






# Significance of FXR agonists in MASLD treatment: a deep dive into lipid alteration by analytical techniques

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**Academic Editor:** Karel Pacak, AKESO Holding, Czech Republic

**Received:** October 15, 2024 **Accepted:** February 14, 2025 **Published:** March 25, 2025

**Cite this article:** Srikanth P, Shaik KM, Patibandla V, Kumar D, Nandi S. Significance of FXR agonists in MASLD treatment: a deep dive into lipid alteration by analytical techniques. *Explor Endocr Metab Dis.* 2025;2:101425. <https://doi.org/10.37349/eemd.2025.101425>

## Abstract

Metabolic dysfunction-associated steatotic liver disease (MASLD) is rapidly emerging as a global health crisis, affecting over 30% of the population and demanding urgent attention. This redefined condition, previously known as non-alcoholic fatty liver disease (NAFLD), reflects a deeper understanding of the intricate interplay between metabolic dysfunction and liver health. At the heart of MASLD lies the troubling accumulation of triglycerides (TGs) in hepatocytes, which precipitates insulin resistance and oxidative stress, ultimately leading to more severe forms like metabolic dysfunction-associated steatohepatitis (MASH). Excitingly, recent research has spotlighted the farnesoid X receptor (FXR) as a groundbreaking therapeutic target. FXR not only regulates lipid metabolism but also combats inflammation and insulin resistance, making it a potential game-changer in the fight against MASLD. With only one FDA-approved drug, resmetirom, currently available, the exploration of FXR agonists opens new avenues for innovative treatments that could revolutionize patient care. By harnessing the power of FXR to restore metabolic balance and integrating advanced strategies like lipidomics and fatty acid profiling, we stand on the brink of transforming how we approach MASLD and its associated complications, paving the way for a healthier future. This review delves into the promising role of FXR in combating MASLD and its implications for related metabolic disorders, emphasizing the urgency for advanced strategies to detect and manage this burgeoning epidemic.

## Keywords

Metabolic dysfunction-associated steatotic liver disease (MASLD), metabolic dysfunction-associated steatohepatitis (MASH), non-alcoholic fatty liver disease (NAFLD), non-alcoholic steatohepatitis (NASH), farnesoid X receptor (FXR), fatty acids, analytical techniques, lipid quantification



## Introduction

The landscape of liver disease nomenclature has recently undergone a significant transformation, igniting curiosity and discussion among healthcare professionals and researchers alike. The shift from non-alcoholic fatty liver disease (NAFLD) to metabolic dysfunction-associated steatotic liver disease (MASLD), along with the transition from non-alcoholic steatohepatitis (NASH) to metabolic dysfunction-associated steatohepatitis (MASH), reflects an evolving scientific understanding that emphasises the metabolic dysfunction underlying these conditions. This rebranding highlights that the “M” in MASLD and MASH signifies metabolic dysfunction, inherently linking these diseases to prevalent metabolic disorders such as obesity and type 2 diabetes, which are critical contributors to liver injury [1, 2]. The reformulation of these terms was guided by a comprehensive Delphi consensus process involving multiple stakeholders in hepatology, aimed at establishing a more accurate and clinically relevant framework for diagnosis and treatment [3, 4]. Currently, the global prevalence of MASLD is estimated at approximately 32%, with projections suggesting it could rise to 55.4% by 2040 due to increasing rates of obesity and type 2 diabetes [5–7]. In India, the prevalence stands at around 38% among adults, indicating a significant public health concern as the nation faces rising obesity rates and associated metabolic disorders [6, 8–10]. Regions such as Latin America report even higher prevalence rates, reaching 44.4%, underscoring the urgent need for targeted interventions globally [6, 11–13].

Despite advancements in nomenclature and understanding, the pathogenesis of MASLD remains incompletely elucidated, with steatosis defined as the presence of triglycerides (TGs) in hepatocytes serving as the primary precursor for the disease [14–17]. The development of hepatic steatosis arises from an imbalance between lipid intake and lipid removal, which is regulated by four key mechanisms: fatty acid uptake, de novo lipogenesis (DNL), fatty acid oxidation, and lipid export. These mechanisms play a crucial role in both the physiological and pathological progression of MASLD [11, 18–20].

Targeting farnesoid X receptor (FXR) presents a promising therapeutic strategy for managing MASLD because FXR is integral in regulating these four key mechanisms of hepatic lipid metabolism [21]. By selectively modulating FXR activity through agonist ligands, it can effectively control lipid acquisition (fatty acid acquisition) and lipid removal to mitigate hepatic steatosis and its progression to more severe liver conditions [22, 23]. Furthermore, FXR agonists have demonstrated significant histological improvements in clinical trials for liver diseases, indicating their potential to enhance treatment outcomes for patients with MASLD [24–29]. This approach not only targets the underlying metabolic dysfunction but also opens avenues for combination therapies that may improve efficacy while minimising side effects associated with current treatments [30].

As research progresses, exploring FXR’s role in MASLD will be crucial for developing innovative therapeutic strategies to effectively manage this complex disease and its associated metabolic disorders [31]. This review explores the role of FXR in liver physiology and the progression of MASLD. We begin by examining FXR’s normal functions and how its dysregulation contributes to disease pathology. We then identify specific lipid classes that alter during the shift from health to disease, highlighting their significance as diagnostic markers and potential biomarkers. Following this, we discuss treatment strategies using FXR agonists and ongoing clinical trials evaluating their effectiveness in managing MASLD. Additionally, we emphasize the crucial roles of fatty acids and lipid classes in the pathogenesis of MASLD and present lipid analyses conducted with hyphenated techniques to reinforce our findings on relevant biomarkers. To ensure clarity and consistency, we will refer to the disease as MASLD and MASH, regardless of whether the underlying populations in the referenced studies were originally classified as patients with NAFLD or NASH.

## Association of MASLD with features of the metabolic syndrome

MASLD condition is characterized by fat deposition in the liver, which can lead to inflammation and fibrosis, ultimately resulting in more severe liver disease, MASH [32–34]. The relationship between MASLD and metabolic syndrome is mutual and bi-directional, highlighting the complexity of these interconnected conditions [35, 36].

## Mutual association with metabolic syndrome

MASLD is frequently associated with several components of metabolic syndrome, including obesity, type 2 diabetes mellitus (T2DM), dyslipidaemia, and hypertension. These conditions collectively contribute to the development and progression of MASLD [37–39]. For instance, individuals with obesity often exhibit increased hepatic fat accumulation, which can exacerbate insulin resistance and lead to T2DM [40–42]. Conversely, the presence of MASLD can worsen metabolic syndrome features; for example, hepatic steatosis can lead to increased insulin resistance, further complicating glucose metabolism and lipid profiles [43–45]. Research shows that a substantial number of individuals, including those who are lean, are affected by MASLD, with about 25% of lean individuals living with HIV also exhibiting the condition, indicating that metabolic dysfunction is not exclusively associated with obesity [6, 37, 46]. These statements emphasize the importance of recognizing MASLD in diverse populations and its potential to influence metabolic health across different demographic groups.

## Bi-directional relationship

The relationship between MASLD and metabolic syndrome is bi-directional [47, 48]. While metabolic syndrome increases the risk of developing MASLD, existing liver disease can also exacerbate features of metabolic syndrome [32, 49, 50]. For example, elevated levels of free fatty acids (FFAs) (for example, oleic acid, palmitic acid, stearic acid, and linoleic acid, etc.) in individuals with MASLD can contribute to systemic insulin resistance and dyslipidaemia by promoting lipogenesis and inhibiting fatty acid oxidation in peripheral tissues [51–53]. Furthermore, the buildup of lipids in the liver can lead to an increased production of pro-inflammatory cytokines [for example, interleukin (IL)-1, IL-6, IL-12, IL-17, etc.], which may worsen insulin sensitivity and promote cardiovascular risk factors associated with metabolic syndrome [54, 55].

## Role of fatty acids

Fatty acid plays a crucial role in intricate relationships. They are central to understanding the pathophysiology of MASLD. The disease is characterized by alterations in hepatic and plasma lipid balance, which are critical for diagnosing and assessing its severity [56]. Hepatic insulin resistance, a key feature of MASLD, correlates with increased levels of diacylglycerol (DAG) and triacylglycerol (TAG), as identified in lipidomics studies [57]. Elevated circulating FFAs may significantly contribute to hepatic lipotoxicity, leading to liver inflammation and fibrosis [32, 58, 59] particularly associated with saturated fatty acids (SFA) such as palmitic acid [60]. In contrast, unsaturated fatty acids (UFA) have been observed to provide protective effects against liver damage and can enhance insulin sensitivity [61–64]. The detrimental impact of SFA on liver health is evident through their accumulation in hepatocytes, which can induce apoptosis and lipotoxicity. Factors influencing fatty acid levels include diet, age, gender and hepatic DNL [65]. Studies indicate that excessive fatty acid accumulation can lead to endoplasmic reticulum stress and hepatocyte injury [66]. The balance between different types of fatty acids is crucial for both the progression of MASLD and the general metabolic health of individuals [64, 67]. Specific lipid profiles have emerged as potential biomarkers for MASLD diagnosis; during the condition, there is an increase in SFA like myristic and palmitic acids [68], while alterations in TG levels and certain phospholipid species may reflect disease progression or treatment response [69–71].

In liver samples from patients with MASLD, there is an observed increase in monounsaturated fatty acids (MUFA) alongside elevated DAG and TAG levels [72, 73]. Inversely, polyunsaturated fatty acids (PUFAs) such as eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) tend to decrease in MASLD conditions [74, 75]. Additionally, activation of FXR has been shown to reduce hepatic lipid levels and intestinal lipid absorption, highlighting potential therapeutic pathways for managing MASLD [76]. Overall, understanding the intricate relationship between various fatty acids and their metabolic implications is essential for addressing MASLD effectively. Fatty acids represent a class of lipid profiles, and various alterations in lipid composition associated with different metabolic disorders are detailed in [Table 1](#).

**Table 1. Alteration of lipids in different metabolic disorders**

Condition	Lipid alterations
Obesity	Obesity is often associated with dyslipidaemia, which presents as elevated plasma levels of TGs, VLDL, and low HDL values. Every 10 pounds of extra fat increases daily cholesterol production by 10 mg. The lipid profile rises in obese individuals [169].
MASLD	Hepatic lipid imbalance results from hepatic absorption of FAs and de novo lipogenesis exceeding fatty acid oxidation and lipid export in MASLD. This leads to modifications in the hepatic metabolism of lipids, potentially precipitating severe complications such as cirrhosis and HCC. Different MASLD patients had various levels of serum lysophosphatidylcholine, sphingomyelin, total cholesterol esters, and TAGs [170].
Type II diabetes	Anomalies of plasma lipids and lipoprotein associated with type II diabetes include reduced HDL cholesterol, an abundance of small dense LDL particles, and elevated TG levels. Every one of these dyslipidaemia traits is linked to an increased risk of CVD [171].
Type II diabetes with MASLD	There is a bidirectional pathophysiological link between MASLD and type 2 diabetes. On the one hand, the latter encourages the development of MASL into MASH, a more advanced form of the disease. HCC and cirrhosis are two conditions that are made more likely by NASH and may necessitate liver transplantation [172].
MASLD + obesity + type II diabetes	The risk of developing cirrhosis and MASH is significantly increased when type 2 diabetes, obesity, and MASLD are present than when MASLD is present, but chronic hyperglycaemia is not. Whether MASLD is a substantial independent predictor of CVD remains debatable [173].

