentanyl. J Pharm Sci. 2010;99:5019–29. [DOI] [PubMed]
- Mura P, Bettinetti GP, Cirri M, Maestrelli F, Sorrenti M, Catenacci L. Solid-state characterization and dissolution properties of naproxen-arginine-hydroxypropyl-beta-cyclodextrin ternary system. Eur J Pharm Biopharm. 2005;59:99–106. [DOI] [PubMed]
- 21. Inoue Y, Watanabe S, Suzuki R, Murata I, Kanamoto I. Evaluation of actarit/γ-cyclodextrin complex prepared by different methods. J Incl Phenom Macrocycl Chem. 2015;81:161–8. [DOI]
- Aigner Z, Berkesi O, Farkas G, Szabó-Révész P. DSC, X-ray and FTIR studies of a gemfibrozil/dimethylβ-cyclodextrin inclusion complex produced by co-grinding. J Pharm Biomed Anal. 2012;57:62–7.
  [DOI] [PubMed]
- Yamamoto K, Tanikawa T, Tomita J, Ishida Y, Nakata D, Terao K, et al. Characterization, Preparation, and Promotion of Plant Growth of 1,3-Diphenylurea/β-Cyclodextrin Derivatives Inclusion Complexes. ACS Omega. 2023;8:34972–81. [DOI] [PubMed] [PMC]