TGs: triglycerides; VLDL: very low-density lipoprotein; HDL: high-density lipoprotein; MASLD: metabolic dysfunction-associated steatotic liver disease; FAs: fatty acids; HCC: hepatocellular carcinoma; TAGs: triacylglycerols; LDL: low-density lipoprotein; CVD: cardiovascular disease; MASL: metabolic dysfunction-associated steatotic liver; MASH: metabolic dysfunction-associated steatohepatitis; NASH: non-alcoholic steatohepatitis

## Physiological role of FXR

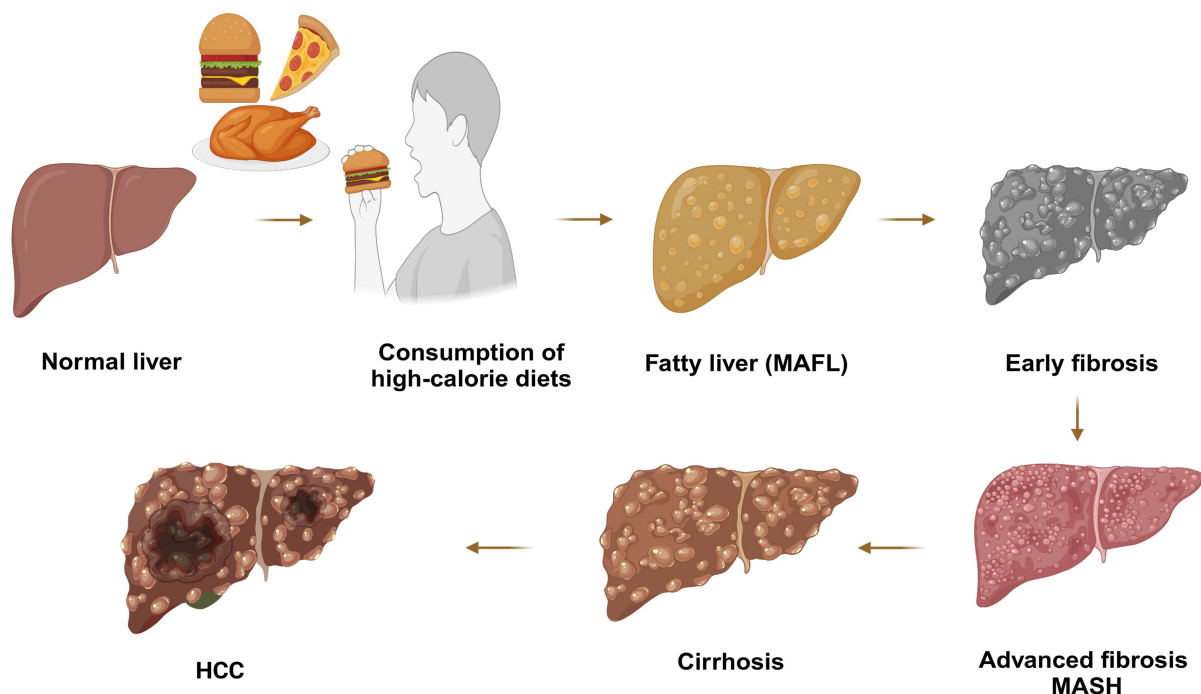
FXR is a key nuclear receptor that acts as a bile acid sensor, playing a significant role in maintaining metabolic homeostasis [77–79]. Predominantly found in the liver and intestine, FXR integrates metabolic signals and is activated by bile acids such as, which are crucial for cholesterol and lipid metabolism [77, 80, 81]. Upon activation, FXR heterodimerizes with retinoid X receptor (RXR) and binds to FXR response elements (FXREs) to regulate genes involved in bile acid synthesis and metabolism [82, 83]. It's essential for coordinating metabolic processes and acts as a therapeutic target for metabolic disorders [31, 84–86]. FXR is an essential modulator of lipid metabolism [87, 88]. Exerting its effects by modulating the activity of several transcription factors, including sterol regulatory element-binding protein-1c (SREBP-1c) and carbohydrate response element-binding protein (ChREBP). SREBP-1c is crucial for the regulation of genes involved in fatty acid and TG synthesis, and FXR activation leads to the repression of SREBP-1c expression. This inhibition is significant as it reduces hepatic TG levels and helps prevent hepatic steatosis, particularly in MASLD [89, 90]. Additionally, FXR interacts with ChREBP to fine-tune lipogenesis based on glucose availability, ensuring lipid synthesis is aligned with the body's energy status [91, 92]. The activation of FXR decreases the expression of fatty acid synthase (FASN), an enzyme essential for fatty acid biosynthesis, and regulates other pathways critical for lipid homeostasis [93, 94]. The small heterodimer partner (SHP) is another important component in this regulatory network. Upon activation by bile acids, FXR induces SHP expression, which acts as a transcriptional repressor of various target genes involved in lipogenesis and bile acid synthesis [95]. This mechanism helps to prevent excessive TG accumulation and maintain normal lipid profiles within hepatocytes [96, 97]. Furthermore, the FXR-SHP pathway interacts with other nuclear receptors such as peroxisome proliferator-activated receptor alpha (PPAR $\alpha$ ), enhancing its regulatory capacity on lipid metabolism [90, 98]. Comprehensive, through the modulation of SREBP-1c and ChREBP, along with the involvement of SHP, FXR plays a pivotal role in coordinating lipid metabolism [99, 100]. Xu et al. [101] studied the impact of the activation of the FXR by obeticholic acid (OCA) significantly inhibits hepatic cholesterol 7 $\alpha$ -hydroxylase (CYP7A1) and sterol 12 $\alpha$ -hydroxylase (Cyp8b1), partly through the induction of SHP. This inhibition leads to a reduced bile acid pool size and altered bile acid composition, which contributes to decreased intestinal cholesterol absorption and enhanced macrophage reverse cholesterol transport (RCT). The reduction in hepatic microsomal cholesterol content triggers an elevation in the expression and functionality of low-density lipoprotein receptors (LDL-R) within the liver [102]. Consequently, this cascade of events leads to a decrease in plasma LDL-cholesterol (LDL-C) levels [103]. In addition to inhibiting CYP7A1, enhanced excretion into bile occurs via the cholesterol transporters known as adenosine triphosphate-binding cassette transporters (ABCG5/8), which can elevate blood cholesterol

levels in knockout (KO) mice [104]. Reduction in intestinal bile acids and promotion of transintestinal cholesterol excretion serve as mechanisms to lower cholesterol levels in the intestine [105]. While FXR induces an anti-atherogenic effect in mice, the scenario differs in humans. This was possible in mice due to the presence of enzymes Cyp2c70 (cytochrome P450 2C70) and Cyp2a12 (cytochrome P450 2A12) [106–108].

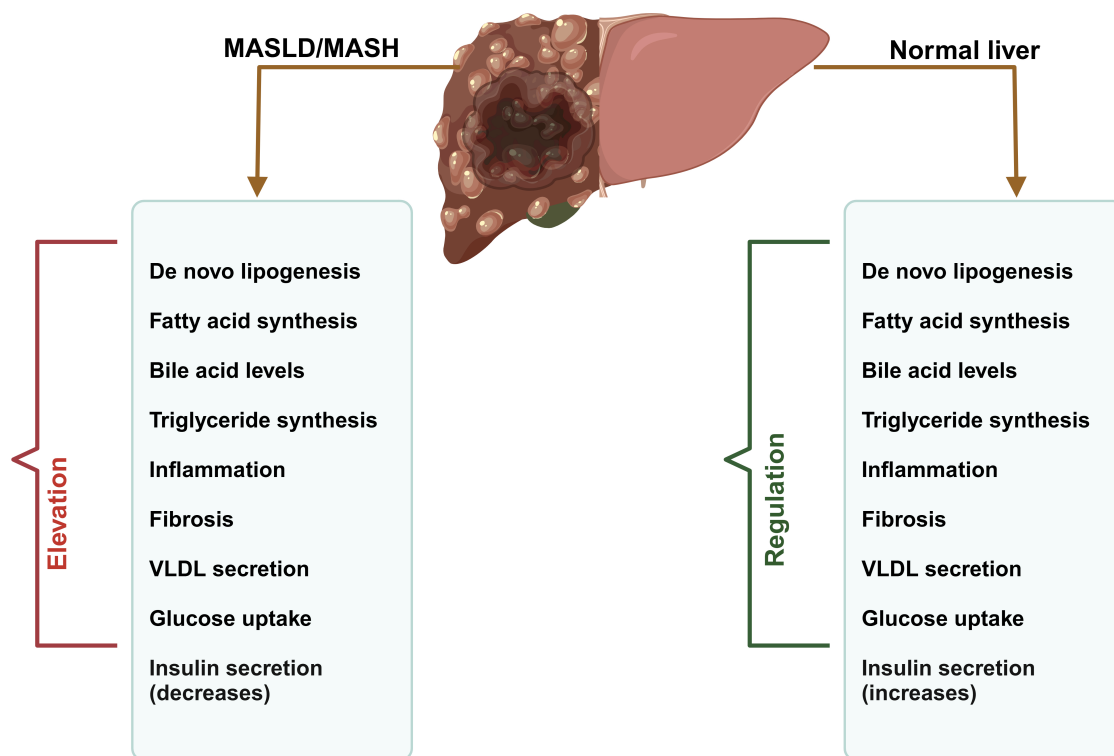
FXR-induced lipoprotein metabolism is complex and differs mainly in ligands and species. FXR is known to reduce high-density lipoprotein (HDL) cholesterol, resulting in a pro-atherogenic profile. The proatherogenic effect is due to the stimulating effects of cholesterol ester transfer protein (CETP), which increases LDL and decreases HDL cholesterol [109]. In addition, the transformation of very low-density lipoprotein (VLDL) rich in TG is converted to cholesterol-rich LDL particles [110]. The major difference between the normal liver and disease progression.

## Pathophysiology of FXR in MASLD and its related disorders

The mechanism behind MASLD primarily involves insulin resistance and lipotoxicity. Insulin resistance leads to increased FFA release from adipose tissue, which accumulates in the liver, causing hepatic steatosis as mentioned in Figure 1 [34, 51]. This accumulation triggers oxidative stress and inflammation, further exacerbated by elevated levels of reactive oxygen species (ROS) that activate pro-inflammatory pathways [111]. Increased ROS levels stimulate kinases such as c-Jun N-terminal kinase (JNK) and p38 mitogen-activated protein kinases (MAPK), promoting inflammation and disrupting insulin signalling. This results in a vicious cycle where inflammation worsens insulin resistance [112]. Additionally, dietary factors like high fructose and saturated fat intake enhance DNL and contribute to the progression from simple steatosis to steatohepatitis. The interplay between gut microbiota dysbiosis and metabolic perturbations also fuels hepatic inflammation and injury [113]. Understanding these specific mechanisms is crucial for developing targeted therapies for MASLD [114], as illustrated by the alterations in disease identifiers depicted in Figure 2.



**Figure 1. Progression from normal liver to cirrhosis and HCC Induced by high-calorie diet.** This visual representation underscores the importance of dietary management in preventing liver-related diseases and emphasizes the cascading effects of excessive calorie intake on liver function. MAFL: metabolic dysfunction-associated fatty liver; MASH: metabolic dysfunction-associated steatohepatitis; HCC: hepatocellular carcinoma. Created in BioRender. Pirangi, S. (2025) [BioRender.com/z08d915](https://doi.org/10.37349/eemd.2025.101425)



**Figure 2. Comparative analysis of liver health: normal vs. MASLD/MASH.** This visual emphasizes the critical role of lipogenic gene regulation in the transition from healthy liver function to advanced liver disease, underscoring the importance of dietary choices in preventing metabolic liver disorders. Due to the regulation of lipogenic genes (*SREBP-1c*, *ChREBP*, *FASN*, and *SHP*) regulated in normal and elevated in MASLD/MASH. MASLD: metabolic dysfunction-associated steatotic liver disease; MASH: metabolic dysfunction-associated steatohepatitis; VLDL: very low-density lipoprotein; *SREBP-1c*: sterol regulatory element-binding protein-1c; *ChREBP*: carbohydrate response element-binding protein; *FASN*: fatty acid synthase; *SHP*: small heterodimer partner. Created in BioRender. Pirangi, S. (2025) [BioRender.com/x79m262](https://BioRender.com/x79m262)

### Impaired lipid regulation

MASLD to MASH represents a significant shift in liver pathology, akin to a small canal (MASLD) flowing into a vast river (MASH). MASLD is characterized by hepatic steatosis, where fat accumulation exceeds 5% of liver weight, often associated with metabolic syndrome features such as obesity, insulin resistance and dyslipidaemia. This stage primarily involves fat accumulation without significant inflammation or fibrosis. In contrast, MASH is a more advanced condition that includes inflammation (steatohepatitis) and can progress to fibrosis, cirrhosis, and even hepatocellular carcinoma (HCC) in Figure 1. MASH is marked by hepatocyte ballooning, inflammatory cell infiltration, and varying degrees of fibrosis. Patients with MASLD may experience stable disease or progress to MASH over time. Few studies indicate that approximately one-third of patients with MASLD develop progressive fibrosis, while others may remain stable or even experience regression. The risk of severe complications increases significantly with advancing fibrosis stages, underscoring the importance of early detection and intervention. Understanding the mechanisms driving this progression is crucial for developing effective diagnostic and therapeutic strategies to manage these prevalent liver diseases effectively. MASLD is closely linked to lipid metabolism, particularly TG and cholesterol metabolism. Increased fatty acid uptake and synthesis, along with decreased lipid degradation, significantly contribute to MASLD. Key transport proteins, including fatty acid transport proteins (FATPs), cluster of differentiation 36 (CD36), and caveolin-1 (CAV-1), facilitate fatty acid uptake in the liver. FATP2 and FATP5 are primarily expressed in the liver, with downregulation of FATP2 reducing fatty acid uptake and improving hepatic steatosis. CD36 is crucial for fatty acid uptake; inhibiting its palmitoylation can alleviate metabolic disorders. CAV-1 also plays a role by promoting autophagy and reducing lipid accumulation. Additionally, lipogenic markers like *FASN* and stearoyl-CoA desaturase (*SCD*) are upregulated in MASLD [115]. Targeting *FASN* through mechanisms such as sorting nexin 8-mediated degradation offers a promising strategy for MASLD prevention. FXR agonists, including nidufexor (LMB763), cilofexor, and EDP-305, are currently undergoing clinical trials for the treatment of MASLD.

These compounds have shown promise in addressing key aspects of the disease. For instance, 1-adamantyl carbonyl-4-phenylpiperazine is an FXR agonist whose derivative, compound 10A, has demonstrated greater efficacy in ameliorating hyperlipidaemia, hepatic steatosis, and insulin resistance [21, 33, 115, 116].

### **Insulin resistance**

The accumulation of TGs in the liver is closely linked to insulin resistance, a hallmark of metabolic syndrome and MASLD [117]. IR is defined as a reduced response to insulin signalling, particularly in insulin-sensitive tissues like adipose tissue and the liver. This impairment leads to decreased glucose uptake and promotes lipolysis, resulting in elevated FFAs that accumulate in the liver and cause fat overload in hepatocytes [118]. The insulin receptor pathway regulates hepatic lipid metabolism through the transcription factor SREBP-1c, which upregulates genes involved in fatty acid biosynthesis, such as acetyl-CoA carboxylase (ACC) and FAS. MASLD-related hepatic IR, inhibition of the IR pathway leads to unregulated DNL, while gluconeogenesis remains unaffected [119].

### **Inflammation and fibrosis**

Chronic inflammation is another critical factor in the progression from simple steatosis to more severe forms of liver disease such as MASH and fibrosis [120]. FXR demonstrates anti-inflammatory effects by inhibiting the nuclear factor-kappa B (NF- $\kappa$ B) signalling pathways that play a role in inflammatory responses [121]. In conditions where FXR signalling is disrupted, there is an increase in pro-inflammatory cytokines and chemokines that promote inflammation and activate hepatic stellate cells (HSCs), leading to excessive extracellular matrix deposition and fibrosis [122]. The fibroblast growth factor (FGF)-15/19 axis is upregulated by FXR in enterocytes, with FGF-19 acting as an enterokinase that travels to the liver via the portal circulation. Upon reaching the liver, FGF-19 interacts with the FGF receptor 4 and  $\beta$ -klotho, leading to the repression of bile acid synthesis and gluconeogenesis. This regulation may support liver regeneration following injury. FXR agonists have demonstrated the ability to reduce fibrosis and steatosis while also exhibiting anti-inflammatory effects. In mice fed a methionine-choline-deficient (MCD) diet, the FXR agonist WAY-362450 effectively reduced liver inflammation and fibrogenesis without leading to an increase in TG levels [123, 124]. In reverse, impaired FXR function can exacerbate inflammatory responses, leading to progressive liver damage [122, 123].

### **Hepatic steatosis**

The buildup of fat in the liver due to impaired FXR signalling not only contributes to steatosis but also increases the risk of developing HCC [125]. Studies have indicated that mice lacking functional FXR develop hepatocellular adenomas and carcinomas spontaneously, suggesting that FXR may act as a tumour suppressor in the liver [126]. By regulating lipid metabolism and inflammatory responses, FXR helps maintain a balance that prevents tumorigenesis [127, 128].

### **HCC**

Recent studies have indicated that FXR may play a protective role against the development of HCC [84, 127]. HCC arises from metabolic dysregulation and chronic inflammation, often driven by increased IL-6 signalling, which plays a leading role in cancer development and progression. FXR has emerged as a significant therapeutic target in HCC, with its agonists, such as OCA, demonstrating tumour-suppressive effects. OCA inhibits the IL-6/Jak-2/STAT3 signalling pathway, thereby reducing STAT3 activation and increasing levels of suppressor of cytokine signalling 3 (SOCS3), a feedback inhibitor of STAT3 [129]. This modulation not only mitigates inflammation but also inhibits HCC proliferation, migration, and invasion, underscoring the potential of FXR agonists in developing effective treatment strategies for HCC [125]. The modulation of lipid metabolism by FXR may help mitigate the risk factors associated with HCC development [84].

### **Extra-hepatic manifestations: impact on chronic kidney disease**

Emerging evidence suggests that FXR also plays a role in extra-hepatic manifestations of MASLD, particularly chronic kidney disease (CKD) [130]. The interplay between liver dysfunction and kidney health

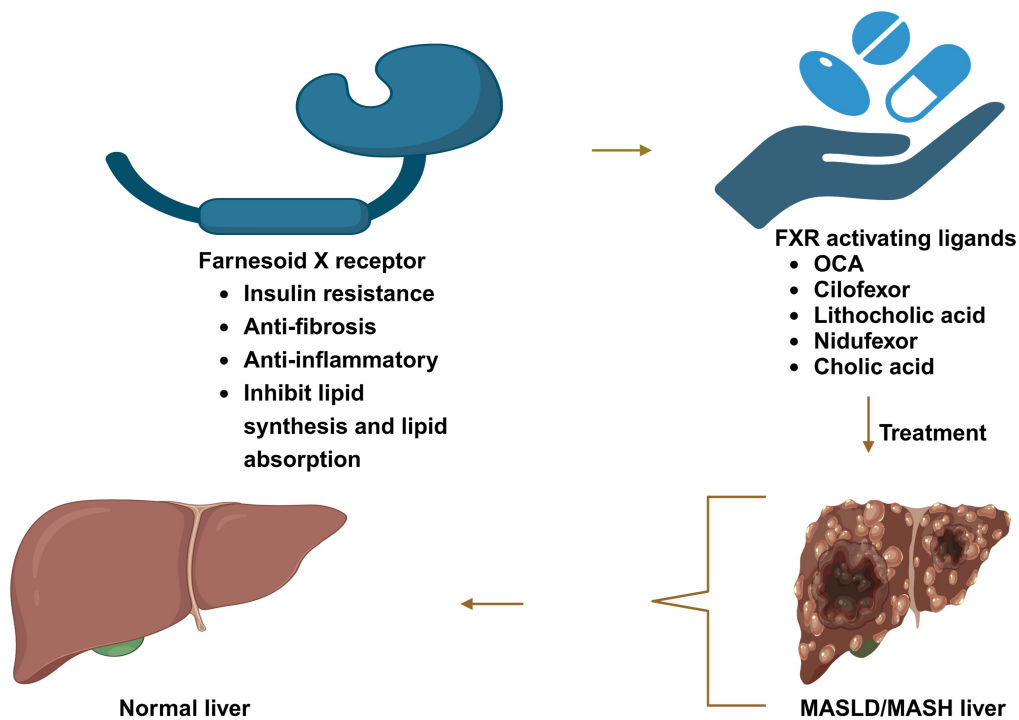
is complex; metabolic abnormalities stemming from hepatic steatosis can adversely affect renal function [131]. Elevated levels of circulating FFAs due to hepatic steatosis can lead to renal lipotoxicity, contributing to CKD progression [132]. FXR activation has been shown to exert protective effects on renal tissues by modulating lipid metabolism and reducing oxidative stress [133]. By regulating lipid uptake and metabolism in renal cells, FXR may help prevent renal injury associated with metabolic disorders [132]. Furthermore, FXR agonists have demonstrated potential in mitigating inflammatory responses within the kidneys, thereby offering a therapeutic avenue for addressing CKD related to MASLD [134].

## Therapeutic applications of FXR agonists

FXR has emerged as a potential therapeutic target for the management of metabolic syndrome and liver disorders, particularly MASLD. The therapeutic potential of FXR agonists is being explored to mitigate the adverse effects associated with MASLD, MASH, and other metabolic disorders, as clearly discussed in Figure 3 [135]. The ligands for the FXR receptor can be classified based on their chemical nature: natural agonists [chenodeoxy cholic acid (CDCA), cholic acid, ursodeoxycholic acid (UDCA)], semisynthetic bile acids (OCA), and synthetic non-steroidal agonists (GW-4064 and WAY 362450) [136]. FXR agonists, such as OCA, have proven effective in reducing hepatic steatosis, enhancing insulin sensitivity, and providing anti-inflammatory effects in clinical trials. FXR agonists, such as OCA, have proven effective in reducing hepatic steatosis, enhancing insulin sensitivity, and providing anti-inflammatory effects in clinical trials. These agents work by restoring normal FXR signalling pathways that regulate lipid metabolism and inflammation [123]. Deoxycholic acid (DCA) and lithocholic acid (LCA) can activate FXR, albeit with lower efficiency. UDCA is a partial agonist inhibiting CDCA activity [137, 138]. FXR agonists have been used recently for the improvement of ailments like MASH, diabetes mellitus, and primary biliary cholangitis (PBC) [139]. OCA, developed by Intercept Pharmaceuticals, is one of the earlier FXR agonists, and its regenerate phase III trial demonstrated reproducibility with phase II data. However, the FDA denied approval for its use in MASH and has currently banned it from phase III clinical trials due to severe side effects, including pruritus; aside from pruritus, OCA was generally well-tolerated [140]. Notably, there was a reduction in HDL cholesterol levels. The U.S. FDA approved OCA for second-line treatment of PBC in 2016 [123, 141].

Clinical data of patients and rodent studies using UDCA showed that the benefits were more effective than clofibrate ones. Small trials showed elevated levels of enzymes, which generally show an increase during liver diseases (alanine aminotransferase, aspartate aminotransferase, gamma-glutamyl transferase, and alkaline phosphatases). These are reduced in the treatment [142]. However, norUDCA, a shortened UDCA derivative, gave promising results in phase II dosing trials in mice models [143]. MET-409, a non-steroidal FXR agonist currently in phase IIa clinical trial, is different in structure from bile acids and has an entirely different chemical nature from other FXR agonists (tropifexor and cilofexor) [144]. The agonist showed potent liver fat reduction, but a transient increase in alanine aminotransferase and FGF-19 was observed [145]. The regulation of fatty acid influx into the hepatic plasma membrane is governed by specialized transporters such as FATP, CD36, and caveolins [146, 147]. Depletion of FATP2, a specific isoform of FATP, has been demonstrated to ameliorate hepatic steatosis and diminish the uptake of fatty acids, suggesting a potential therapeutic avenue for mitigating excessive lipid accumulation in the liver [148]. The transportation of elongated fatty acids is facilitated by CD36, a mechanism regulated by PPAR-gamma [149]. There exists a clear association between the expression of CD36 and the concentration of TGs within hepatic tissue [150]. Investigations carried out in obese murine models propose that administration of GW4064 induces a subsequent decline in the expression of CD36 at a molecular scale, thereby reducing circulating lipid levels [151]. Caveolin functions in the formation of lipid droplets by associating fatty acids with binding proteins (FABP) [152]. Vitamin D receptor-interacting protein 205 (DRIP205), coactivator-associated arginine methyltransferase-1 (CARM-1), and protein arginine methyltransferase 1 (PRMT-1) are some of the secondary coactivators that play a role in FXR-mediated transcription [153]. CARM-1 acts by its protein methylase activity, known to interact indirectly with nuclear receptors of the p160 family. Steroid receptor coactivator 1 (SRC-1), one of the nuclear receptors of the p160 family, acts on RXR-FXR heterodimers. Peroxisome proliferator-activated receptor-gamma coactivator-1 alpha (PGC-1 alpha), a





**Figure 3. The role of FXR agonists in restoring liver health.** This visual emphasizes the critical role of FXR agonists as promising therapeutic agents in managing liver diseases, showcasing their ability to restore liver health and improve overall metabolic function. OCA: obeticholic acid; MASLD: metabolic dysfunction-associated steatotic liver disease; NASH: non-alcoholic steatohepatitis. Created in BioRender. Pirangi, S. (2025) [BioRender.com/u291065](https://BioRender.com/u291065)

well-known coactivator of PPAR-gamma, shows more FXR coactivating properties than SRC-1. FXR activates PGC-1 alpha by binding to the NH<sub>2</sub> position [154–156].

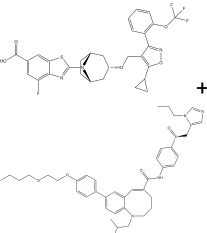
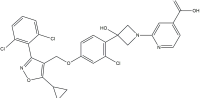
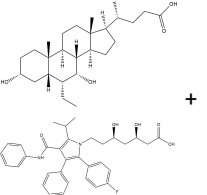
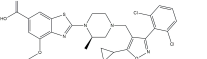
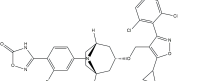
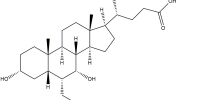
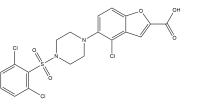
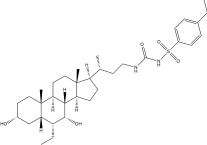
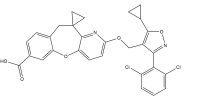
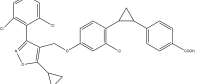
### Clinical trials and US FDA-approved drugs

FXR is crucial in countering these pathological conditions, as it exerts effects on each of them. Here are a few clinical trial drugs and ligands targeting FXR, with treatment mentioned in Table 2. Rezdiffra (resmetirom), a novel thyroid hormone receptor beta (THRβ) agonist, was first approved for treating adults with noncirrhotic, MASH with moderate to advanced liver scarring [157]. Combined with diet and exercise, this drug reduces liver fat accumulation by selectively stimulating THRβ in the liver. However, Rezdiffra is associated with several side effects like pruritus, diarrhoea, nausea, vomiting, constipation, abdominal pain, and dizziness are commonly reported. Additionally, it may cause liver injury (hepatotoxicity) and gallbladder-related complications such as gallstones (cholelithiasis) and inflammation of the gallbladder (cholecystitis). Therefore, patients undergoing treatment with Rezdiffra should be closely monitored [158]. Due to the numerous severe side effects associated with Rezdiffra, there is an urgent need for new therapeutic options for MASLD. Given these concerns, FXR agonist drugs emerge as a promising alternative to fulfil this requirement, offering a potential pathway for safer and more effective treatment strategies for MASLD while minimising adverse effects.

### Exploring lipid diversity: a new frontier in understanding MASLD

Lipidomics is an emerging field that provides a comprehensive analysis of lipid compositions in biological samples, offering valuable insights into MASLD [159]. Studies of liver biopsies from MASLD patients have revealed significant alterations in lipid profiles, particularly in glycerophospholipids and sphingolipids. Notably, an increased *n*-6/*n*-3 fatty acid ratio and decreased levels of essential long-chain PUFAs, such as arachidonic and EPA, have been observed, indicating a shift in lipid metabolism that contributes to disease progression [53, 160]. In addition to liver tissue analysis, lipidomic studies of plasma samples have identified specific lipid changes that may serve as non-invasive biomarkers for disease progression. For

**Table 2. Represents FXR agonist drugs for the MASLD/MASH under clinical trial phase II/III trails**

Primary mechanism	Agent (trial name)	Structure	Clinical trials	NCT number (clinicaltrials.gov)	Reference
FXR agonist and a chemokine receptor type 2/5 antagonist	LJN452 and CVC (ceniciviroc)		Phase 2b	NCT03517540	[174, 175]
FXR agonist	Cilofexor (GS-9674)		Phase 2	NCT02854605	[176]
FXR agonist and statin	Obeticholic acid and atorvastatin		Phase 2	NCT02633956	[177]
Non bile acid FXR agonist	HPG1860		Phase 2a	NCT05338034	[178]
FXR agonist	CS0159		Phase 2	NCT05591079	[179]
FXR agonist	Obeticholic acid		Phase 3	NCT03439254	[180]
FXR agonist	EYP001a (vonafexor)		Phase 2	NCT03812029	[181]
FXR agonist	EDP-305		Phase 2	NCT03421431	[182]
Tricyclic FXR agonist	HEC96719		Phase 2	NCT05397379	[183]
Non-steroidal FXR agonist	PX-104		Phase 2	NCT01999101	[184]

FXR: farnesoid X receptor; MASLD: metabolic dysfunction-associated steatotic liver disease; MASH: metabolic dysfunction-associated steatohepatitis

example, significant alterations in phosphatidylcholine (PC), phosphatidylethanolamine (PE), and sphingomyelin levels have been documented between healthy individuals and those with varying stages of MASLD, including MASH as detailed described in Table 3 [161, 162]. These lipid changes highlight the critical role of lipid metabolism in the development and progression of MASLD. The accumulation of lipids within hepatocytes leads to increased metabolic demands, resulting in the production of ROS and oxidative stress [163]. This process is exacerbated by obesity-related expansion of adipose tissue and insulin resistance, which promote the release of FFAs and activate inflammatory pathways. Elevated levels of ceramides a class of sphingolipids have been linked to lipotoxicity and hepatic inflammation, further contributing to the pathogenesis of MASLD [34].

**Table 3. Quantification of fatty acids by using various analytical techniques**

Authors name	Disease condition analysed	Metabolites altered	Analytical technique	Healthy vs. NASH patients
Masoodi et al. [185]	MASLD and MASH	Amino acids, fatty acids, triglycerides, phospholipids, and bile acids	Metabolomics and lipidomic by MS	Altered amino acid metabolism, and lipid disturbances
Kartsoli et al. [186]	MASLD	Liver lipid species	Lipidomic approach by LC-MS/MS	Impact of liver lipid species on MASH development and progression
Garcia-Jaramillo et al. [187]	MASH	Hepatic neutral and membrane lipids	Lipidomic analysis by UPLC-TOF-MS/MS	Profound alterations in lipid composition
Wang et al. [188]	MASLD and MASH	The main types of lipids are glycerides, glycerophospholipids, sphingolipids, fatty acyl lipids, and sterol lipids.	Lipidomic profiling by UPLC-MS/MS GC-FID	Differential lipid profiles in urine samples
Piras et al. [189]	MASLD	Circulating lipid biomarkers	LC-MS	Potential biomarkers for MASLD
Tan et al. [190]	MASLD and MASH	Circulating fatty acids, triglycerides, phospholipids, and bile acids	Metabolomics and lipidomics by imaging particle detector	Altered metabolic pathways
Kalopitas et al. [191]	MASLD and MASH	Potential differences in plasma lipids	Lipidomic profiling by LC-MS and GC-MS	Comparison in MASH, MASLD, and healthy
Zhu et al. [192]	NAFL and NASH	Urinary extracellular vesicles	Lipidomic analysis by mass-based approach	Lipidomic changes as potential markers
Gaggini et al. [193]	NAFLD	Circulating lipids	Metabolomics by mass-based approach	Links between lipids and NAFLD

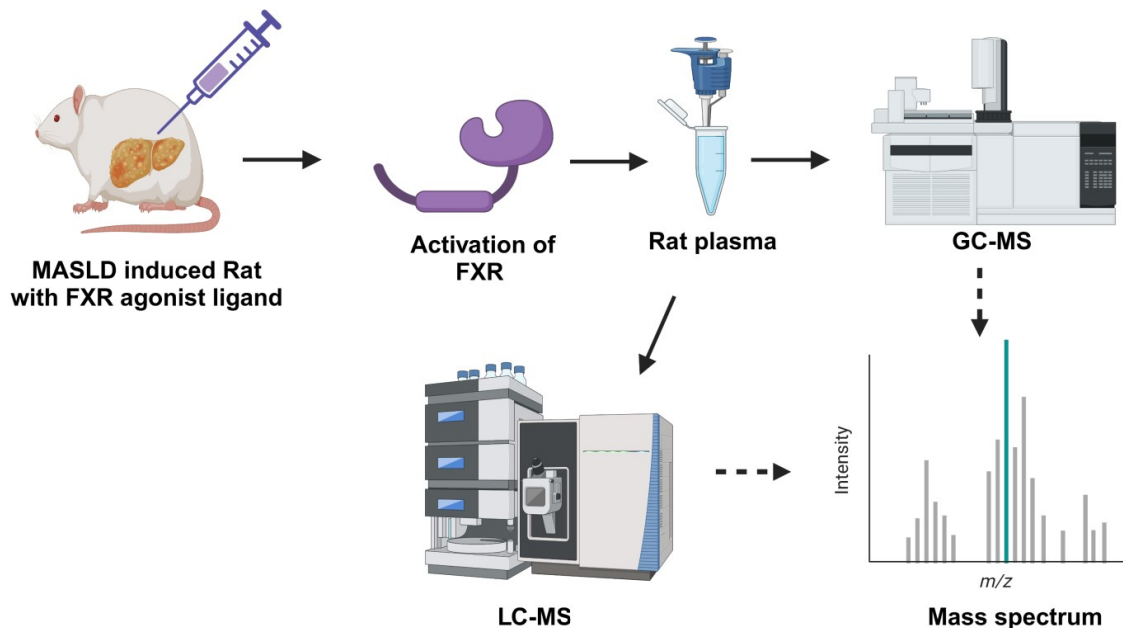
NASH: non-alcoholic steatohepatitis; MASLD: metabolic dysfunction-associated steatotic liver disease; LC-MS: liquid chromatography-mass spectrometry; UPLC-TOF-MS: ultra-performance liquid chromatography time-of-flight mass spectrometry; UPLC-MS: ultra-performance liquid chromatography-mass spectrometry; GC-FID: gas chromatography flame ionization detector; GC-MS: gas chromatography-mass spectrometry; NAFL: non-alcoholic fatty liver; NAFLD: non-alcoholic fatty liver disease

To analyse these complex lipid profiles, researchers utilize hyphenated techniques such as liquid chromatography-mass spectrometry (LC-MS) and gas chromatography-mass spectrometry (GC-MS) as represented in Figure 4. These methodologies enable the identification and quantification of thousands of lipid species, enhancing our understanding of their roles in liver disease [164–166]. Lipidomic profiling is crucial for uncovering the specific lipid alterations associated with MASLD [60, 167]. By identifying potential biomarkers for distinguishing between MASLD and MASH, lipidomics offers promising avenues for early diagnosis and targeted therapeutic strategies aimed at restoring lipid homeostasis and improving patient outcomes [168].

## Conclusions

In this review, we have clearly articulated the renaming of NAFLD to MASLD and NASH to MASH. We explored the association of MASLD with features of metabolic syndrome, emphasizing the central role of fatty acids in disease development and progression. Additionally, we examined the role of FXR in both physiological and pathological conditions, detailing how hepatic steatosis can progress to severe outcomes like fibrosis and CKD. The therapeutic potential of FXR agonists in treating MASLD and other metabolic diseases was discussed, alongside a review of clinical trial drugs targeting the FXR receptor. Furthermore, we highlighted the importance of lipidomic studies for quantifying fatty acids responsible for MASLD and MASH pathogenesis using advanced analytical techniques.

Despite these insights, significant research gaps remain due to the complex pathology of MASLD. Over 30% of the global population is affected by MAFLD, highlighting this silent epidemic's growing prevalence. Current diagnostic techniques are limited, necessitating improvements in lipidomics through advanced methods like LC-MS/MS and GC-MS/MS to accurately assess disease severity. Optimizing sample preparation protocols and minimizing matrix effects are essential steps forward. Currently, only one drug,



**Figure 4. Quantification of lipid profiles in MASLD-induced rats treated with FXR agonists.** This visual emphasizes the potential of FXR agonist therapy as a promising intervention for MASLD and illustrates the critical role of lipid profiling in monitoring disease progression and therapeutic outcomes. MASLD: metabolic dysfunction-associated steatotic liver disease; FXR: farnesoid X receptor; GC-MS: gas chromatography-mass spectrometry; LC-MS: liquid chromatography-mass spectrometry. Created in BioRender. Pirangi, S. (2025) [BioRender.com/x10t173](https://BioRender.com/x10t173)

resmetirom, has been approved by the FDA for MASLD treatment. While FXR agonists show promise in mitigating disease pathology, side effects have hindered others, such as OCA, from progressing to phase III trials. There is a pressing need for early-stage identification of MASLD and further advancements in lipidomic studies, alongside the development of new therapeutic options to address this widespread health issue. By pursuing these avenues, we can enhance our understanding and management of MASLD, ultimately improving patient outcomes in this prevalent liver disease.

## Abbreviations

CARM-1: coactivator-associated arginine methyltransferase-1

CAV-1: caveolin-1

CD36: cluster of differentiation 36

CDCA: chenodeoxy cholic acid

ChREBP: carbohydrate response element-binding protein

CKD: chronic kidney disease

DAG: diacylglycerol

FASN: fatty acid synthase

FATPs: fatty acid transport proteins

FFAs: free fatty acids

FGF-15: fibroblast growth factor-15

FXR: farnesoid X receptor

FXREs: farnesoid X receptor response elements

GC-FID: gas chromatography flame ionization detector

GC-MS: gas chromatography-mass spectrometry

HCC: hepatocellular carcinoma

HDL: high-density lipoprotein  
LC-MS: liquid chromatography-mass spectrometry  
LDL: low-density lipoprotein  
MASH: metabolic associated-steatotic hepatitis  
MASLD: metabolic-associated steatotic liver diseases  
MS: mass spectrometry  
MUFA: monounsaturated fatty acids  
NAFL: non-alcoholic fatty liver  
NAFLD: non-alcoholic fatty liver disease  
NASH: non-alcoholic steatohepatitis  
OCA: obeticholic acid  
PBC: primary biliary cholangitis  
PPAR $\alpha$ : peroxisome proliferator-activated receptor alpha  
PUFAs: polyunsaturated fatty acids  
RXR: retinoid X receptor  
SCD: stearoyl-CoA desaturase  
SFA: saturated fatty acids  
SHP: small heterodimer partner  
SRC-1: steroid receptor coactivator 1  
SREBP-1c: sterol regulatory element-binding protein-1c  
T2DM: type 2 diabetes mellitus  
TAG: triacylglycerol  
TGs: triglycerides  
THR $\beta$ : thyroid hormone receptor beta  
UDCA: ursodeoxycholic acid  
UFA: unsaturated fatty acids  
VLDL: very low-density lipoprotein

## **Declarations**

### **Acknowledgments**

The authors sincerely thank NIPER, S.A.S Nagar, and the Department of Pharmaceuticals, Ministry of Chemicals and Fertilizers, India, for providing infrastructure and Ph.D. fellowship.

### **Author contributions**

PS: Conceptualization, Investigation, Writing—original draft, Writing—review & editing. KMS, VP, and DK: Writing—original draft, Writing—review & editing. SN: Conceptualization, Investigation, Validation, Writing—review & editing, Supervision.

### **Conflicts of interest**

The authors declare that there are no conflicts of interest.

### Ethical approval

Not applicable.

### Consent to participate

Not applicable.

### Consent to publication

Not applicable.

### Availability of data and materials

Not applicable.

### Funding

Not applicable.

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## References

1. Rinella ME, Lazarus JV, Ratziu V, Francque SM, Sanyal AJ, Kanwal F, et al.; NAFLD Nomenclature consensus group. A multisociety Delphi consensus statement on new fatty liver disease nomenclature. *Hepatology*. 2023;78:1966–86. [DOI] [PubMed] [PMC]
2. Kuchay MS, Misra A. From non-alcoholic fatty liver disease (NAFLD) to metabolic-associated fatty liver disease (MAFLD): A journey over 40 years. *Diabetes Metab Syndr*. 2020;14:695–6. [DOI] [PubMed]
3. Dale K, Fallouh Y, Alkhouri N. MASLD and MASH: how a change of nomenclature may impact our approach in treating liver disease. *Expert Opin Investig Drugs*. 2024;33:1095–7. [DOI] [PubMed]
4. European Association for the Study of the Liver (EASL); {European Association for the Study of Diabetes (EASD)}; {European Association for the Study of Obesity (EASO)}. EASL-EASD-EASO Clinical Practice Guidelines on the management of metabolic dysfunction-associated steatotic liver disease (MASLD). *J Hepatol*. 2024;81:492–542. [DOI] [PubMed]
5. New MASLD Nomenclature [Internet]. American Association for the Study of Liver Diseases; c2025 [cited 2024 Dec 15]. Available from: <https://www.aasld.org/new-masld-nomenclature>
6. Younossi ZM, Kalligeros M, Henry L. Epidemiology of Metabolic Dysfunction-Associated Steatotic Liver Disease. *Clin Mol Hepatol*. 2024. [DOI] [PubMed]
7. Dabbah S, Mishani I, Davidov Y, Ben Ari Z. Implementation of Machine Learning Algorithms to Screen for Advanced Liver Fibrosis in Metabolic Dysfunction-Associated Steatotic Liver Disease: An In-Depth Explanatory Analysis. *Digestion*. 2024. [DOI] [PubMed]
8. Targher G, Byrne CD, Tilg H. MASLD: a systemic metabolic disorder with cardiovascular and malignant complications. *Gut*. 2024;73:691–702. [DOI] [PubMed]
9. Jan B, Dar MI, Choudhary B, Basist P, Khan R, Alhalmi A. Cardiovascular Diseases Among Indian Older Adults: A Comprehensive Review. *Cardiovasc Ther*. 2024;2024:6894693. [DOI] [PubMed] [PMC]
10. Murtaza G, Riaz S, Zafar M, Ahsan Raza M, Kaleem I, Imran H, et al. Examining the growing challenge: Prevalence of diabetes in young adults (Review). *Med Int (Lond)*. 2024;5:2. [DOI] [PubMed] [PMC]

11. Marchesini G, Vettor R, Pinzani M. MASLD emerging from the fog of fatty liver. *J Hepatol.* 2024;80:178–80. [DOI] [PubMed]
12. Miao L, Targher G, Byrne CD, Cao YY, Zheng MH. Current status and future trends of the global burden of MASLD. *Trends Endocrinol Metab.* 2024;35:697–707. [DOI] [PubMed]
13. Kozlitina J, Sookoian S. Global Epidemiological Impact of *PNPLA3* I148M on Liver Disease. *Liver Int.* 2025;45:e16123. [DOI] [PubMed] [PMC]
14. Brouwers B, Rao G, Tang YY, Rodríguez Á, Glass LC, Hartman ML. Incretin-based investigational therapies for the treatment of MASLD/MASH. *Diabetes Research and Clinical Practice.* 2024;211:111675. [DOI]
15. Soto A, Spongberg C, Martinino A, Giovinazzo F. Exploring the Multifaceted Landscape of MASLD: A Comprehensive Synthesis of Recent Studies, from Pathophysiology to Organoids and Beyond. *Biomedicines.* 2024;12:397. [DOI] [PubMed] [PMC]
16. Koenig AB, Tan A, Abdelaal H, Monge F, Younossi ZM, Goodman ZD. Review article: Hepatic steatosis and its associations with acute and chronic liver diseases. *Aliment Pharmacol Ther.* 2024;60:167–200. [DOI] [PubMed]
17. Reid MV, Fredrickson G, Mashek DG. Mechanisms coupling lipid droplets to MASLD pathophysiology. *Hepatology.* 2024. [DOI] [PubMed]
18. Xi Y, Li H. Role of farnesoid X receptor in hepatic steatosis in nonalcoholic fatty liver disease. *Biomed Pharmacother.* 2020;121:109609. [DOI] [PubMed]
19. Chen HQ. Nutrient mTORC1 signaling contributes to hepatic lipid metabolism in the pathogenesis of non-alcoholic fatty liver disease. *Liver Research.* 2020;4:15–22. [DOI]
20. Lee Y, Choi D, Park J, Kim JG, Choi T, Youn D. The Effects of Warm Acupuncture on the Expression of *AMPK* in High-Fat Diet-Induced MAFLD Rats. *Curr Issues Mol Biol.* 2024;46:11580–92. [DOI] [PubMed] [PMC]
21. Feng X, Zhang R, Yang Z, Zhang K, Xing J. Mechanism of Metabolic Dysfunction-associated Steatotic Liver Disease: Important role of lipid metabolism. *J Clin Transl Hepatol.* 2024;12:815–26. [DOI] [PubMed] [PMC]
22. New pipeline of drug candidates offers hope for effective MASLD treatments [Internet]. Xia & He Publishing Inc.; c2024 [cited 2024 Dec 22]. Available from: <https://www.news-medical.net/news/20240925/New-pipeline-of-drug-candidates-offers-hope-for-effective-MASLD-treatments.aspx>
23. Long J, Xu Y, Zhang X, Wu B, Wang C. Role of FXR in the development of NAFLD and intervention strategies of small molecules. *Arch Biochem Biophys.* 2024;757:110024. [DOI] [PubMed]
24. Kumar J, Hasan M, Mohsin S, Alzaher, MH, Nagar T, Jamil A, et al. Assessing the efficacy of farnesoid X receptor agonists in the management of metabolic dysfunction-associated steatotic liver disease: A systematic review and meta-analysis: Efficacy of Farnesoid X Receptor Agonists in Metabolic Dysfunction-associated Steatotic Liver Disease: Systematic Review and Meta-analysis. *Clinics and Research in Hepatology and Gastroenterology.* 2025;49:02530. [DOI]
25. Chen J, Wang R, Xiong F, Sun H, Kemper B, Li W, et al. Hammerhead-type FXR agonists induce an enhancer RNA *Fincor* that ameliorates nonalcoholic steatohepatitis in mice. *Elife.* 2024;13:RP91438. [DOI] [PubMed] [PMC]
26. Ali AH, Carey EJ, Lindor KD. Recent advances in the development of farnesoid X receptor agonists. *Ann Transl Med.* 2015;3:5. [DOI] [PubMed] [PMC]
27. Hollenback D, Hambruch E, Fink G, Birkel M, Schulz A, Hornberger M, et al. Development of Cilofexor, an Intestinally-Biased Farnesoid X Receptor Agonist, for the Treatment of Fatty Liver Disease. *J Pharmacol Exp Ther.* 2024;389:61–75. [DOI] [PubMed]
28. Sanyal AJ, Lopez P, Lawitz EJ, Lucas KJ, Loeffler J, Kim W, et al. Tropifexor for nonalcoholic steatohepatitis: an adaptive, randomized, placebo-controlled phase 2a/b trial. *Nat Med.* 2023;29:392–400. [DOI] [PubMed] [PMC]

29. Adorini L, Trauner M. FXR agonists in NASH treatment. *J Hepatol.* 2023;79:1317–31. [DOI] [PubMed]
30. Branković M, Dukić M, Gmizić T, Popadić V, Nikolić N, Sekulić A, et al. New Therapeutic Approaches for the Treatment of Patients with Metabolic Dysfunction-Associated Steatotic Liver Disease (MASLD) and Increased Cardiovascular Risk. *Diagnostics (Basel).* 2024;14:229. [DOI] [PubMed] [PMC]
31. Antwi MB, Jennings A, Lefere S, Clarisse D, Geerts A, Devisscher L, et al. Unlocking therapeutic potential: exploring cross-talk among emerging nuclear receptors to combat metabolic dysfunction in steatotic liver disease. *npj Metab Health Dis.* 2024;2:13. [DOI]
32. Sandireddy R, Sakthivel S, Gupta P, Behari J, Tripathi M, Singh BK. Systemic impacts of metabolic dysfunction-associated steatotic liver disease (MASLD) and metabolic dysfunction-associated steatohepatitis (MASH) on heart, muscle, and kidney related diseases. *Front Cell Dev Biol.* 2024;12: 1433857. [DOI] [PubMed] [PMC]
33. Ma Y, Wang J, Xiao W, Fan X. A review of MASLD-related hepatocellular carcinoma: progress in pathogenesis, early detection, and therapeutic interventions. *Front Med (Lausanne).* 2024;11: 1410668. [DOI] [PubMed] [PMC]
34. Schwärzler J, Grabherr F, Grander C, Adolph TE, Tilg H. The pathophysiology of MASLD: an immunometabolic perspective. *Expert Rev Clin Immunol.* 2024;20:375–86. [DOI] [PubMed]
35. Lim S, Kim J, Targher G. Links between metabolic syndrome and metabolic dysfunction-associated fatty liver disease. *Trends Endocrinol Metab.* 2021;32:500–14. [DOI] [PubMed]
36. Ma C, Yan K, Wang Z, Zhang Q, Gao L, Xu T, et al. The association between hypertension and nonalcoholic fatty liver disease (NAFLD): literature evidence and systems biology analysis. *Bioengineered.* 2021;12:2187–202. [DOI] [PubMed] [PMC]
37. Chan WK, Chuah KH, Rajaram RB, Lim LL, Ratnasingam J, Vethakkan SR. Metabolic Dysfunction-Associated Steatotic Liver Disease (MASLD): A State-of-the-Art Review. *J Obes Metab Syndr.* 2023; 32:197–213. [DOI] [PubMed] [PMC]
38. Chew NWS, Pan XH, Chong B, Chandramouli C, Muthiah M, Lam CSP. Type 2 diabetes mellitus and cardiometabolic outcomes in metabolic dysfunction-associated steatotic liver disease population. *Diabetes Research and Clinical Practice.* 2024;211:111652. [DOI] [PubMed]
39. Mellekjær A, Kjær MB, Haldrup D, Grønbaek H, Thomsen KL. Management of cardiovascular risk in patients with metabolic dysfunction-associated steatotic liver disease. *Eur J Intern Med.* 2024;122: 28–34. [DOI] [PubMed]
40. Wondmkun YT. Obesity, Insulin Resistance, and Type 2 Diabetes: Associations and Therapeutic Implications. *Diabetes Metab Syndr Obes.* 2020;13:3611–6. [DOI] [PubMed] [PMC]
41. Ahmed B, Sultana R, Greene MW. Adipose tissue and insulin resistance in obese. *Biomed Pharmacother.* 2021;137:111315. [DOI] [PubMed]
42. Roden M, Petersen KF, Shulman GI. Insulin Resistance in Type 2 Diabetes. *Textbook of Diabetes.* 2024. [DOI]
43. Petrie E, Gray M, Bril F. Metabolic characteristics of patients with MetALD: Caveats of a new definition. *Liver Int.* 2024;44:2929–38. [DOI] [PubMed]
44. Sakurai Y, Kubota N, Yamauchi T, Kadowaki T. Role of Insulin Resistance in MAFLD. *Int J Mol Sci.* 2021;22:4156. [DOI] [PubMed] [PMC]
45. Jiang S, Young JL, Wang K, Qian Y, Cai L. Diabetic-induced alterations in hepatic glucose and lipid metabolism: The role of type 1 and type 2 diabetes mellitus (Review). *Mol Med Rep.* 2020;22: 603–11. [DOI] [PubMed] [PMC]
46. Kalopitas G, Arvanitakis K, Tsachouridou O, Malandris K, Koufakis T, Metallidis S, et al. Metabolic Dysfunction-Associated Steatotic Liver Disease in People Living with HIV-Limitations on Antiretroviral Therapy Selection. *Life (Basel).* 2024;14:742. [DOI] [PubMed] [PMC]



47. Yang C, He Q, Chen Z, Qin J, Lei F, Liu Y, et al. A Bidirectional Relationship Between Hyperuricemia and Metabolic Dysfunction-Associated Fatty Liver Disease. *Front Endocrinol (Lausanne)*. 2022;13: 821689. [DOI] [PubMed] [PMC]
48. Muzica CM, Sfarti C, Trifan A, Zenovia S, Cuciureanu T, Nastasa R, et al. Nonalcoholic Fatty Liver Disease and Type 2 Diabetes Mellitus: A Bidirectional Relationship. *Can J Gastroenterol Hepatol*. 2020;2020:6638306. [DOI] [PubMed] [PMC]
49. Lekakis V, Papatheodoridis GV. Natural history of metabolic dysfunction-associated steatotic liver disease. *Eur J Intern Med*. 2024;122:3–10. [DOI] [PubMed]
50. Zheng H, Sechi LA, Navarese EP, Casu G, Vidili G. Metabolic dysfunction-associated steatotic liver disease and cardiovascular risk: a comprehensive review. *Cardiovasc Diabetol*. 2024;23:346. [DOI] [PubMed] [PMC]
51. Habib S. Team players in the pathogenesis of metabolic dysfunctions-associated steatotic liver disease: The basis of development of pharmacotherapy. *World J Gastrointest Pathophysiol*. 2024;15: 93606. [DOI] [PubMed] [PMC]
52. Rao G, Peng X, Li X, An K, He H, Fu X, et al. Unmasking the enigma of lipid metabolism in metabolic dysfunction-associated steatotic liver disease: from mechanism to the clinic. *Front Med (Lausanne)*. 2023;10:1294267. [DOI] [PubMed] [PMC]
53. Carli F, Della Pepa G, Sabatini S, Vidal Puig A, Gastaldelli A. Lipid metabolism in MASLD and MASH: From mechanism to the clinic. *JHEP Rep*. 2024;6:101185. [DOI] [PubMed] [PMC]
54. Silveira Rossi JL, Barbalho SM, Reverete de Araujo R, Bechara MD, Sloan KP, Sloan LA. Metabolic syndrome and cardiovascular diseases: Going beyond traditional risk factors. *Diabetes Metab Res Rev*. 2022;38:e3502. [DOI] [PubMed]
55. Ziolkowska S, Binienda A, Jabłkowski M, Szemraj J, Czarny P. The Interplay between Insulin Resistance, Inflammation, Oxidative Stress, Base Excision Repair and Metabolic Syndrome in Nonalcoholic Fatty Liver Disease. *Int J Mol Sci*. 2021;22:11128. [DOI] [PubMed] [PMC]
56. Ju-Seop K, editor. *Non-alcoholic Fatty Liver Disease*. London: IntechOpen; 2023.
57. Petersen MC, Shulman GI. Roles of Diacylglycerols and Ceramides in Hepatic Insulin Resistance. *Trends Pharmacol Sci*. 2017;38:649–65. [DOI] [PubMed] [PMC]
58. Schilcher K, Dayoub R, Kubitzka M, Riepl J, Klein K, Buechler C, et al. Saturated Fat-Mediated Upregulation of IL-32 and CCL20 in Hepatocytes Contributes to Higher Expression of These Fibrosis-Driving Molecules in MASLD. *Int J Mol Sci*. 2023;24:13222. [DOI] [PubMed] [PMC]
59. Wang X, Zhang L, Dong B. Molecular mechanisms in MASLD/MASH-related HCC. *Hepatology*. 2024; 10.1097/HEP.0000000000000786. [DOI] [PubMed]
60. Núñez-Sánchez MÁ, Martínez-Sánchez MA, Martínez-Montoro JI, Balaguer-Román A, Murcia-García E, Fernández-Ruiz VE, et al. Lipidomic Analysis Reveals Alterations in Hepatic FA Profile Associated With MASLD Stage in Patients With Obesity. *The Journal of Clinical Endocrinology & Metabolism*. 2024;109:1781–92. [DOI]
61. Deng KQ, Huang X, Lei F, Zhang XJ, Zhang P, She ZG, et al. Role of hepatic lipid species in the progression of nonalcoholic fatty liver disease. *Am J Physiol Cell Physiol*. 2022;323:C630–9. [DOI] [PubMed]
62. Valenzuela R, Ortiz M, Hernández-Rodas MC, Echeverría F, Videla LA. Targeting n-3 Polyunsaturated Fatty Acids in Non-Alcoholic Fatty Liver Disease. *Curr Med Chem*. 2020;27:5250–72. [DOI] [PubMed]
63. Zhang W, Lin H, Cheng W, Huang Z, Zhang W. Protective Effect and Mechanism of Plant-Based Monoterpenoids in Non-alcoholic Fatty Liver Diseases. *J Agric Food Chem*. 2022;70:4839–59. [DOI] [PubMed]
64. Shang T, Liu L, Zhou J, Zhang M, Hu Q, Fang M, et al. Protective effects of various ratios of DHA/EPA supplementation on high-fat diet-induced liver damage in mice. *Lipids Health Dis*. 2017;16:65. [DOI] [PubMed] [PMC]

65. Ten Hove M, Pater L, Storm G, Weiskirchen S, Weiskirchen R, Lammers T, et al. The hepatic lipidome: From basic science to clinical translation. *Adv Drug Deliv Rev.* 2020;159:180–97. [DOI] [PubMed]
66. Alsabeeh N, Chausse B, Kakimoto PA, Kowaltowski AJ, Shirihai O. Cell culture models of fatty acid overload: Problems and solutions. *Biochim Biophys Acta Mol Cell Biol Lipids.* 2018;1863:143–51. [DOI] [PubMed] [PMC]
67. Stefan N, Yki-Järvinen H, Neuschwander-Tetri BA. Metabolic dysfunction-associated steatotic liver disease: heterogeneous pathomechanisms and effectiveness of metabolism-based treatment. *Lancet Diabetes Endocrinol.* 2025;13:134–48. [DOI] [PubMed]
68. Scavo MP, Negro R, Arrè V, Depalo N, Carrieri L, Rizzi F, et al. The oleic/palmitic acid imbalance in exosomes isolated from NAFLD patients induces necroptosis of liver cells via the elongase-6/RIP-1 pathway. *Cell Death Dis.* 2023;14:635. [DOI] [PubMed] [PMC]
69. Abdelhameed F, Kite C, Lagojda L, Dallaway A, Chatha KK, Chaggar SS, et al. Non-invasive Scores and Serum Biomarkers for Fatty Liver in the Era of Metabolic Dysfunction-associated Steatotic Liver Disease (MASLD): A Comprehensive Review From NAFLD to MAFLD and MASLD. *Curr Obes Rep.* 2024;13:510–31. [DOI] [PubMed] [PMC]
70. Verschuren L, Mak AL, van Koppen A, Özsezen S, Difrancesco S, Caspers MPM, et al. Development of a novel non-invasive biomarker panel for hepatic fibrosis in MASLD. *Nat Commun.* 2024;15:4564. [DOI] [PubMed] [PMC]
71. Thakral N, Desalegn H, Diaz LA, Cabrera D, Loomba R, Arrese M, et al. A Precision Medicine Guided Approach to the Utilization of Biomarkers in MASLD. *Semin Liver Dis.* 2024;44:273–86. [DOI] [PubMed]
72. Maki KC, Dicklin MR, Kirkpatrick CF. Saturated fats and cardiovascular health: Current evidence and controversies. *J Clin Lipidol.* 2021;15:765–72. [DOI] [PubMed]
73. Mei J, Qian M, Hou Y, Liang M, Chen Y, Wang C, et al. Association of saturated fatty acids with cancer risk: a systematic review and meta-analysis. *Lipids Health Dis.* 2024;23:32. [DOI] [PubMed] [PMC]
74. Duan H, Song W, Zhao J, Yan W. Polyunsaturated Fatty Acids (PUFAs): Sources, Digestion, Absorption, Application and Their Potential Adjunctive Effects on Visual Fatigue. *Nutrients.* 2023;15:2633. [DOI] [PubMed] [PMC]
75. Dyall SC, Balas L, Bazan NG, Brenna JT, Chiang N, da Costa Souza F, et al. Polyunsaturated fatty acids and fatty acid-derived lipid mediators: Recent advances in the understanding of their biosynthesis, structures, and functions. *Prog Lipid Res.* 2022;86:101165. [DOI] [PubMed] [PMC]
76. Clifford BL, Sedgeman LR, Williams KJ, Morand P, Cheng A, Jarrett KE, et al. FXR activation protects against NAFLD via bile-acid-dependent reductions in lipid absorption. *Cell Metab.* 2021;33:1671–84.e4. [DOI] [PubMed] [PMC]
77. Ma K, Saha PK, Chan L, Moore DD. Farnesoid X receptor is essential for normal glucose homeostasis. *J Clin Invest.* 2006;116:1102–9. [DOI] [PubMed] [PMC]
78. Zhang L, Wang YD, Chen WD, Wang X, Lou G, Liu N, et al. Promotion of liver regeneration/repair by farnesoid X receptor in both liver and intestine in mice. *Hepatology.* 2012;56:2336–43. [DOI] [PubMed] [PMC]
79. Jiang C, Xie C, Li F, Zhang L, Nichols RG, Krausz KW, et al. Intestinal farnesoid X receptor signaling promotes nonalcoholic fatty liver disease. *J Clin Invest.* 2015;125:386–402. [DOI] [PubMed] [PMC]
80. Mori H, Svegliati Baroni G, Marzioni M, Di Nicola F, Santori P, Maroni L, et al. Farnesoid X Receptor, Bile Acid Metabolism, and Gut Microbiota. *Metabolites.* 2022;12:647. [DOI] [PubMed] [PMC]
81. Chiang JYL, Ferrell JM. Discovery of farnesoid X receptor and its role in bile acid metabolism. *Mol Cell Endocrinol.* 2022;548:111618. [DOI] [PubMed] [PMC]
82. Ge MX, Shao RG, He HW. Advances in understanding the regulatory mechanism of cholesterol 7 $\alpha$ -hydroxylase. *Biochem Pharmacol.* 2019;164:152–64. [DOI] [PubMed]
83. Sinha RA. Targeting nuclear receptors for NASH/MASH: From bench to bedside. *Liver Res.* 2024;8:34–45. [DOI] [PubMed] [PMC]

84. Tang Y, Fan Y, Wang Y, Wang D, Huang Q, Chen T, et al. A Current Understanding of FXR in NAFLD: The multifaceted regulatory role of FXR and novel lead discovery for drug development. *Biomed Pharmacother.* 2024;175:116658. [DOI] [PubMed]
85. Wen YQ, Zou ZY, Zhao GG, Zhang MJ, Zhang YX, Wang GH, et al. FXR activation remodels hepatic and intestinal transcriptional landscapes in metabolic dysfunction-associated steatohepatitis. *Acta Pharmacol Sin.* 2024;45:2313–27. [DOI] [PubMed]
86. Li Y, Wang L, Yi Q, Luo L, Xiong Y. Regulation of bile acids and their receptor FXR in metabolic diseases. *Front Nutr.* 2024;11:1447878. [DOI] [PubMed] [PMC]
87. Kumari A, Pal Pathak D, Asthana S. Bile acids mediated potential functional interaction between FXR and FATP5 in the regulation of Lipid Metabolism. *Int J Biol Sci.* 2020;16:2308–22. [DOI] [PubMed] [PMC]
88. Panzitt K, Wagner M. FXR in liver physiology: Multiple faces to regulate liver metabolism. *Biochim Biophys Acta Mol Basis Dis.* 2021;1867:166133. [DOI] [PubMed]
89. Ni M, Zhang B, Zhao J, Feng Q, Peng J, Hu Y, et al. Biological mechanisms and related natural modulators of liver X receptor in nonalcoholic fatty liver disease. *Biomed Pharmacother.* 2019;113:108778. [DOI] [PubMed]
90. Li Z, Zheng D, Zhang T, Ruan S, Li N, Yu Y, et al. The roles of nuclear receptors in cholesterol metabolism and reverse cholesterol transport in nonalcoholic fatty liver disease. *Hepatol Commun.* 2023;8:e0343. [DOI] [PubMed] [PMC]
91. Yang Z, Danzeng A, Liu Q, Zeng C, Xu L, Mo J, et al. The Role of Nuclear Receptors in the Pathogenesis and Treatment of Non-alcoholic Fatty Liver Disease. *Int J Biol Sci.* 2024;20:113–26. [DOI] [PubMed] [PMC]
92. Fleishman JS, Kumar S. Bile acid metabolism and signaling in health and disease: molecular mechanisms and therapeutic targets. *Signal Transduct Target Ther.* 2024;9:97. [DOI] [PubMed] [PMC]
93. Li L, Xu S, Wang W, Li X, Wang H, Yang Q, et al. Bruceine A alleviates alcoholic liver disease by inhibiting AIM2 inflammasome activation via activating FXR. *Phytomedicine.* 2024;130:155693. [DOI] [PubMed]
94. Chen S, Sun S, Feng Y, Li X, Yin G, Liang P, et al. Diosgenin attenuates nonalcoholic hepatic steatosis through the hepatic FXR-SHP-SREBP1C/PPAR $\alpha$ /CD36 pathway. *Eur J Pharmacol.* 2023;952:175808. [DOI] [PubMed]
95. Wang Y, Xu H, Zhou X, Chen W, Zhou H. Dysregulated bile acid homeostasis: unveiling its role in metabolic diseases. *Med Rev (2021).* 2024;4:262–83. [DOI] [PubMed] [PMC]
96. Yu J, Huang J, Xia T, Li M, Li R. Molecular mechanisms of hepatic lipid metabolism disorders: Focus on mitochondrial quality control. *Port Hypertens Cirrhosis.* 2024;3:217–33. [DOI]
97. Liang Y, Qi J, Yu D, Wang Z, Li W, Long F, et al. Ferulic Acid Alleviates Lipid and Bile Acid Metabolism Disorders by Targeting FASN and CYP7A1 in Iron Overload-Treated Mice. *Antioxidants (Basel).* 2024;13:1277. [DOI] [PubMed] [PMC]
98. Ding C, Wang Z, Dou X, Yang Q, Ning Y, Kao S, et al. Farnesoid X receptor: From Structure to Function and Its Pharmacology in Liver Fibrosis. *Aging Dis.* 2024;15:1508–36. [DOI] [PubMed] [PMC]
99. Zhang L, Chen J, Yang X, Shen C, Huang J, Zhang D, et al. Hepatic *Zbtb18* (Zinc Finger and BTB Domain Containing 18) alleviates hepatic steatohepatitis via *FXR* (Farnesoid X Receptor). *Signal Transduct Target Ther.* 2024;9:20. [DOI] [PubMed] [PMC]
100. Lee LE, Doke T, Mukhi D, Susztak K. The key role of altered tubule cell lipid metabolism in kidney disease development. *Kidney Int.* 2024;106:24–34. [DOI] [PubMed]
101. Xu Y, Li F, Zalzal M, Xu J, Gonzalez FJ, Adorini L, et al. Farnesoid X receptor activation increases reverse cholesterol transport by modulating bile acid composition and cholesterol absorption in mice. *Hepatology.* 2016;64:1072–85. [DOI] [PubMed] [PMC]

102. Trapani L, Segatto M, Pallottini V. Regulation and deregulation of cholesterol homeostasis: The liver as a metabolic “power station”. *World J Hepatol.* 2012;4:184–90. [DOI] [PubMed] [PMC]
103. Goldstein JL, Brown MS. The LDL Receptor. *Arterioscler Thromb Vasc Biol.* 2009;29:431–8. [DOI] [PubMed] [PMC]
104. Zein AA, Kaur R, Hussein TOK, Graf GA, Lee JY. ABCG5/G8: a structural view to pathophysiology of the hepatobiliary cholesterol secretion. *Biochem Soc Trans.* 2019;47:1259–68. [DOI] [PubMed] [PMC]
105. Nakano T, Inoue I, Murakoshi T. A Newly Integrated Model for Intestinal Cholesterol Absorption and Efflux Reappraises How Plant Sterol Intake Reduces Circulating Cholesterol Levels. *Nutrients.* 2019; 11:310. [DOI] [PubMed] [PMC]
106. Yntema T, Eijgenraam TR, Kloosterhuis NJ, Havinga R, Koster MH, Hovingh MV, et al. The impact of a humanized bile acid composition on atherosclerosis development in hypercholesterolaemic *Cyp2c70* knockout mice. *Sci Rep.* 2025;15:2100. [DOI] [PubMed] [PMC]
107. Bhattacharya A, Taylor RE, Guo GL. *In vivo* mouse models to study bile acid synthesis and signaling. *Hepatobiliary Pancreat Dis Int.* 2023;22:466–73. [DOI] [PubMed] [PMC]
108. Honda A, Miyazaki T, Iwamoto J, Hirayama T, Morishita Y, Monma T, et al. Regulation of bile acid metabolism in mouse models with hydrophobic bile acid composition. *J Lipid Res.* 2020;61:54–69. [DOI] [PubMed] [PMC]
109. Deng S, Liu J, Niu C. HDL and Cholesterol Ester Transfer Protein (CETP). *Adv Exp Med Biol.* 2022; 1377:13–26. [DOI] [PubMed]
110. Ginsberg HN, Packard CJ, Chapman MJ, Borén J, Aguilar-Salinas CA, Averno M, et al. Triglyceride-rich lipoproteins and their remnants: metabolic insights, role in atherosclerotic cardiovascular disease, and emerging therapeutic strategies—a consensus statement from the European Atherosclerosis Society. *Eur Heart J.* 2021;42:4791–806. [DOI] [PubMed] [PMC]
111. Weinberg Sibony R, Segev O, Dor S, Raz I. Overview of oxidative stress and inflammation in diabetes. *J Diabetes.* 2024;16:e70014. [DOI]
112. Khoi CS, Lin TY, Chiang CK. Targeting Insulin Resistance, Reactive Oxygen Species, Inflammation, Programmed Cell Death, ER Stress, and Mitochondrial Dysfunction for the Therapeutic Prevention of Free Fatty Acid-Induced Vascular Endothelial Lipotoxicity. *Antioxidants (Basel).* 2024;13:1486. [DOI] [PubMed] [PMC]
113. Parola M, Pinzani M. Liver fibrosis in NAFLD/NASH: from pathophysiology towards diagnostic and therapeutic strategies. *Mol Aspects Med.* 2024;95:101231. [DOI] [PubMed]
114. Zhu Y, Liu H, Zhang M, Guo GL. Fatty liver diseases, bile acids, and FXR. *Acta Pharm Sin B.* 2016;6: 409–12. [DOI] [PubMed] [PMC]
115. FJanikiewicz J, Hanzelka K, Dziewulska A, Kozinski K, Dobrzyn P, Bernas T, et al. Inhibition of SCD1 impairs palmitate-derived autophagy at the step of autophagosome-lysosome fusion in pancreatic  $\beta$ -cells. *J Lipid Res.* 2015;56:1901–11. [DOI] [PubMed] [PMC]
116. Fiorucci S, Distrutti E, Carino A, Zampella A, Biagioli M. Bile acids and their receptors in metabolic disorders. *Prog Lipid Res.* 2021;82:101094. [DOI] [PubMed]
117. Vesković M, Šutulović N, Hrnčić D, Stanojlović O, Macut D, Mladenović D. The Interconnection between Hepatic Insulin Resistance and Metabolic Dysfunction-Associated Steatotic Liver Disease—The Transition from an Adipocentric to Liver-Centric Approach. *Curr Issues Mol Biol.* 2023;45: 9084–102. [DOI] [PubMed] [PMC]
118. Islam T. Targeting Liver Cell Metabolism and Function in Non-Alcoholic Fatty Liver Disease [dissertation]. Portugal: Universidade de Lisboa; 2021.
119. Palma R, Pronio A, Romeo M, Scognamiglio F, Ventriglia L, Ormando VM, et al. The Role of Insulin Resistance in Fueling NAFLD Pathogenesis: From Molecular Mechanisms to Clinical Implications. *J Clin Med.* 2022;11:3649. [DOI] [PubMed] [PMC]

120. Bansal SK, Bansal MB. Pathogenesis of MASLD and MASH - role of insulin resistance and lipotoxicity. *Aliment Pharmacol Ther.* 2024;59:S10–22. [DOI] [PubMed]
121. Ren N, Wang WF, Zou L, Zhao YL, Miao H, Zhao YY. The nuclear factor kappa B signaling pathway is a master regulator of renal fibrosis. *Front Pharmacol.* 2024;14:1335094. [DOI] [PubMed] [PMC]
122. Wang WL, Lian H, Liang Y, Ye Y, Tam PKH, Chen Y. Molecular Mechanisms of Fibrosis in Cholestatic Liver Diseases and Regenerative Medicine-Based Therapies. *Cells.* 2024;13:1997. [DOI] [PubMed] [PMC]
123. Wang K, Zhang Y, Wang G, Hao H, Wang H. FXR agonists for MASH therapy: Lessons and perspectives from obeticholic acid. *Med Res Rev.* 2024;44:568–86. [DOI] [PubMed]
124. Puengel T, Liu H, Guillot A, Heymann F, Tacke F, Peiseler M. Nuclear Receptors Linking Metabolism, Inflammation, and Fibrosis in Nonalcoholic Fatty Liver Disease. *Int J Mol Sci.* 2022;23:2668. [DOI] [PubMed] [PMC]
125. Yeo YH, Abdelmalek M, Khan S, Moylan CA, Rodriguez L, Villanueva A, et al. Current and emerging strategies for the prevention of hepatocellular carcinoma. *Nat Rev Gastroenterol Hepatol.* 2024. [DOI] [PubMed]
126. Huang P, Zhao H, Dai H, Li J, Pan X, Pan W, et al. FXR deficiency induced ferroptosis via modulation of the CBP-dependent p53 acetylation to suppress breast cancer growth and metastasis. *Cell Death Dis.* 2024;15:826. [DOI] [PubMed] [PMC]
127. Belka M, Gostyńska-Stawna A, Stawny M, Krajka-Kuźniak V. Activation of Nrf2 and FXR via Natural Compounds in Liver Inflammatory Disease. *Int J Mol Sci.* 2024;25:11213. [DOI] [PubMed] [PMC]
128. Anand S, Patel TN. Integrating the metabolic and molecular circuits in diabetes, obesity and cancer: a comprehensive review. *Discov Oncol.* 2024;15:779. [DOI] [PubMed] [PMC]
129. Attia YM, Tawfiq RA, Ali AA, Elmazar MM. The FXR Agonist, Obeticholic Acid, Suppresses HCC Proliferation & Metastasis: Role of IL-6/STAT3 Signalling Pathway. *Sci Rep.* 2017;7:12502. [DOI] [PubMed] [PMC]
130. Lonardo A. Association of NAFLD/NASH, and MAFLD/MASLD with chronic kidney disease: an updated narrative review. *Metab Target Organ Damage.* 2024;4:16. [DOI]
131. Rad NK, Heydari Z, Tamimi AH, Zahmatkesh E, Shpichka A, Barekat M, et al. Review on Kidney-Liver Crosstalk: Pathophysiology of Their Disorders. *Cell J.* 2024;26:98–111. [DOI] [PubMed] [PMC]
132. Rong J, Zhang Z, Peng X, Li P, Zhao T, Zhong Y. Mechanisms of hepatic and renal injury in lipid metabolism disorders in metabolic syndrome. *Int J Biol Sci.* 2024;20:4783–98. [DOI] [PubMed] [PMC]
133. Zhang X, Wu W, Li Y, Peng Z. Exploring the role and therapeutic potential of lipid metabolism in acute kidney injury. *Ren Fail.* 2024;46:2403652. [DOI] [PubMed] [PMC]
134. Ghose S, Satariano M, Korada S, Cahill T, Shah R, Raina R. Advancements in diabetic kidney disease management: integrating innovative therapies and targeted drug development. *Am J Physiol Endocrinol Metab.* 2024;326:E791–806. [DOI] [PubMed]
135. Almeqdadi M, Gordon FD. Farnesoid X Receptor Agonists: A Promising Therapeutic Strategy for Gastrointestinal Diseases. *Gastro Hep Adv.* 2023;3:344–52. [DOI] [PubMed] [PMC]
136. Petrescu AD, DeMorrow S. Farnesoid X Receptor as Target for Therapies to Treat Cholestasis-Induced Liver Injury. *Cells.* 2021;10:1846. [DOI] [PubMed] [PMC]
137. Sheng W, Ji G, Zhang L. The Effect of Lithocholic Acid on the Gut-Liver Axis. *Front Pharmacol.* 2022; 13:910493. [DOI] [PubMed] [PMC]
138. Kiriya Y, Nochi H. The Role of Gut Microbiota-Derived Lithocholic Acid, Deoxycholic Acid and Their Derivatives on the Function and Differentiation of Immune Cells. *Microorganisms.* 2023;11: 2730. [DOI] [PubMed] [PMC]
139. Radun R, Trauner M. Role of FXR in Bile Acid and Metabolic Homeostasis in NASH: Pathogenetic Concepts and Therapeutic Opportunities. *Semin Liver Dis.* 2021;41:461–75. [DOI] [PubMed] [PMC]

140. FDA Accepts Intercept's New Drug Application for OCA for the Treatment of Pre-Cirrhotic Liver Fibrosis Due to NASH [Internet]. Morristown: Intercept Pharmaceuticals, Inc.; c2024 [cited 2024 Sep 30]. Available from: <https://www.globenewswire.com/news-release/2023/01/19/2591748/23024/en/FDA-Accepts-Intercept-s-New-Drug-Application-for-OCA-for-the-Treatment-of-Pre-Cirrhotic-Liver-Fibrosis-Due-to-NASH.html>
141. Jiang L, Zhang H, Xiao D, Wei H, Chen Y. Farnesoid X receptor (FXR): Structures and ligands. *Comput Struct Biotechnol J*. 2021;19:2148–59. [DOI] [PubMed] [PMC]
142. Laurin J, Lindor KD, Crippin JS, Gossard A, Gores GJ, Ludwig J, et al. Ursodeoxycholic Acid or Clofibrate in the Treatment of Non-Alcohol-Induced Steatohepatitis: A Pilot Study. *Hepatology*. 1996; 23:1464–7. [DOI] [PubMed]
143. Halilbasic E, Steinacher D, Trauner M. Nor-Ursodeoxycholic Acid as a Novel Therapeutic Approach for Cholestatic and Metabolic Liver Diseases. *Dig Dis*. 2017;35:288–92. [DOI] [PubMed]
144. Harrison SA, Bashir MR, Lee KJ, Shim-Lopez J, Lee J, Wagner B, et al. A structurally optimized FXR agonist, MET409, reduced liver fat content over 12 weeks in patients with non-alcoholic steatohepatitis. *J Hepatol*. 2021;75:25–33. [DOI] [PubMed]
145. Kremoser C. FXR agonists for NASH: How are they different and what difference do they make? *J Hepatol*. 2021;75:12–5. [DOI] [PubMed]
146. Black PN, Sandoval A, Arias-Barrau E, DiRusso CC. Targeting the Fatty Acid Transport Proteins (FATP) to Understand the Mechanisms Linking Fatty Acid Transport to Metabolism. *Immunol Endocr Metab Agents Med Chem*. 2009;9:11–7. [DOI] [PubMed] [PMC]
147. Samovski D, Jacome-Sosa M, Abumrad NA. Fatty Acid Transport and Signaling: Mechanisms and Physiological Implications. *Annu Rev Physiol*. 2023;85:317–37. [DOI] [PubMed]
148. Falcon A, Doege H, Fluitt A, Tsang B, Watson N, Kay MA, et al. FATP2 is a hepatic fatty acid transporter and peroxisomal very long-chain acyl-CoA synthetase. *Am J Physiol Endocrinol Metab*. 2010;299:E384–93. [DOI] [PubMed] [PMC]
149. Li Y, Huang X, Yang G, Xu K, Yin Y, Brecchia G, et al. CD36 favours fat sensing and transport to govern lipid metabolism. *Prog Lipid Res*. 2022;88:101193. [DOI] [PubMed]
150. Chen Y, Zhang J, Cui W, Silverstein RL. CD36, a signaling receptor and fatty acid transporter that regulates immune cell metabolism and fate. *J Exp Med*. 2022;219:e20211314. [DOI] [PubMed] [PMC]
151. Singh N, Yadav M, Singh AK, Kumar H, Dwivedi SK, Mishra JS, et al. Synthetic FXR Agonist GW4064 Is a Modulator of Multiple G Protein-Coupled Receptors. *Mol Endocrinol*. 2014;28:659–73. [DOI] [PubMed] [PMC]
152. Dalton CM, Schlegel C, Hunter CJ. Caveolin-1: A Review of Intracellular Functions, Tissue-Specific Roles, and Epithelial Tight Junction Regulation. *Biology (Basel)*. 2023;12:1402. [DOI] [PubMed] [PMC]
153. Hwang JW, Cho Y, Bae GU, Kim SN, Kim YK. Protein arginine methyltransferases: promising targets for cancer therapy. *Exp Mol Med*. 2021;53:788–808. [DOI] [PubMed] [PMC]
154. Santos M, Hwang JW, Bedford MT. CARM<sub>1</sub> arginine methyltransferase as a therapeutic target for cancer. *J Biol Chem*. 2023;299:105124. [DOI] [PubMed] [PMC]
155. Ma Z, Lyu X, Qin N, Liu H, Zhang M, Lai Y, et al. Coactivator-associated arginine methyltransferase 1: A versatile player in cell differentiation and development. *Genes Dis*. 2022;10:2383–92. [DOI] [PubMed] [PMC]
156. Suresh S, Huard S, Dubois T. CARM1/PRMT4: Making Its Mark beyond Its Function as a Transcriptional Coactivator. *Trends Cell Biol*. 2021;31:402–17. [DOI] [PubMed]
157. Petta S, Targher G, Romeo S, Pajvani UB, Zheng MH, Aghemo A, et al. The first MASH drug therapy on the horizon: Current perspectives of resmetirom. *Liver Int*. 2024;44:1526–36. [DOI] [PubMed]
158. Kokkorakis M, Boutari C, Hill MA, Kotsis V, Loomba R, Sanyal AJ, et al. Resmetirom, the first approved drug for the management of metabolic dysfunction-associated steatohepatitis: Trials, opportunities, and challenges. *Metabolism*. 2024;154:155835. [DOI] [PubMed]

159. Mouskeftara T, Kalopitas G, Liapikos T, Arvanitakis K, Theocharidou E, Germanidis G, et al. A Comprehensive Analysis of Liver Lipidomics Signature in Adults with Metabolic Dysfunction-Associated Steatohepatitis-A Pilot Study. *Int J Mol Sci.* 2024;25:13067. [DOI] [PubMed] [PMC]
160. Berry KA, Hankin JA, Barkley RM, Spraggins JM, Caprioli RM, Murphy RC. MALDI Imaging of Lipid Biochemistry in Tissues by Mass Spectrometry. *Chem Rev.* 2011;111:6491–512. [DOI] [PubMed] [PMC]
161. Flam E, Haas JT, Staels B. Liver metabolism in human MASLD: A review of recent advancements using human tissue metabolomics. *Atherosclerosis.* 2025;400:119054. [DOI] [PubMed]
162. Mazi TA, Borkowski K, Newman JW, Fiehn O, Bowlus CL, Sarkar S, et al. Ethnicity-specific alterations of plasma and hepatic lipidomic profiles are related to high NAFLD rate and severity in Hispanic Americans, a pilot study. *Free Radic Biol Med.* 2021;172:490–502. [DOI] [PubMed] [PMC]
163. Reiniers MJ, van Golen RF, van Gulik TM, Heger M. Reactive oxygen and nitrogen species in steatotic hepatocytes: a molecular perspective on the pathophysiology of ischemia-reperfusion injury in the fatty liver. *Antioxid Redox Signal.* 2014;21:1119–42. [DOI] [PubMed] [PMC]
164. Metz TO, Zhang Q, Page JS, Shen Y, Callister SJ, Jacobs JM, et al. Future of Liquid Chromatography-Mass Spectrometry in Metabolic Profiling and Metabolomic Studies for Biomarker Discovery. *Biomark Med.* 2007;1:159–85. [DOI] [PubMed] [PMC]
165. Mika A, Sledzinski T, Stepnowski P. Current Progress of Lipid Analysis in Metabolic Diseases by Mass Spectrometry Methods. *Curr Med Chem.* 2019;26:60–103. [DOI] [PubMed]
166. Jurowski K, Kochan K, Walczak J, Barańska M, Piekoszewski W, Buszewski B. Analytical Techniques in Lipidomics: State of the Art. *Crit Rev Anal Chem.* 2017;47:418–37. [DOI] [PubMed]
167. Muralidharan S, Lee JWJ, Lim YS, Muthiah M, Tan E, Demicioglu D, et al. Serum lipidomic signatures in patients with varying histological severity of metabolic-dysfunction associated steatotic liver disease. *Metabolism.* 2025;162:156063. [DOI] [PubMed]
168. Eeda V, Patil NY, Joshi AD, Awasthi V. Advancements in metabolic-associated steatotic liver disease research: Diagnostics, small molecule developments, and future directions. *Hepatol Res.* 2024;54:222–34. [DOI] [PubMed]
169. Stadler JT, Marsche G. Obesity-Related Changes in High-Density Lipoprotein Metabolism and Function. *Int J Mol Sci.* 2020;21:8985. [DOI] [PubMed] [PMC]
170. Ipsen DH, Lykkesfeldt J, Tveden-Nyborg P. Molecular mechanisms of hepatic lipid accumulation in non-alcoholic fatty liver disease. *Cell Mol Life Sci.* 2018;75:3313–27. [DOI] [PubMed] [PMC]
171. Jialal I, Singh G. Management of diabetic dyslipidemia: An update. *World J Diabetes.* 2019;10:280–90. [DOI] [PubMed] [PMC]
172. Targher G, Corey KE, Byrne CD, Roden M. The complex link between NAFLD and type 2 diabetes mellitus - mechanisms and treatments. *Nat Rev Gastroenterol Hepatol.* 2021;18:599–612. [DOI] [PubMed]
173. Pouwels S, Sakran N, Graham Y, Leal A, Pintar T, Yang W, et al. Non-alcoholic fatty liver disease (NAFLD): a review of pathophysiology, clinical management and effects of weight loss. *BMC Endocr Disord.* 2022;22:63. [DOI] [PubMed] [PMC]
174. Tully DC, Rucker PV, Chianelli D, Williams J, Vidal A, Alper PB, et al. Discovery of Tropifexor (LJN452), a Highly Potent Non-bile Acid FXR Agonist for the Treatment of Cholestatic Liver Diseases and Nonalcoholic Steatohepatitis (NASH). *J Med Chem.* 2017;60:9960–73. [DOI] [PubMed]
175. Ratziu V, Sanyal A, Harrison SA, Wong VW, Francque S, Goodman Z, et al. Cenicriviroc Treatment for Adults With Nonalcoholic Steatohepatitis and Fibrosis: Final Analysis of the Phase 2b CENTAUR Study. *Hepatology.* 2020;72:892–905. [DOI] [PubMed]
176. Trauner M, Gulamhusein A, Hameed B, Caldwell S, Shiffman ML, Landis C, et al. The Nonsteroidal Farnesoid X Receptor Agonist Cilofexor (GS-9674) Improves Markers of Cholestasis and Liver Injury in Patients With Primary Sclerosing Cholangitis. *Hepatology.* 2019;70:788–801. [DOI] [PubMed] [PMC]

177. Lu H, Ban Z, Xiao K, Sun M, Liu Y, Chen F, et al. Hepatic-Accumulated Obeticholic Acid and Atorvastatin Self-Assembled Nanocrystals Potentiate Ameliorative Effects in Treatment of Metabolic-Associated Fatty Liver Disease. *Adv Sci (Weinh)*. 2024;11:e2308866. [DOI] [PubMed] [PMC]
178. Mo C, Xu X, Zhang P, Peng Y, Zhao X, Chen S, et al. Discovery of HPG1860, a Structurally Novel Nonbile Acid FXR Agonist Currently in Clinical Development for the Treatment of Nonalcoholic Steatohepatitis. *J Med Chem*. 2023;66:9363–75. [DOI] [PubMed]
179. Xie Z, Li Y, Cheng L, Huang Y, Rao W, Shi H, et al. Potential therapeutic strategies for MASH: from preclinical to clinical development. *Life Metab*. 2024;3:loae029. [DOI] [PubMed] [PMC]
180. Polyzos SA, Kountouras J, Mantzoros CS. Obeticholic acid for the treatment of nonalcoholic steatohepatitis: Expectations and concerns. *Metabolism*. 2020;104:154144. [DOI] [PubMed]
181. Ratziu V, Harrison SA, Loustaud-Ratti V, Bureau C, Lawitz E, Abdelmalek M, et al. Hepatic and renal improvements with FXR agonist vonafexor in individuals with suspected fibrotic NASH. *J Hepatol*. 2023;78:479–92. [DOI] [PubMed]
182. Ratziu V, Rinella ME, Neuschwander-Tetri BA, Lawitz E, Denham D, Kayali Z, et al. EDP-305 in patients with NASH: A phase II double-blind placebo-controlled dose-ranging study. *J Hepatol*. 2022;76:506–17. [DOI] [PubMed]
183. Cao S, Yang X, Zhang Z, Wu J, Chi B, Chen H, et al. Discovery of a tricyclic farnesoid X receptor agonist HEC96719, a clinical candidate for treatment of non-alcoholic steatohepatitis. *Eur J Med Chem*. 2022;230:114089. [DOI] [PubMed]
184. Traussnigg S, Halilbasic E, Hofer H, Munda P, Stojakovic T, Fauler G, et al. Open-label phase II study evaluating safety and efficacy of the non-steroidal farnesoid X receptor agonist PX-104 in non-alcoholic fatty liver disease. *Wien Klin Wochenschr*. 2021;133:441–51. [DOI] [PubMed] [PMC]
185. Masoodi M, Gastaldelli A, Hyötyläinen T, Arretxe E, Alonso C, Gaggini M, et al. Metabolomics and lipidomics in NAFLD: biomarkers and non-invasive diagnostic tests. *Nat Rev Gastroenterol Hepatol*. 2021;18:835–56. [DOI] [PubMed]
186. Kartsoli S, Kostara CE, Tsimihodimos V, Bairaktari ET, Christodoulou DK. Lipidomics in non-alcoholic fatty liver disease. *World J Hepatol*. 2020;12:436–50. [DOI] [PubMed] [PMC]
187. Garcia-Jaramillo M, Spooner MH, Löhr CV, Wong CP, Zhang W, Jump DB. Lipidomic and transcriptomic analysis of western diet-induced nonalcoholic steatohepatitis (NASH) in female *Ldlr*<sup>-/-</sup> mice. *PLoS One*. 2019;14:e0214387. [DOI] [PubMed] [PMC]
188. Wang T, Ma G, Nie S, Williamson NA, Reid GE, Gasser RB. Lipid composition and abundance in the reproductive and alimentary tracts of female *Haemonchus contortus*. *Parasit Vectors*. 2020;13:338. [DOI] [PubMed] [PMC]
189. Piras C, Noto A, Ibba L, Deidda M, Fanos V, Muntoni S, et al. Contribution of Metabolomics to the Understanding of NAFLD and NASH Syndromes: A Systematic Review. *Metabolites*. 2021;11:694. [DOI] [PubMed] [PMC]
190. Tan EY, Muthiah MD, Sanyal AJ. Metabolomics at the cutting edge of risk prediction of MASLD. *Cell Rep Med*. 2024;5:101853. [DOI] [PubMed] [PMC]
191. Kalopitas G, Mouskeftara T, Liapikos T, Arvanitakis K, Ioannidis A, Malandris K, et al. Plasma Lipids Profile in the Prediction of Non-Alcoholic Steatohepatitis in Adults: A Case-Control Study. *Int J Mol Sci*. 2023;24:12717. [DOI] [PubMed] [PMC]
192. Zhu Q, Li H, Ao Z, Xu H, Luo J, Kaurich C, et al. Lipidomic identification of urinary extracellular vesicles for non-alcoholic steatohepatitis diagnosis. *J Nanobiotechnology*. 2022;20:349. [DOI] [PubMed] [PMC]
193. Gaggini M, Carli F, Rosso C, Buzzigoli E, Marietti M, Della Latta V, et al. Altered amino acid concentrations in NAFLD: Impact of obesity and insulin resistance. *Hepatology*. 2018;67:145–58. [DOI] [PubMed]